

A Review on the Dietary Flavonoid Kaempferol

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Abstract: Epidemiological studies have revealed that a diet rich in plant-derived foods has a protective effect on human health. Identifying bioactive dietary constituents is an active area of scientific investigation that may lead to new drug discovery. Kaempferol (3,5,7-trihydroxy-2-(4-hydroxyphenyl)-4H-1-benzopyran-4-one) is a flavonoid found in many edible plants (e.g. tea, broccoli, cabbage, kale, beans, endive, leek, tomato, strawberries and grapes) and in plants or botanical products commonly used in traditional medicine (e.g. *Ginkgo biloba*, *Tilia spp*, *Equisetum spp*, *Moringa oleifera*, *Sophora japonica* and propolis). Some epidemiological studies have found a positive association between the consumption of foods containing kaempferol and a reduced risk of developing several disorders such as cancer and cardiovascular diseases. Numerous preclinical studies have shown that kaempferol and some glycosides of kaempferol have a wide range of pharmacological activities, including antioxidant, anti-inflammatory, antimicrobial, anticancer, cardioprotective, neuroprotective, antidiabetic, anti-osteoporotic, estrogenic/antiestrogenic, anxiolytic, analgesic and anti-allergic activities. In this article, the distribution of kaempferol in the plant kingdom and its pharmacological properties are reviewed. The pharmacokinetics (e.g. oral bioavailability, metabolism, plasma levels) and safety of kaempferol are also analyzed. This information may help understand the health benefits of kaempferol-containing plants and may contribute to develop this flavonoid as a possible agent for the prevention and treatment of some diseases.

Keywords: Flavonoids, antioxidant, anti-inflammatory, cancer, kaempferol, astragalin, tiliroside, kaempferitrin, robinin.

1. INTRODUCTION

Flavonoids are a group of plant secondary metabolites characterized by a diphenylpropane structure. They are widely distributed in the plant kingdom and are common constituents of fruits, vegetables and some beverages. Flavonoids may play a role in the decreased risk of chronic diseases associated with a diet rich in plant-derived foods. A positive relationship between the ingestion of foods containing flavonoids and a reduced risk of developing cancer and cardiovascular diseases has indeed been observed in some epidemiological studies [1-5]. *In vitro* and *in vivo* investigations have shown plausible mechanisms by which flavonoids may confer cancer and cardiovascular protection [6]. Evidence also suggests that certain flavonoids may be useful in the treatment of several diseases [7-12]. Some of this evidence comes from the study of plants used in traditional medicine to treat a wide range of pathologies, which has revealed that flavonoids are common bioactive constituents of these plants [10].

The flavonoid kaempferol (3,5,7-trihydroxy-2-(4-hydroxyphenyl)-4H-1-benzopyran-4-one) is a yellow compound with a low molecular weight (MW: 286.2 g/mol) that is commonly found in plant-derived foods and in plants used in traditional medicine. Although there are over two thousand articles in PubMed reporting the isolation and/or biological properties of this flavonoid, there is not any report summarizing or analyzing all this information. The purpose

of this article is to review the distribution and biological activities of kaempferol. First we show a compilation of plant species and foods where kaempferol and/or glycosides of kaempferol have been identified. Then, after reviewing the epidemiological evidence linking the consumption of kaempferol-containing foods to the incidence of several diseases, we provide an overview of the pharmacological and toxicological properties of this polyphenol. Finally, we analyze the bioavailability and metabolism of kaempferol, which may help identify which of the many biological activities of this flavonoid may be relevant in an *in vivo* situation. This information may help understand the preventive and therapeutic properties of kaempferol-containing plants and may help develop this flavonoid as a possible agent for the prevention and treatment of some diseases.

2. DISTRIBUTION IN THE PLANT KINGDOM AND DIETARY SOURCES

Like other flavonoids, kaempferol has a diphenylpropane structure (C6-C3-C6) and is synthesized by condensation of 4-coumaroyl-CoA (C6-C3) with three molecules of malonyl-CoA (C6) [13, 14]. This reaction, catalyzed by the enzyme chalcone synthase (EC 2.3.1.74), results in the formation of the flavonoid naringenin chalcone (C6-C3-C6). This chalcone is transformed into the flavanone naringenin by the enzyme chalcone isomerase (EC 5.5.1.6), which catalyzes the closure of the C3 ring. The enzyme flavanone 3-dioxygenase (EC 1.14.11.9) introduces a hydroxyl group in naringenin at C3 to form dihydrokaempferol. Finally, the enzyme flavonol synthase (EC 1.14.11.23) introduces a double bond in dihydrokaempferol at C2-C3 to produce kaempferol (Fig. 1). Because the enzymes involved in the

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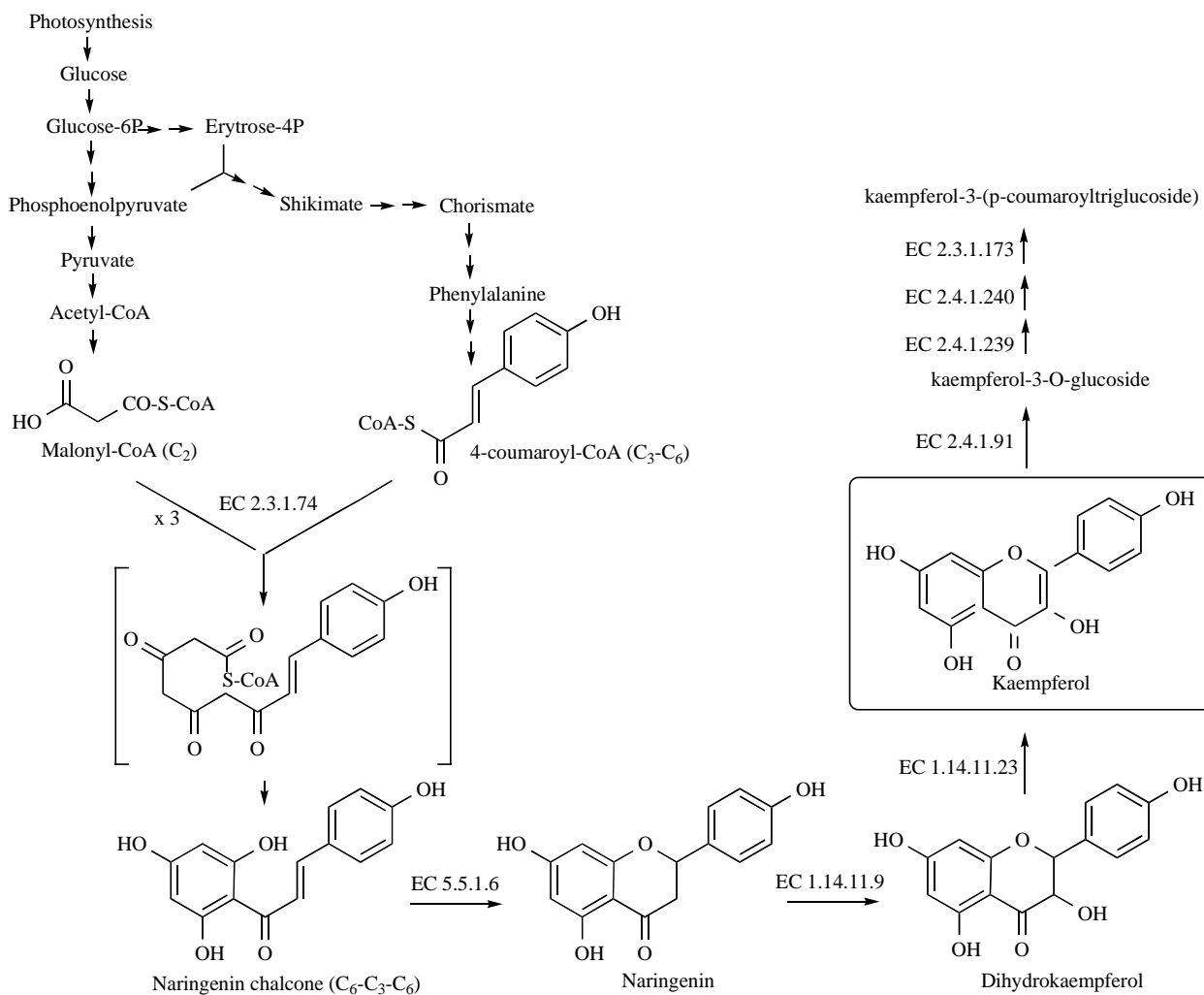


Fig. (1). Biosynthesis of kaempferol and some glycosides of kaempferol (see text for further details). EC 2.3.1.74: chalcone synthase; EC 5.5.1.6: chalcone isomerase; EC 1.14.11.9: flavanone 3-dioxygenase; EC 1.14.11.23: flavonol synthase; EC 2.4.1.91: flavonol 3-*O*-glucosyltransferase; EC 2.4.1.239: flavonol-3-*O*-glucoside glucosyltransferase; EC 2.4.1.240: flavonol-3-*O*-diglycoside glucosyltransferase; EC 2.3.1.173: flavonol-3-*O*-triglycoside *p*-coumaroyltransferase.

biosynthesis of kaempferol are relatively common in the plant kingdom, it is not surprising that this flavonoid is widely distributed in plants. Sugars such as glucose, rhamnose, galactose and rutinose are usually bound to kaempferol to form glycosides. Some glycosides of kaempferol are very common in nature (e.g. kaempferol-3-*O*-glucoside, also called astragalol), because their biosynthesis only requires additional enzymes that are widespread in the plant kingdom (e.g. flavonol 3-*O*-glucosyltransferase, EC 2.4.1.91). The enzymes involved in the biosynthesis of some other kaempferol glycosides are more restricted in nature and, therefore, these glycosides will only be synthesized by plant species with the genetic information required to code for such enzymes. For instance, kaempferol-3-(*p*-coumaroyltriglycoside) is not widely distributed in plants, as its biosynthesis requires the presence of three additional enzymes that are not widespread: flavonol-3-*O*-glucoside glucosyltransferase (EC 2.4.1.239), flavonol-3-*O*-diglycoside glucosyltransferase (EC 2.4.1.240) and flavonol-3-*O*-triglycoside *p*-coumaroyltransferase (EC 2.3.1.173). Table 1 compiles plant species in which

kaempferol and/or kaempferol glycosides have been identified. This table contains over 400 plant species and shows the botanical family, the type of glycoside and the bibliographical reference. Fig. (2) shows the structure of kaempferol and selected glycosides of kaempferol.

Kaempferol has been identified in many botanical families and has been found in Pteridophyta, Pinophyta and Magnoliophyta. In Pteridophyta, kaempferol and/or some of its glycosides have been identified in Aspidiaceae, Aspleniaceae, Blechnaceae, Cyatheaceae, Dennstaedtiaceae, Equisetaceae, Ophioglossaceae, Polypodiaceae and Schizaeaceae (see Table 1 for references). In Pinophyta (Gymnosperms) kaempferol and its glycosides have been identified in Cephalotaxaceae [15], Ginkgoaceae [16-21] and Taxaceae [22]. In the division Magnoliophyta (Angiosperms), kaempferol has been found both in Magnoliopsida (Dicotyledons) and Liliopsida (Monocotyledons). In Monocotyledons, kaempferol has been identified in Alliaceae, Araceae, Asphodelaceae, Dioscoreaceae, Hemerocallidaceae, Hostaceae, Iridaceae,

Table 1. Plant Species that Contain Kaempferol and/or Glycosides of Kaempferol

Species	Family	Compounds	Reference
<i>Abutilon grandiflorum</i>	Malvaceae	Kaempferol 3-O- β -(6"-p-coumaroyl)-glucopyranoside, kaempferol 3-O- β -glucopyranoside and kaempferol 3-O- β -rutinoside	[353]
<i>Abutilon theophrasti</i>	Malvaceae	Kaempferol 3-O- β -(6"-p-coumaroyl)-glucopyranoside, kaempferol 3-O- β -glucopyranoside, kaempferol 3-O- α -rhamnopyranosyl-(1 \rightarrow 6)- β -glucopyranoside and kaempferol 7-O- β -diglucoside	[354]
<i>Acacia nilotica</i>	Leguminosae	Kaempferol	[355]
<i>Acaena splendens</i>	Rosaceae	7-O-acetyl-3-O- β -D-glucosyl-kaempferol	[356]
<i>Acalypha hispida</i>	Euphorbiaceae	Kaempferol 3-O-rutinoside	[357]
<i>Acanthopanax sieboldianus</i>	Araliaceae	Kaempferol 3-O-rutinoside	[358]
<i>Aceriphyllum rossii</i>	Saxifragaceae	Kaempferol 3-O- β -D-glucopyranoside (astragalin) and kaempferol 3-O- α -L-rhamnopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside	[359]
<i>Aconitum</i> spp	Ranunculaceae	Kaempferol 3-O-(6-trans-caffeoyl)- β -glucopyranosyl-(1 \rightarrow 2)- β -glucopyranoside-7-O- α -rhamnopyranoside, kaempferol 3-O-(6-trans-p-coumaroyl)- β -glucopyranosyl-(1 \rightarrow 2)- β -glucopyranoside-7-O- α -rhamnopyranoside, kaempferol 7-O-(6-trans-caffeoyl)- β -glucopyranosyl-(1 \rightarrow 3)- α -rhamnopyranoside-3-O- β -glucopyranoside, kaempferol 7-O-(6-trans-p-coumaroyl)- β -glucopyranosyl-(1 \rightarrow 3)- α -rhamnopyranoside-3-O- β -glucopyranoside, kaempferol 3-O- β -(2"-acetyl)galactopyranoside, kaempferol 3-O- β -(2"-acetyl)galactopyranoside-7-O- α -arabinopyranoside and kaempferol 3,7-di-O- α -rhamnopyranoside	[360]
<i>Aconitum napellus</i>	Ranunculaceae	Kaempferol 7-O-(6-trans-caffeoyl)- β -glucopyranosyl-(1 \rightarrow 3)- α -rhamnopyranoside-3-O- β -glucopyranoside and kaempferol 7-O-(6-trans-p-coumaroyl)- β -glucopyranosyl-(1 \rightarrow 3)- α -rhamnopyranoside-3-O- β -glucopyranoside	[361]
<i>Aconitum naviculare</i>	Ranunculaceae	3-O-[β -D-glucopyranosyl-(1 \rightarrow 3)-(4-O-trans-p-coumaroyl)- α -L-rhamnopyranosyl-(1 \rightarrow 6)- β -D-glucopyranosyl]-7-O-[β -D-glucopyranosyl-(1 \rightarrow 3)- α -L-rhamnopyranosyl]kaempferol	[362]
<i>Aconitum paniculatum</i>	Ranunculaceae	Kaempferol 3-O- β -(2' '-acetyl)galactopyranoside and kaempferol 3-O- β -(2' '-acetyl)galactopyranoside-7-O- α -arabinopyranoside	[363]
<i>Aconitum variegatum</i>	Ranunculaceae	Kaempferol 3-O- β -D-galactopyranoside-7-O- α -L-arabinopyranoside and kaempferol 3-O- β -D-glucopyranoside	[364]
<i>Actinidia valvata</i>	Actinidiaceae	Kaempferol, kaempferol 3-O- α -L-rhamnopyranosyl-(1 \rightarrow 3)-(2,4-di-O-acetyl- α -L-rhamnopyranosyl) (1 \rightarrow 6) β -D-galactopyranoside, kaempferol 3-O- α -L-rhamnopyranosyl-(1 \rightarrow 3)-(4-O-acetyl- α -L-rhamnopyranosyl)-(1 \rightarrow 6)- β -D-galactopyranoside and kaempferol 3-O- β -D-galactopyranoside	[365]
<i>Adina racemosa</i>	Rubiaceae	Kaempferol 3-O- α -L-rhamnopyranosyl-(1 \rightarrow 6)-[(4-O-trans-p-coumaroyl)- α -L-rhamnopyranosyl (1 \rightarrow 2)]-(4-O-trans-p-coumaroyl)- β -D-galactopyranoside	[366]
<i>Ailanthus excelsa</i>	Simaroubaceae	Kaempferol 3-O- α -arabinopyranoside and kaempferol 3-O- β -galactopyranoside	[234]
<i>Alangium salviifolium</i>	Alangiaceae	Kaempferol and kaempferol 3-O- β -D-glycopyranoside	[367]
<i>Albizia lebbek</i>	Leguminosae	Tri-O-glycoside kaempferol	[368]
<i>Allium cepa</i>	Alliaceae	Kaempferol	[369]

(Table 1). Contd.....

Species	Family	Compounds	Reference
<i>Allium hirtifolium</i>	Alliaceae	Kaempferol 3-O- β -D-rhamnopyranosyl-(1 \rightarrow 2)-glucopyranoside, kaempferol 3-O- β -D-glucopyranosyl-(1 \rightarrow 4)-glucopyranoside, kaempferol 3-O-glucopyranoside and kaempferol 7-O-glucopyranoside	[370]
<i>Allium neapolitanum</i>	Alliaceae	Kaempferol 3-O-[(2-O- α -L-rhamnopyranosyl-4-O- β -D-glucopyranosyl)- β -D-glucopyranoside]	[371]
<i>Allium porrum</i>	Alliaceae	Kaempferol 3-O-[2-O-(trans-3-methoxy-4-hydroxycinnamoyl)- β -D-galactopyranosyl]-(1 \rightarrow 4)-O- β -D-glucopyranoside, kaempferol 3-O-[2-O-(trans-3-methoxy-4-hydroxycinnamoyl)- β -D-glucopyranosyl]-(1 \rightarrow 6)-O- β -D-glucopyranoside and other kaempferol glycosides	[372]
<i>Allium triquetrum</i>	Alliaceae	Kaempferol glycosides	[373]
<i>Allium ursinum</i>	Alliaceae	Kaempferol 3-O- β -neohesperidoside-7-O-[2-O-(trans-p-coumaroyl)- β -D-glucopyranoside, kaempferol 3-O- β -neohesperidoside-7-O-[2-O-(trans-feruloyl)- β -D-glucopyranoside, kaempferol 3-O- β -neohesperidoside-7-O-[2-O-(trans-p-coumaroyl)-3-O- β -D-glucopyranosyl]- β -D-glucopyranoside, kaempferol 3-O- β -glucopyranoside and kaempferol 3-O- β -neohesperidoside	[374]
<i>Allium victorialis</i>	Alliaceae	Kaempferol 3, 4'-di-O- β -D-glucoside	[375]
<i>Aloe vera</i>	Asphodelaceae	Kaempferol	[376]
<i>Alomia myriadenia</i>	Asteraceae	Kaempferol 7-methylether (rhamnocitrin)	[377]
<i>Alternanthera tenella</i>	Amaranthaceae	Kaempferol	[378]
<i>Althaea rosea</i>	Malvaceae	Kaempferol	[379]
<i>Amaranthus spinosus</i>	Amaranthaceae	Kaempferol glycosides	[380]
<i>Amburana cearensis</i>	Leguminosae	Kaempferol	[381]
<i>Ammi majus</i>	Apiaceae	Kaempferol 3-O-glucoside and kaempferol 3-O-[2''-(4''-acetyl-rhamnosyl)-(1 \rightarrow 2)-6''-glucosyl] glucoside	[382]
<i>Amoora cucullata</i>	Meliaceae	Kaempferol 3-O- β -D-glucopyranoside	[383]
<i>Anaphalis aureopunctata</i>	Asteraceae	Kaempferol 3-O-acetyl-6-O-(p-coumaroyl)- β -D-glucopyranoside	[384]
<i>Annona purpurea</i>	Annonaceae	Kaempferol 3-O-rhamnoside	[385]
<i>Apocynum venetum</i>	Apocynaceae	Kaempferol and kaempferol 6'-O-acetate	[290, 386]
<i>Arabidopsis mutants</i>	Brassicaceae	Kaempferol glycosides	[387]
<i>Arabidopsis thaliana</i>	Brassicaceae	Kaempferol 3-O- β -[β -D-glucopyranosyl(1 \rightarrow 6)D-glucopyranoside]-7-O- α -L-rhamnopyranoside, kaempferol 3-O- β -D-glucopyranoside-7-O- α -L-rhamnopyranoside and kaempferol 3-O- α -L-rhamnopyranoside-7-O- α -L-rhamnopyranoside	[388]
<i>Ardisia colorata</i>	Myrsinaceae	Kaempferol	[389]
<i>Ardisia japonica</i>	Epacridaceae	Kaempferol 3-O- α -L-rhamnopyranoside and kaempferol 3,7-O- α -L-di-rhamnopyranoside	[390]
<i>Argyrea speciosa</i>	Convolvulaceae	Kaempferol 7-O methyl 3-sulphate	[209]
<i>Asclepias incarnata</i>	Asclepiadaceae	Kaempferol 3- β -glucopyranoside	[391]
<i>Asclepias syriaca</i>	Asclepiadaceae	Kaempferol, kaempferol 3-O- β -galactopyranoside, kaempferol 3-O- β -xylopyranosyl-(1 \rightarrow 2)- β -galactopyranoside, kaempferol 3-O- β -glucopyranosyl-(1 \rightarrow 2)- β -galactopyranoside and kaempferol 3-O- α -rhamnopyranosyl-(1 \rightarrow 2)- β -galactopyranoside	[392]

(Table 1). Contd.....

Species	Family	Compounds	Reference
<i>Asplenium prolongatum</i>	Aspleniaceae	Kaempferol 3-O- α -L-rhamnopyranoside-7-O-[6-feruloyl- β -D-glucopyranosyl-(1 \rightarrow 3)- α -L-rhamnopyranoside]	[393]
<i>Asplenium trichomanes</i>	Aspleniaceae	Kaempferol 3-O- α -[2'acetyl]-arabinofuranosyl-7-O- α -L-rhamnopyranoside	[394]
<i>Astragalus caprinus</i>	Leguminosae	Kaempferol 3-O-[[β -D-xylopyranosyl(1 \rightarrow 3)- α -L-rhamnopyranosyl(1 \rightarrow 6)][β -D-apiofuranosyl(1 \rightarrow 2)]]- β -D-galactopyranosyl	[395, 396]
<i>Astragalus shikokianus</i>	Leguminosae	Kaempferol 3-O- α -L-rhamnopyranosyl -(1 \rightarrow 6)-[α -L-rhamnopyranosyl-(1 \rightarrow 2)]- β -D-galactopyranosyl-7-O- α -L-rhamnopyranoside (astrasikokioside I) and kaempferol 3-O- α -L-rhamnopyranosyl-(1 \rightarrow 2)- β -D-galactopyranosyl-7-O- α -L-rhamnopyranoside	[397]
<i>Baseonema acuminatum</i>	Asclepiadaceae	Kaempferol 3-O-(6"-galloyl)- β -D-glucopyranoside	[398]
<i>Bauhinia forficata</i>	Leguminosae	Kaempferol 3,7-O- α -di-rhamnoside (kaempferitrin)	[246]
<i>Bauhinia malabarica</i>	Leguminosae	Kaempferol and 6,8-di-C-methylkaempferol 3-methylether	[399]
<i>Bauhinia megalandra</i>	Leguminosae	Kaempferol 3-O- α -rhamnoside	[400]
<i>Bauhinia microstachya</i>	Leguminosae	Kaempferol 3-O-rhamnosyl	[401]
<i>Bauhinia variegata</i>	Leguminosae	Kaempferol, kaempferol 7,4'-dimethyl ether 3-O- β -D-glucopyranoside and kaempferol 3-O- β -D-glucopyranoside	[99]
<i>Berchemia floribunda</i>	Rhamnaceae	Kaempferol and kaempferol 3-O- α -L-arabinofuranoside	[175]
<i>Blackstonia perfoliata</i>	Gentianaceae	Kaempferol 3-O- α -L-rhamnopyranosyl-(1 \rightarrow 2)- β -D-galactopyranoside and kaempferol 3-O- α -L-rhamnopyranosyl-(1 \rightarrow 2)- β -D-galactopyranoside-7-O- β -D-glucopyranoside	[402, 403]
<i>Blechnum novae-zelandiae</i>	Blechnaceae	Kaempferol 3-O- β -D-glucuronopyranoside	[404]
<i>Brassica</i> spp	Brassicaceae	Kaempferol 3-O- β -D-sophoroside-7-O- β -D-glucoside and kaempferol 3-O- β -D-(2-sinapoylsophoroside)-7-O- β -D-glucoside	[405]
<i>Brassica campestris</i>	Brassicaceae	Kaempferol and kaempferol 3-O-hydroxyferuloylsophoroside-7-O-glucoside	[406]
<i>Brassica juncea</i>	Brassicaceae	Kaempferol 7-O- β -D-glucopyranosyl-(1 \rightarrow 3)-[β -D-glucopyranosyl-(1 \rightarrow 6)]-glucopyranoside	[407]
<i>Brassica oleracea</i>	Brassicaceae	Kaempferol and kaempferol 3-sinapoyl-di-glucoside-7-di-glucoside	[408]
<i>Brassica rapa</i>	Brassicaceae	Kaempferol	[409]
<i>Bunias orientalis</i>	Brassicaceae	Kaempferol and kaempferol glycosides	[410]
<i>Bunium persicum</i>	Apiaceae	Kaempferol	[411]
<i>Bupleurum flavum</i>	Apiaceae	Kaempferol	[412]
<i>Calligonum comosum</i>	Polygonaceae	Kaempferol 3-O-rhamnopyranoside and kaempferol 3-O-glucuronide	[413]
<i>Callistemon lanceolatus</i>	Myrtaceae	Kaempferol 3-O- β -D-galacturonopyranoside	[414]
<i>Calluna vulgaris</i>	Ericaceae	Kaempferol 3-O- β -D-galactoside	[85]
<i>Camellia sinensis</i>	Theaceae	Kaempferol 3-O-(2-O- β -D-galactopyranosyl-6-O- α -L-rhamnopyranosyl)- β -D-glucopyranoside, kaempferol 3-O-(2-O- β -D-xylopyranosyl-6-O- α -L-rhamnopyranosyl)- β -D-glucopyranoside, kaempferol 3-glucosyl-(1 \rightarrow 3)-rhamnosyl(1 \rightarrow 6)galactosides, kaempferol 3-O- β -D-galactopyranosyl-(1 \rightarrow 2)- β -D-glucopyranoside and kaempferol 3-O-[2-coumaroyl-3-O- β -D-glucosyl-3-O- β -D-glucosylrutinoside]	[415-420]

(Table 1). Contd.....

Species	Family	Compounds	Reference
<i>Campanula alliariifolia</i>	Campanulaceae	Kaempferol 3-O-glucoside	[421]
<i>Campanula barbata</i>	Campanulaceae	Kaempferol 3-O-glucoside and kaempferol 3-O-rutinoside	[422]
<i>Camptosorus sibiricus</i>	Aspleniaceae	Kaempferol 3-O-(6-trans-caffeoyl)- β -D-glucopyranosyl-(1 \rightarrow 2)- β -D-glucopyranoside, kaempferol 3-O-(6-trans-caffeoyl)- β -D-glucopyranosyl-(1 \rightarrow 2)- β -D-glucopyranoside-7-O- β -D-glucopyranoside and kaempferol 3-O-(6-trans-p-coumaroyl)- β -D-glucopyranosyl-(1 \rightarrow 2)- β -D-glucopyranoside-7-O- β -D-glucopyranoside	[423]
<i>Canavalia gladiata</i>	Leguminosae	Kaempferol 3-O- β -D-galactopyranosyl-7-O- α -L-rhamnopyranoside	[424]
<i>Cannabis sativa</i>	Cannabaceae	Kaempferol 3-O-sophoroside	[425]
<i>Capparis spinosa</i>	Capparaceae	Kaempferol derivatives	[46]
<i>Carthamus lanatus</i>	Asteraceae	Kaempferol 3-O- β -D-sophoroside	[426]
<i>Carthamus tinctorius</i>	Asteraceae	Kaempferol 7-O- β -D-glucopyranoside	[427]
<i>Cassia alata</i>	Leguminosae	Kaempferol 3-O-gentiobioside	[428, 429]
<i>Cassia angustifolia</i>	Leguminosae	Kaempferol	[430]
<i>Cassia nodosa</i>	Leguminosae	Kaempferol 3-O-rhamnoside	[431]
<i>Cassia siamea</i>	Leguminosae	Kaempferol	[432]
<i>Cassipourea gummiflua</i>	Rhizophoraceae	Kaempferol 3-O- α -L-rhamnopyranoside	[433]
<i>Celastrus hindsii</i>	Celastraceae	Kaempferol 3-rutinoside	[434]
<i>Celastrus tatarinovii</i>	Celastraceae	Heterosides derivatives of kaempferol	[435]
<i>Centaurea hierapolitana</i>	Asteraceae	Kaempferol 3-O-rutinoside	[436]
<i>Centella asiatica</i>	Apiaceae	Kaempferol and kaempferol 3-O- β -D-glucoside	[437, 438]
<i>Cephalocereus senilis</i>	Cactaceae	Kaempferol 7-rhamnoside, kaempferol 3-rhamnosyl-(1 \rightarrow 6)-galactoside-7-rhamnoside and kaempferol 3-O- β -D-glucopyranosyl-(1 \rightarrow 2)-O-[α -L-rhamnopyranosyl-(1 \rightarrow 6)]- β -D-galactopyranoside-7-O- α -L-rhamnopyranoside	[439]
<i>Cephalotaxus koreana</i>	Cephalotaxaceae	Kaempferol 3-O- α -L-rhamnopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside	[15]
<i>Chenopodium murale</i>	Chenopodiaceae	Kaempferol 3-O-[(4- β -D-apiofuranosyl)- α -L-rhamnopyranoside]-7-O- α -L-rhamnopyranoside, kaempferol 3-O-[(4- β -D-xylopyranosyl)- α -L-rhamnopyranoside]-7-O- α -L-rhamnopyranoside and kaempferol 3-O- β -D-glucopyranoside-7-O- α -L-rhamnopyranoside.	[440]
<i>Chenopodium quinoa</i>	Chenopodiaceae	Kaempferol 3-apiofuranosyl-(1 \rightarrow 2)-rhamnopyranosyl-(1 \rightarrow 6)-galactoside and kaempferol 3-apiofuranosyl-(1 \rightarrow 2)-rhamnopyranosyl-(1 \rightarrow 6)-galactoside	[441]
<i>Chionanthus retusus</i>	Oleaceae	Kaempferol	[442]
<i>Chromolaena odorata</i>	Asteraceae	Kaempferol 4'-methyl ether and kaempferol 3-O-rutinoside	[443]
<i>Chusqueira spinosa</i>	Asteraceae	Kaempferol 3-O-glucuronide, kaempferol 3-O-rutinoside and kaempferol 3-O-glucoside	[444]
<i>Cichorium endivia</i>	Asteraceae	Kaempferol 3-O-glucoside, kaempferol 3-O-glucuronide and kaempferol 3-O-(6-O-malonyl)glucoside	[445]

(Table 1). Contd.....

Species	Family	Compounds	Reference
<i>Cinnamomum osmophloeum</i>	Lauraceae	Kaempferol 3-O-β-D-glucopyranosyl-(1→4)-α-L-rhamnopyranosyl-7-O-α-L-rhamnopyranoside, kaempferol 3-O-β-D-apiofuranosyl-(1→2)-α-L-arabinofuranosyl-7-O-α-L-rhamnopyranoside and kaempferol 3-O-β-D-apiofuranosyl-(1→4)-α-L-rhamnopyranosyl-7-O-α-L-rhamnopyranoside	[446]
<i>Cirsium rivulare</i>	Asteraceae	Kaempferol 3-galactoside	[447]
<i>Cissus sicyoides</i>	Vitaceae	Kaempferol 3-rhamnoside	[448]
<i>Cistus ladanifer</i>	Cistaceae	Kaempferol 3,7-di-O-methyl, kaempferol 3-O-(6"-galloyl)-β-D-glucopyranoside and kaempferol 3-O-methyl	[449, 450]
<i>Cistus laurifolius</i>	Cistaceae	Kaempferol 3,7-O-dimethyl and kaempferol 3,4'-O-dimethyl	[451, 452]
<i>Citrus aurantifolia</i>	Rutaceae	Kaempferol 3-O-β-rutinoside, kaempferol 3-O-β-D-glucopyranoside-6"-(3-hydroxy-3-methyl glutarate) and kaempferol 3-O-β-D-glucopyranoside-6"-(3-hydroxy-3-methyl glutarate)-7-O-β-D-glucopyranoside	[453]
<i>Clitoria ternatea</i>	Leguminosae	Kaempferol, kaempferol 3-(2-rhamnosylrutinoside) and kaempferol 3-O-(2"-O-α-rhamnosyl-6"-O-malonyl)-β-glucoside	[454]
<i>Cnidoscopus aconitifolius</i>	Euphorbiaceae	Kaempferol 3-O-glycosides	[455]
<i>Cnidoscopus chayamansa</i>	Euphorbiaceae	Kaempferol 3-O-glycosides	[455]
<i>Colubrina asiatica</i>	Rhamnaceae	Kaempferol 3-O-rutinoside	[456]
<i>Consolida oliveriana</i>	Ranunculaceae	Kaempferol	[229]
<i>Conyza aegyptiaca</i>	Asteraceae	Kaempferol 3-O-β-D-glucopyranoside	[457]
<i>Conyza filaginoides</i>	Asteraceae	Kaempferol 3-O-(6"-O-caffeoyl)-β-D-galactopyranoside	[458]
<i>Cornus kousa</i>	Cornaceae	Kaempferol 3-O-rhamnoside and kaempferol 3-O-glucoside	[68]
<i>Corylus avellana</i>	Corylaceae	Kaempferol 3-rhamnoside	[459]
<i>Crassocephalum crepidioides</i>	Asteraceae	Kaempferol glycosides	[50]
<i>Crescentia alata</i>	Bignoniaceae	Kaempferol and kaempferol 3-O-rutinoside	[460]
<i>Crocus antalyensis</i>	Iridaceae	Kaempferol 3-O-β-D-(2-O-α-rhamnopyranosyl)glucopyranoside, kaempferol 3-O-β-D-(2-O-β-D-glucopyranosyl)glucopyranoside and kaempferol 3,4'-di-O-β-D-glucopyranoside	[461]
<i>Crocus sativus</i>	Iridaceae	Kaempferol and kaempferol 7-O-β-d-glucopyranoside	[462, 463]
<i>Crocus speciosus</i>	Iridaceae	Kaempferol 3-O-α-(2,3-di-O-β-D-glucopyranosyl)rhamnopyranoside and kaempferol 3-O-α-(2-O-β-D-glucopyranosyl)rhamnopyranoside	[461]
<i>Croton cajucara</i>	Euphorbiaceae	Kaempferol 3,4',7-trimethylether and kaempferol 3,7-dimethylether	[464]
<i>Croton gossypifolius</i>	Euphorbiaceae	Kaempferol 3-O-rhamnopyranoside	[465]
<i>Crypteronia paniculata</i>	Crypteroniaceae	Kaempferol 3-O-α-L-rhamnoside	[466]
<i>Cudrania tricuspidata</i>	Moraceae	Kaempferol, 6-p-hydroxybenzyl kaempferol 7-O-β-D-glucopyranoside and kaempferol 7-O-β-D-glucopyranoside	[467, 468]
<i>Cuphea pinetorum</i>	Lythraceae	Kaempferol	[226]
<i>Cuscuta australis</i>	Convolvulaceae	Kaempferol	[469]
<i>Cuscuta chinensis</i>	Convolvulaceae	Kaempferol	[470]
<i>Cussonia racemosa</i>	Araliaceae	Kaempferol rutinoside	[471]
<i>Cyathea phalerata</i>	Cyatheaceae	kaempferol 3-neohesperidoside	[257]

(Table 1). Contd.....

Species	Family	Compounds	Reference
<i>Cynanchum acutum</i>	Asclepiadaceae	Kaempferol 3-O- β -galacturonopyranoside	[252]
<i>Cynanchum chinense</i>	Asclepiadaceae	Kaempferol 3-O- β -D-glucopyranoside-7-O- α -L-rhamnopyranosyl and kaempferol 3-O- α -L-rhamnopyranoside-7-O- α -L-rhamnopyranosyl	[472]
<i>Daphniphyllum calycinum</i>	Daphniphyllaceae	Kaempferol 3-O-neohesperidoside	[473]
<i>Datura suaveolens</i>	Solanaceae	Kaempferol 3-O- α -L-arabinopyranosyl-7-O- β -D-glucopyranoside and kaempferol 3-O- α -L-arabinopyranoside	[474]
<i>Delphinium gracile</i>	Ranunculaceae	Kaempferol 3-O-[[β -D-glucopyranosyl-(1 \rightarrow 3)-4-O-(E-p-coumaroyl)- α -L-rhamnopyranosyl-(1 \rightarrow 6)]] β -D-glucopyranoside-7-O-(4-O-acetyl)- α -L-rhamnopyranoside and kaempferol 3-O-[[β -D-glucopyranosyl-(1 \rightarrow 3)-4-O-(E-p-coumaroyl)- α -D-rhamnopyranosyl (1 \rightarrow 6)]] β -D-glucopyranoside-7-O-(4-O-acetyl)- α -D-rhamnopyranoside	[475]
<i>Dendrophthoe falcata</i>	Loranthaceae	Kaempferol 3-O- α -L-rhamnopyranoside	[476]
<i>Dennstaedtia scabra</i>	Dennstaedtiaceae	Kaempferol	[477]
<i>Derris trifoliata</i>	Leguminosae	Kaempferol 3-O-[(6'''-feruloyl)- β -D-glucopyranosyl-(1 \rightarrow 3)]-[α -L-rhamnopyranosyl-(1 \rightarrow 6)]- β -D-glucopyranoside, kaempferol 3-O- α -L-rhamnopyranosyl-(1 \rightarrow 6)- β -D-glucopyranosyl-(1 \rightarrow 3)- β -D-glucopyranoside, kaempferol 3-O- α -L-rhamnopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside and kaempferol 3-O- α -L-rhamnopyranosyl-(1 \rightarrow 6)- β -D-glucopyranosyl-(1 \rightarrow 2)- β -D-glucopyranoside	[478, 479]
<i>Dianthus barbatus</i>	Caryophyllaceae	Kaempferol 3-O- β -D-sophoroside	[480]
<i>Diodia teres</i>	Rubiaceae	Kaempferol 3-O-rutinoside	[481]
<i>Dioscorea bulbifera</i>	Dioscoreaceae	Kaempferol 3,5-dimethyl ether	[482]
<i>Diospyros crassiflora</i>	Ebenaceae	Kaempferol 3-O- α -L-rhamnopyranosyl-(1 \rightarrow 2)- β -D-glucopyranoside	[483]
<i>Diospyros kaki</i>	Ebenaceae	Kaempferol 3-O-(2''-O-galloyl)-glucoside, kaempferol 3-O- β -D-galactopyranoside and kaempferol 3-O- β -D-glucopyranoside	[235, 484]
<i>Diospyros lotus</i>	Ebenaceae	Kaempferol	[485]
<i>Dipladenia martiana</i>	Apocynaceae	Kaempferol, kaempferol 3-O- β -D-glucopyranoside (astragalin)	[486]
<i>Diplotaxis erucoides</i>	Brassicaceae	Kaempferol and kaempferol glycosides	[410]
<i>Diplotaxis tenuifolia</i>	Brassicaceae	Kaempferol and kaempferol glycosides	[410]
<i>Dorycnium rectum</i>	Leguminosae	Kaempferol-3,7-O- α -di-rhamnopyranoside, kaempferol-3-O- β -glucopyranoside-7- α -rhamnopyranoside	[487]
<i>Draba nemorosa</i>	Brassicaceae	Kaempferol glycosides	[488]
<i>Dracocephalum peregrinum</i>	Lamiaceae	Kaempferol 3-O-glucopyranoside	[489]
<i>Drynaria fortunei</i>	Polypodiaceae	Kaempferol 3-O- β -D-glucopyranoside-7-O- α -L-arabinofuranoside	[490]
<i>Dryopteris crassirhizoma</i>	Aspidiaceae	Kaempferol glycosides	[218]
<i>Dyosma versipellis</i>	Berberidaceae	Kaempferol	[491]
<i>Echinops echinatus</i>	Asteraceae	Kaempferol, kaempferol 4'-methylether, kaempferol 7-methylether and kaempferol 3-O- α -L-rhamnoside	[492]
<i>Echites hirsuta</i>	Apocynaceae	Kaempferol	[493]
<i>Elateriospermum tapos</i>	Euphorbiaceae	Kaempferol	[494]
<i>Ellipeiopsis cherreensis</i>	Annonaceae	Kaempferol 3-O-rutinoside	[495]

(Table 1). Contd.....

Species	Family	Compounds	Reference
<i>Epimedium sagittatum</i>	Berberidaceae	Kaempferol, kaempferol 3-O-(2"-E-p-coumaroyl,4"-Z-p-coumaroyl)- α -L-rhamnopyranoside and kaempferol 3-O-(3"-Z-p-coumaroyl,4"-E-p-coumaroyl)- α -L-rhamnopyranoside	[496, 497]
<i>Equisetum arvense</i>	Equisetaceae	Kaempferol 3-O-glucoside	[498]
<i>Equisetum debile</i>	Equisetaceae	Kaempferol 3-O-sophoroside, kaempferol 3,7-O- β -D-diglucofuranoside and kaempferol 3-O-sophoroside-7-O- β -D-glucofuranoside	[499]
<i>Equisetum myriochaetum</i>	Equisetaceae	Kaempferol glucosides	[248]
<i>Equisetum palustre</i>	Equisetaceae	Kaempferol 3-O-1"- β -D-glucofuranosyl-3-O-1"- β -D-glucofuranoside	[500]
<i>Eriobotrya japonica</i>	Rosaceae	Kaempferol 3-rhamnosides	[501]
<i>Eruca sativa</i>	Brassicaceae	Kaempferol and kaempferol glycosides	[410]
<i>Eucalyptus</i> spp	Myrtaceae	Kaempferol	[502]
<i>Eucalyptus occidentalis</i>	Myrtaceae	6,8-di-C-methylkaempferol 3-methyl ether	[177]
<i>Euonymus alatus</i>	Celastraceae	Kaempferol	[260]
<i>Euphorbia aleppica</i>	Euphorbiaceae	Kaempferol	[503]
<i>Euphorbia pekinensis</i>	Euphorbiaceae	Kaempferol 3-O-(2"-O-galloyl)- β -D-glucoside	[221]
<i>Euphorbia petiolata</i>	Euphorbiaceae	Kaempferol 3-O-(2-O-galloyl)-glucoside, kaempferol 3-O-glucoside and kaempferol 3-O-rhamnoside	[504]
<i>Euscaphis Japonica</i>	Staphyleaceae	Kaempferol and kaempferol 3-O- β -D-glucofuranoside	[505]
<i>Evolvulus alsinoides</i>	Convolvulaceae	Kaempferol 7-O- β -glucofuranoside, kaempferol 4'-O- β -D-glucofuranosyl-(1 \rightarrow 2)- β -D-glucofuranoside, kaempferol 4'-O- α -L-rhamnopyranosyl-(1 \rightarrow 6)- β -D-glucofuranoside and kaempferol 3-O- β -glucofuranoside	[506, 507]
<i>Fagonia arabica</i>	Zygophyllaceae	Kaempferol 7-O-rhamnoside	[508]
<i>Fagonia taekholmiana</i>	Zygophyllaceae	Kaempferol 3-O-glucoside, kaempferol 3,7-di-O-rhamnoside and kaempferol 3-O- β -L-arabinopyranosyl-(1 \rightarrow 4)- α -L-rhamnopyranoside-7-O- α -L-rhamnopyranoside	[509]
<i>Ficaria verna</i>	Ranunculaceae	Kaempferol 3-O- β -D-(6"- α -L-rhamnopyranosyl)-glucofuranoside (nicotiflorin)	[510]
<i>Ficus pandurata</i>	Moraceae	Kaempferol 3-O- β -neohesperidoside	[511]
<i>Foeniculum vulgare</i>	Apiaceae	Kaempferol 3-O-rutinoside and kaempferol 3-O-glucoside	[512]
<i>Forsteronia refracta</i>	Apocynaceae	Kaempferol 3-O-(2",4"-O-di-acetyl- α -L-rhamnopyranoside), kaempferol 3-O-(3",4"-O-diacetyl- α -L-rhamnopyranoside) and kaempferol 3-O-(4"-O-acetyl- α -L-rhamnopyranoside)	[513]
<i>Fragaria ananassa</i>	Rosaceae	Kaempferol 3- β -D-(6-O-trans-p-coumaroyl)glucofuranoside and kaempferol 3- β -D-(6-O-cis-p-coumaroyl)glucofuranoside	[514, 515]
<i>Frankenia laevis</i>	Frankeniaceae	Kaempferol 3,7-di-sodium sulphate	[516]
<i>Galega officinalis</i>	Leguminosae	Kaempferol 3-[2Gal-(4-acetyl-rhamnosyl)robinobioside] and kaempferol 3-(2Gal-rhamnosylrobinobioside)	[517]
<i>Geranium bellum</i>	Geraniaceae	Kaempferol and Kaempferol 3-O- β -D-glucofuranoside	[518]
<i>Geranium carolinianum</i>	Geraniaceae	Kaempferol	[519]
<i>Geranium potentillaefolium</i>	Geraniaceae	Kaempferol	[518]

(Table 1). Contd.....

Species	Family	Compounds	Reference
<i>Geranium pratense</i>	Geraniaceae	Kaempferol 3-O- β -galactopyranoside and kaempferol 3-O- β -glucopyranoside	[83]
<i>Ginkgo biloba</i>	Ginkgoaceae	Kaempferol, kaempferol 3-O- α -[6"-p-coumaroylglucosyl- β -1,4-rhamnopyranoside), kaempferol 3-O-(2"-O- β -D-glucopyranosyl)- α -L-rhamnopyranoside, kaempferol 3-O-glucosyl(1 \rightarrow 2)rhamnoside, kaempferol 3-O-rhamnosyl-(1 \rightarrow 2)-rhamnosyl-(1 \rightarrow 6)-glucoside, kaempferol 3-O- α -L-[6"-p-coumaroyl- β -D-glucopyranosyl-(1 \rightarrow 2)-rhamnopyranoside] and kaempferol 3-O- α -L-[6"-p-coumaroyl- β -D-glucopyranosyl-(1 \rightarrow 2)-rhamnopyranoside]-7-O- β -D-glucopyranoside	[16-21]
<i>Glycine max</i>	Leguminosae	Kaempferol glycosides	[520]
<i>Glycyrrhiza spp</i>	Leguminosae	Kaempferol 3-O-methyl ether	[521]
<i>Gnidia involucrata</i>	Thymelaeaceae	Kaempferol 3-O-glucoside	[522]
<i>Goodyera schlechtendaliana</i>	Orchidaceae	Kaempferol 3-O-rutinoside	[523]
<i>Grindelia robusta</i>	Asteraceae	6-OH-kaempferol-3,6-dimethylether	[524]
<i>Gymnema sylvestre</i>	Asclepiadaceae	Kaempferol 3-O- β -D-glucopyranosyl-(1 \rightarrow 4)- α -L-rhamnopyranosyl-(1 \rightarrow 6)- β -D-galactopyranoside	[525]
<i>Gynostemma cardiospermum</i>	Cucurbitaceae	Kaempferol	[526]
<i>Hedyosmum bonplandianum</i>	Chloranthaceae	Kaempferol 3-O-[α -L-rhamnopyranosyl (1 \rightarrow 6)- β -D-glucopyranoside] and kaempferol 3-O-[β -D-glucopyranoside]	[300]
<i>Hedyotis diffusa</i>	Rubiaceae	Kaempferol 3-O-(2-O- β -D-glucopyranosyl)- β -D-galactopyranoside and kaempferol 3-O-[2-O-(6-O-E-feruloyl)- β -D-glucopyranosyl]- β -D-galactopyranoside	[281]
<i>Helichrysum italicum</i>	Asteraceae	Kaempferol 3-O- β -D-(6"-E-p-coumaroyl)-glucopyranoside (tiliroside)	[527]
<i>Helleborus niger</i>	Ranunculaceae	Kaempferol 3-O- α -L-arabinopyranosyl-(1 \rightarrow 2)- β -D-galactopyranoside-7-O- β -D-glucopyranoside	[528]
<i>Hemerocallis spp</i>	Hemerocallidaceae	Kaempferol 3-O-glycoside	[529]
<i>Heterotheca inuloides</i>	Asteraceae	Kaempferol	[530]
<i>Hippophae rhamnoides</i>	Elaeagnaceae	Kaempferol	[531]
<i>Hosta ventricosa</i>	Hostaceae	Kaempferol glycosides	[532]
<i>Hydrangea macrophylla</i>	Hydrangeaceae	Kaempferol oligoglycosides	[533]
<i>Hypericum brasiliense</i>	Clusiaceae	Kaempferol	[534]
<i>Hypericum perforatum</i>	Clusiaceae	Kaempferol	[535]
<i>Ilex pernyi</i>	Aquifoliaceae	Kaempferol 3-O-sambubioside	[536]
<i>Impatiens balsamina</i>	Balsaminaceae	Kaempferol, kaempferol 3-glucoside, kaempferol 3-glucosylrhamnoside, kaempferol 3-rutinoside and kaempferol 3-(p-coumaroyl)glucoside	[210, 537, 538]
<i>Impatiens textori</i>	Balsaminaceae	Kaempferol, kaempferol 3-glucoside and kaempferol 3-rhamnosyldiglycoside	[539]
<i>Indigofera hebeptala</i>	Leguminosae	Kaempferol 3-omicron- α -L-rhamnopyranosyl-(1 \rightarrow 2)- β -D-galactopyranoside-7-O- α -L-arabinofuranoside, kaempferol 3-O- α -rhamnopyranosyl-(1 \rightarrow 6)- β -D-galactopyranoside-7-O- α -L-arabinofuranoside, kaempferol 3-omicron- α -L-arabinopyranoside-7-O- α -L-rhamnopyranoside, kaempferol 3-O- α -L-rhamnopyranoside-7-O- α -L-arabinopyranoside and kaempferol 7-rhamnoside	[540]

(Table 1). Contd.....

Species	Family	Compounds	Reference
<i>Indigofera suffruticosa</i>	Leguminosae	Kaempferol	[541]
<i>Indigofera truxillensis</i>	Leguminosae	Kaempferol	[541]
<i>Ixeridium gracile</i>	Asteraceae	Kaempferol	[542]
<i>Kalanchoe pinnata</i>	Crassulaceae	Kaempferol 3-O- α -L-arabinopyranosyl-(1 \rightarrow 2)- α -L-rhamnopyranoside (kapinnatoside)	[230]
<i>Kanahia laniflora</i>	Asclepiadaceae	Kaempferol 3-O-(6-O- α -L-rhamnopyranosyl)- β -D-glucopyranoside and kaempferol 3-O-(2,6-di-O- α -L-rhamnopyranosyl)- β -D-glucopyranoside	[543]
<i>Kitaibelia vitifolia</i>	Malvaceae	Kaempferol 3-O-(6"-p-coumaroyl)- β -glucoside, kaempferol 3-O- β -xylopyranosyl-(1 \rightarrow 2)- β -glucopyranoside and kaempferol 3-O-sambubioside-7-O-glucoside	[544]
<i>Koelreuteria henryi</i>	Sapindaceae	Kaempferol, kaempferol 3-O- α -rhamnopyranoside and kaempferol 3-O- α -arabinopyranoside	[545, 546]
<i>Koelreuteria paniculata</i>	Sapindaceae	Kaempferol 3-O-arabinopyranoside and kaempferol 3-O- α -L-rhamnoside	[547]
<i>Lactuca scariola</i>	Asteraceae	Kaempferol	[548]
<i>Lamium album</i>	Lamiaceae	Kaempferol 3-O-glucoside	[549]
<i>Lamium amplexicaule</i>	Lamiaceae	Kaempferol 3-O-[β -D-glucopyranosyl-(1 \rightarrow 4)] [α -L-rhamnopyranosyl-(1 \rightarrow 6)]- β -D-glucopyranoside	[550]
<i>Laurus nobilis</i>	Lauraceae	Kaempferol 3-O- α -L-(2",4"-di-E-p-coumaroyl)-rhamnoside and kaempferol 3-O- α -L-(2"-Z-p-coumaroyl-4"-E-p-coumaroyl)-rhamnoside	[211]
<i>Lavatera trimestris</i>	Malvaceae	Kaempferol 7-O- β -D-glucoside and kaempferol 3-O- β -D-glucoside	[551]
<i>Leonurus persicus</i>	Lamiaceae	Kaempferol 3-O-glucoside	[552]
<i>Leptothyrsa sprucei</i>	Rutaceae	Kaempferol 3-O- α -L-rhamnopyranoside	[553]
<i>Licania licaniaeflora</i>	Chrysobalanaceae	Kaempferol 3-O- α -rhamnoside	[554]
<i>Ligustrum sinense</i>	Oleaceae	Kaempferol 3-O- α -L-rhamnopyranosyl-7-O- β -D-glucopyranoside	[555]
<i>Lilium candidum</i>	Liliaceae	Kaempferol	[556]
<i>Lilium longiflorum</i>	Liliaceae	Kaempferol and kaempferol glycosides	[557]
<i>Lilium pumilum</i>	Liliaceae	Kaempferol 3-O-rutinoside	[558]
<i>Lonicera japonica</i>	Caprifoliaceae	Kaempferol 3-O- β -D-glucopyranoside	[559]
<i>Loranthus tanakae</i>	Loranthaceae	Kaempferol 3-O- α -L-rhamnoside	[165]
<i>Lotus polyphyllos</i>	Leguminosae	Kaempferol 3,7-di-O-glucosides and kaempferol 3-O- β -(6"-O-E-p-coumaroylglucoside)-7-O- β -glucoside	[560]
<i>Lycium barbarum</i>	Solanaceae	Kaempferol, kaempferol 3-O-rutinoside and kaempferol 3-O-rutinoside-7-O-glucoside	[561, 562]
<i>Lygodium Flexuosum</i>	Schizaeaceae	Kaempferol, kaempferol 3- β -D-glucoside	[563]
<i>Lysimachia vulgaris</i>	Primulaceae	Heterosides derivatives of kaempferol	[564]
<i>Machilus philippinensis</i>	Lauraceae	Kaempferol 3-O- α -L-rhamnopyranoside 3",4"-di-E-P-coumaric acid ester and kaempferol 3-O- α -L-rhamnopyranoside 3"-E,4"-Z-di-p-coumaric acid ester	[565]
<i>Magnolia fargesii</i>	Magnoliaceae	Kaempferol 3-O- β -D-(6"-O-coumaroyl)glucopyranoside (tiliroside)	[566]

(Table 1). Contd.....

Species	Family	Compounds	Reference
<i>Malva crispa</i>	Malvaceae	Kaempferol glycosides	[567]
<i>Malus domestica</i>	Rosaceae	Kaempferol 3-O-rhamnoside	[568]
<i>Manihot esculenta</i>	Euphorbiaceae	Kaempferol 3-O-rutinoside	[569]
<i>Maytenus aquifolium</i>	Celastraceae	Kaempferol 3-O- α -L-rhamnopyranosyl-(1 \rightarrow 6)-O-[β -D-glucopyranosyl-(1 \rightarrow 3)-O- α -L-rhamnopyranosyl-(1 \rightarrow 2)-O- β -D-galactopyranoside	[570]
<i>Maytenus ilicifolia</i>	Celastraceae	Kaempferol 3-O- α -L-rhamnopyranosyl-(1 \rightarrow 6)-O-[α -L-arabinopyranosyl-(1 \rightarrow 3)-O- α -L-rhamnopyranosyl-(1 \rightarrow 2)]-O- β -D-galactopyranoside and other kaempferol glycosides	[571]
<i>Meconopsis</i> spp	Papaveraceae	Kaempferol 3-O-(6-O- β -D-glucopyranosyl)- β -D-glucopyranoside and kaempferol 3-O-(6-O- β -D-glucopyranosyl)- β -D-galactopyranoside	[572]
<i>Meconopsis quintuplinervia</i>	Papaveraceae	Kaempferol 3-O- β -D-glucopyranoside and kaempferol 3-O-[β -D-glucopyranosyl-(1 \rightarrow 2)- β -D-glucopyranoside]	[573]
<i>Melastoma malabathricum</i>	Melastomataceae	Kaempferol 3-O-(2",6"-di-O-p-trans-coumaroyl)- β -glucoside	[574]
<i>Melilotus neapolitana</i>	Leguminosae	Kaempferol 3-O-rutinoside and kaempferol 3-O-glucoside	[575]
<i>Miconia cabucu</i>	Melastomataceae	Kaempferol 3-O- β -D-(6"-coumaroyl)-glucopyranoside	[576]
<i>Mitracarpus scaber</i>	Rubiaceae	Kaempferol 3-O-rutinoside	[577]
<i>Momina sylvatica</i>	Polygalaceae	Kaempferol 3-O- β -D-glucosyl-(1 \rightarrow 2)-O-[α -L-rhamnosyl-(1 \rightarrow 6)]- β -D-galactoside and other kaempferol glycosides	[578]
<i>Morinda citrifolia</i>	Rubiaceae	Kaempferol	[109]
<i>Morinda morindoides</i>	Rubiaceae	Kaempferol and kaempferol 7-O-[α -L-rhamnopyranosyl-(1 \rightarrow 6)]-[β -D-glucopyranosyl-(1 \rightarrow 2)]- β -D-glucopyranoside	[579-581]
<i>Moringa oleifera</i>	Moringaceae	Kaempferol, kaempferol 3-O-[β -glucosyl-(1 \rightarrow 2)]-[α -rhamnosyl-(1 \rightarrow 6)]- β -glucoside-7-O- α -rhamnoside, kaempferol 3-O-[α -rhamnosyl-(1 \rightarrow 2)]-[α -rhamnosyl-(1 \rightarrow 4)]- β -glucoside-7-O- α -rhamnoside and kaempferol 3-O- α -rhamnoside	[582-584]
<i>Morus alba</i>	Moraceae	Kaempferol 3-O- β -D-glucopyranoside (astragalol), kaempferol 3-O-(6"-O-acetyl)- β -D-glucopyranoside and kaempferol 3-O- α -L-rhamnopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside	[585]
<i>Morus insignis</i>	Moraceae	Kaempferol 3-O- β -glucopyranoside	[253]
<i>Musa</i> spp	Musaceae	Kaempferol 3-O-rutinoside	[586]
<i>Nelumbo nucifera</i>	Nelumbonaceae	Kaempferol and kaempferol glycosides	[98, 587]
<i>Neocheiropteris palmatopedata</i>	Polypodiaceae	Kaempferol glycosides	[588]
<i>Nepenthes gracilis</i>	Nepenthaceae	Kaempferol	[589]
<i>Nephelium lappaceum</i>	Sapindaceae	Kaempferol 3-O- β -D-glucopyranoside-7-O- α -L-rhamnopyranoside	[590]
<i>Nicotiana tabacum</i>	Solanaceae	Kaempferol 3-rutinoside	[591]
<i>Nymphaea candida</i>	Nymphaeaceae	Kaempferol, kaempferol 3-O-(2"-O-galloylrutinoside), kaempferol 3-O- β -D-glucopyranoside, kaempferol 3-O- α -L-rhamnopyranoside, kaempferol 3-O- α -L-rhamnopyranosylglucopyranoside and kaempferol 7-O- β -D-glucopyranoside-3-(O- α -L-rhamnopyranosylglucopyranoside)	[282]
<i>Nymphaea odorata</i>	Nymphaeaceae	Kaempferol 3-O- α -L-rhamnopyranoside	[592]
<i>Ochna beddomei</i>	Ochnaceae	Kaempferol, kaempferol 3-O-rhamnoside and kaempferol 3-O-glucoside	[593]

(Table 1). Contd.....

Species	Family	Compounds	Reference
<i>Ochradenus baccatus</i>	Resedaceae	Kaempferol 3-O-β-D-glucopyranoside (astragalín)	[594]
<i>Olea europaea</i>	Oleaceae	Kaempferol	[595]
<i>Ophioglossum petiolatum</i>	Ophioglossaceae	Kaempferol	[596]
<i>Opuntia dillenii</i>	Cactaceae	Kaempferol 7-O-β-D-glucopyranosyl-(1→4)-β-D-glucopyranoside	[597]
<i>Opuntia ficus-indica</i>	Cactaceae	Kaempferol, kaempferol 3-methyl ether	[598]
<i>Origanum dictamnus</i>	Lamiaceae	Kaempferol	[599]
<i>Orostachys japonicus</i>	Crassulaceae	Kaempferol	[600]
<i>Oxytropis falcate</i>	Leguminosae	Kaempferol	[601]
<i>Paeonia suffruticosa</i>	Paeoniaceae	Kaempferol 7-O-glucoside	[602]
<i>Papaver nudicaule</i>	Papaveraceae	Kaempferol 3-O-β-sophoroside, kaempferol 3-O-β-sophoroside-7-O-β-glucoside and other kaempferol derivatives	[603]
<i>Parthenocissus tricuspidata</i>	Vitaceae	Kaempferol and kaempferol 3,5,7,4'-O-tetramethyl	[604]
<i>Pedilanthus tithymalooides</i>	Euphorbiaceae	Kaempferol 3-O-β-D-glucopyranoside-6''-(3-hydroxy-3-methylglutarate)	[605]
<i>Pelargonium quercifolium</i>	Geraniaceae	Kaempferol	[606]
<i>Pemphis acidula</i>	Lythraceae	Kaempferol 6''-O-galloyl-β-D-glycosides	[607]
<i>Pentachondra pumila</i>	Epacridaceae	Kaempferol 3-(2,4-di-E-p-coumaroylrhamnoside)	[608]
<i>Persicaria lapathifolia</i>	Polygonaceae	Kaempferol 3-O-β-D-(6''-p-hydroxybenzoyl)-galactopyranoside	[609]
<i>Peumus boldus</i>	Monimiaceae	Kaempferol glycosides	[610]
<i>Phaleria macrocarpa</i>	Thymelaeaceae	Kaempferol 3-O-β-D-glucoside	[611]
<i>Phaseolus vulgaris</i>	Leguminosae	Kaempferol 3-O-[xylosyl-(1→2)]-rhamnosyl-(1→6)-glucoside, kaempferol 3-O-glucuronide, kaempferol 3-O-β-D-glucopyranoside and kaempferol 3-O-rutinoside	[612, 613]
<i>Phellodendron amurense</i>	Rutaceae	Kaempferol 3-O-β-D-glucoside	[614]
<i>Phlomis aurea</i>	Lamiaceae	Kaempferol 3-O-β-D-glucopyranosyl-(1-6)-β-D-glucopyranoside	[615]
<i>Phlomis caucasica</i>	Lamiaceae	Kaempferol 3-O-glucoside	[616]
<i>Phyllanthus acidus</i>	Euphorbiaceae	Kaempferol	[617]
<i>Phyllanthus emblica</i>	Euphorbiaceae	Kaempferol 3-O-α-L-(6''-methyl)-rhamnopyranoside and kaempferol 3-O-α-L-(6''-ethyl)-rhamnopyranoside	[618]
<i>Phytolacca americana</i>	Phytolaccaceae	Kaempferol 3-O-β-D-glucopyranoside, kaempferol 3-O-β-D-xylopyranosyl-(1→2)-β-D-glucopyranoside, kaempferol 3-O-α-L-rhamnopyranosyl-(1→2)-β-D-glucopyranoside and kaempferol 3-O-di-glucoside	[619]
<i>Pinus densiflora</i>	Pinaceae	Kaempferol 3-O-β-D-glucoside and 6-C-methyl kaempferol 3-O-β-D-glucoside	[620]
<i>Pistacia vera</i>	Anacardiaceae	Kaempferol	[621]
<i>Pisum sativum</i>	Leguminosae	Kaempferol 3-sophorotrioside	[622]
<i>Planchonia grandis</i>	Lecythidaceae	Kaempferol	[623]
<i>Platanus occidentalis</i>	Platanaceae	Kaempferol 3-O-α-L-(2'',3''-di-E-p-coumaroyl)rhamnoside, kaempferol 3-O-α-L-(2''-E-p-coumaroyl-3''-Z-p-coumaroyl)rhamnoside, kaempferol 3-O-α-L-(2''-Z-p-coumaroyl-3''-E-p-coumaroyl)rhamnoside and kaempferol 3-O-α-L-(2'',3''-di-Z-p-coumaroyl)rhamnoside	[624]

(Table 1). Contd.....

Species	Family	Compounds	Reference
<i>Platanus orientalis</i>	Platanaceae	Kaempferol 3-O- α -L-(2"-E-p-coumaroyl)-rhamnopyranoside, kaempferol 3-O- β -D-(6"-E-p-coumaroyl)-glucopyranoside and kaempferol 3-O- α -L-(2",3"-di-E-p-coumaroyl)-rhamnopyranoside	[625]
<i>Pleurospermum franchetianum</i>	Apiaceae	Kaempferol 3,7-di-O- α -L-rhamnopyranoside and kaempferol 3-O- β -D-glucopyranosyl-7-O- α -L-rhamnopyranoside	[626]
<i>Pogonatherum crinitum</i>	Poaceae	Kaempferol, kaempferol 3-O- α -L-rhamnopyranoside and kaempferol 3-O-rutinoside	[627]
<i>Polygala japonica</i>	Polygalaceae	Kaempferol 3-gentiobioside and kaempferol glycosides	[628]
<i>Polygonum</i> spp	Polygonaceae	Kaempferol 3-O-glucoside-2"-gallate	[629]
<i>Polygonum amphibium</i>	Polygonaceae	Kaempferol 3-O- α -rhamnosyl-(1 \rightarrow 2)- β -glucuronide	[630]
<i>Polygonum salicifolium</i>	Polygonaceae	Kaempferol 3-O- β -D-glucopyranoside (astragalin) and kaempferol 3-O- β -D-galactopyranoside	[631]
<i>Polygonum tinctorium</i>	Polygonaceae	Kaempferol	[208]
<i>Polypodium decumanum</i>	Polypodiaceae	Kaempferol glycosides	[632]
<i>Polypodium triseriale</i>	Polypodiaceae	Kaempferol glycosides	[632]
<i>Pongamia pinnata</i>	Leguminosae	Kaempferol, kaempferol 3-O- β -D-rutinoside and kaempferol 3-O- β -D-glucopyranoside	[633]
<i>Populus davidiana</i>	Salicaceae	Kaempferol	[634]
<i>Potentilla argentea</i>	Rosaceae	Kaempferol 3-O- β -D-(6"-E-p-coumaroyl)-glucopyranoside	[168]
<i>Primula</i> spp	Primulaceae	Kaempferol 3-O-(2-O- α -L-rhamnopyranosyl-6-O- β -D-xylopyranosyl- β -D-glucopyranoside)	[635]
<i>Prunus amygdalus</i>	Rosaceae	Kaempferol 3-O- α -L-rhamnopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside	[636]
<i>Prunus davidiana</i>	Rosaceae	Kaempferol and kaempferol 7-O- β -D-glucoside	[637]
<i>Prunus serotina</i>	Rosaceae	Kaempferol 3-O- α -L-arabinofuranoside (juglanin)	[638]
<i>Prunus serrulata</i>	Rosaceae	Kaempferol 3-O- α -arabinofuranoside, kaempferol 3-O- β -xylopyranoside, kaempferol 3-O- β -glucopyranoside and kaempferol 3-O- β -xylopyranosyl-(1 \rightarrow 2)- β -glucopyranoside	[639]
<i>Prunus spinosa</i>	Rosaceae	Kaempferol, kaempferol 3-O- α -L-arabinofuranoside, kaempferol 3-O- α -L-rhamnopyranoside, kaempferol 7-O- α -L-rhamnopyranoside, kaempferol 3-O- β -D-xylopyranoside, kaempferol 3,7-di-O- α -L-rhamnopyranoside and kaempferol 3-O-(2"-E-p-coumaroyl)- α -L-arabinofuranoside	[640, 641]
<i>Prunus tomentosa</i>	Rosaceae	Kaempferol, kaempferol 3-O- α -L-rhamnopyranoside, kaempferol 3-O- β -D-(6-acetyl)-glucopyranosyl-(1 \rightarrow 4)- α -L-rhamnopyranoside (multiflorin A) and kaempferol 3-O- β -D-glucopyranosyl-(1 \rightarrow 4)- α -L-rhamnopyranoside (multiflorin B)	[104]
<i>Psidium guajava</i>	Myrtaceae	Kaempferol	[642]
<i>Pteridium aquilinum</i>	Dennstaedtiaceae	Kaempferol 3-O- β -D-glucopyranoside	[643]
<i>Pterogyne nitens</i>	Caesalpiniaceae	Kaempferol and kaempferol 3,7-O- α -di-rhamnoside (kaempferitrin)	[644]
<i>Punica granatum</i>	Punicaceae	Kaempferol	[645]
<i>Pyrenacantha staudtii</i>	Icacinaceae	Kaempferol 3-O- β -rhamnopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside	[646]
<i>Pyrrosia petiolosa</i>	Polypodiaceae	Kaempferol 3-O- β -D-glucopyranoside-7-O- α -L-arabinofuranoside	[647]

(Table 1). Contd.....

Species	Family	Compounds	Reference
<i>Pyrus communis</i>	Rosaceae	Kaempferol 3-O-β-D-glucopyranoside, 8-methoxykaempferol 3-O-β-D-(2"-O-α-D-glucopyranosyl)-glucopyranoside and 8-methoxykaempferol 3-O-β-D-(2"-O-α-L-rhamnopyranosyl)-glucopyranoside	[648]
<i>Quercus dentata</i>	Fagaceae	Kaempferol 3-O-β-D-glucopyranoside, kaempferol 3-O-(6"-trans-p-coumaroyl)-β-D-glucopyranoside, kaempferol 3-O-(2",6"-di-trans-p-coumaroyl)-β-D-glucopyranoside and kaempferol 3-O-(2",4"-di-acetyl-3"-cis-p-coumaroyl-6"-trans-p-coumaroyl)-β-D-glucopyranoside	[649]
<i>Randia formosa</i>	Rubiaceae	Kaempferol 3-O-rutinoside	[650]
<i>Reseda muricata</i>	Resedaceae	Kaempferol 3-O-β-D-glucopyranosyl-(1"→2")-O-α-L-rhamnopyranoside 7-O-β-D-glucopyranoside and kaempferol 3-O-β-D-glucopyranosyl-(1"→2")-O-α-L-rhamnopyranoside 7-O-β-D-(6"-O-E-coumaryl)glucopyranoside)	[651]
<i>Rhamnus nakaharai</i>	Rhamnaceae	Kaempferol	[652]
<i>Rhamnus nipalensis</i>	Rhamnaceae	Kaempferol 4'-methylether	[653]
<i>Rhamnus petiolaris</i>	Rhamnaceae	Kaempferol 3-rhamninoside	[654]
<i>Rhamnus procumbens</i>	Rhamnaceae	Kaempferol	[655]
<i>Rhamnus thymifolius</i>	Rhamnaceae	Kaempferol 3-O-α-L-rhamnopyranosyl-(1→3)-(4-O-acetyl)-O-α-L-rhamnopyranosyl-(1→6)-O-β-D-galactopyranoside and kaempferol 4'-O-α-L-rhamnopyranosyl-(1→3)-O-α-L-rhamnopyranosyl-(1→6)-O-β-D-galactopyranoside	[656]
<i>Rheum nobile</i>	Polygonaceae	Kaempferol glycoside	[657]
<i>Rhodiola rosea</i>	Crassulaceae	Kaempferol	[217]
<i>Rhodiola sachalinensis</i>	Crassulaceae	Kaempferol, kaempferol 3-O-β-D-xylofuranosyl-(1→2)-β-D-glucopyranoside, kaempferol 7-O-α-L-rhamnopyranoside and kaempferol 3-O-β-D-glucopyranosyl-(1→2)-β-D-glucopyranoside	[658-660]
<i>Rhus verniciflua</i>	Anacardiaceae	Kaempferol and kaempferol 3-O-glucoside	[91]
<i>Ribes nigrum</i>	Grossulariaceae	Kaempferol 3-O-(6"-malonyl)glucoside	[661]
<i>Rodgersia podophylla</i>	Saxifragaceae	Kaempferol 3-O-α-L-5"-acetyl-arabinofuranoside and kaempferol 3-O-α-L-3"-acetyl-arabinofuranoside	[662]
<i>Rosa spp</i>	Rosaceae	Kaempferol	[663]
<i>Rosa canina</i>	Rosaceae	Kaempferol 3-O-β-D-(6"-E-p-coumaroyl)-glucopyranoside (tiliroside)	[664]
<i>Rosa damascena</i>	Rosaceae	Kaempferol, kaempferol 3-O-glucoside and other kaempferol glycosides	[219, 665]
<i>Rosa hybrids</i>	Rosaceae	Kaempferol	[438]
<i>Rosmarinus officinalis</i>	Lamiaceae	Kaempferol	[666]
<i>Rubus idaeus</i>	Rosaceae	Kaempferol, kaempferol 3-O-β-D-galactosides, kaempferol 3-O-β-L-arabinopyranoside and kaempferol 3-O-β-D-(6"-E-p-coumaroyl)-glucoside (tiliroside)	[667, 668]
<i>Rubus sanctus</i>	Rosaceae	Kaempferol 3-O-(6"-O-galloyl)-(4)C(1)-β-D-galactopyranoside	[669]
<i>Rubus ulmifolius</i>	Rosaceae	Kaempferol 3-O-β-D-glucuronide	[670, 671]
<i>Rumex chalepensis</i>	Polygonaceae	Kaempferol 3-rhamnosyl-(1→6) galactoside	[672]

(Table 1). Contd.....

Species	Family	Compounds	Reference
<i>Rumex japonicus</i>	Polygonaceae	Kaempferol 3-O- β -D-glucoside	[673]
<i>Sageretia theezans</i>	Rhamnaceae	Kaempferol 3-O- α -L-rhamnopyranoside	[674]
<i>Salix raddeana</i>	Salicaceae	Kaempferol 3-O-glucoside	[675]
<i>Sambucus nigra</i>	Caprifoliaceae	Kaempferol	[676]
<i>Sanguisorba minor</i>	Rosaceae	Kaempferol 3-O-[2"-galloyl-O- β -glucosyl-(1" \rightarrow 2")-O- β -glucoside)] and kaempferol 3-O-mono-glycosides	[677]
<i>Sauropus androgynus</i>	Euphorbiaceae	Kaempferol and 3-O- β -D-glucosyl-(1 \rightarrow 6)- β -D-glucosyl-kaempferol	[23, 244]
<i>Scabiosa hymettia</i>	Dipsacaceae	Kaempferol 3-O-[3-O-acetyl-6-O-(E)-p-coumaroyl]- β -D-glucopyranoside and kaempferol 3-O- β -D-glucopyranoside (astragallicin)	[678]
<i>Scopolia carniolica</i>	Solanaceae	Kaempferol 3-O-(2-glucosyl)-galactoside-7-O-glucoside	[679]
<i>Scopolia caucasica</i>	Solanaceae	Kaempferol 3-O-(2-glucosyl)-galactoside-7-O-glucoside and kaempferol 3-O-(2-glucosyl)-galactoside	[680]
<i>Scopolia lurida</i>	Solanaceae	Kaempferol glycosides	[679]
<i>Scopolia sinensis</i>	Solanaceae	Kaempferol 3-O-(2-glucosyl)-galactoside-7-O-glucoside	[679]
<i>Scrophularia ilwensis</i>	Scrophulariaceae	Kaempferol 3-O-rutinoside	[681]
<i>Sebastiania brasiliensis</i>	Euphorbiaceae	Kaempferol	[682]
<i>Securigera securidaca</i>	Leguminosae	Kaempferol	[683]
<i>Securinea virosa</i>	Euphorbiaceae	Kaempferol 3-O-(4-galloyl)- β -D-glucopyranoside	[684]
<i>Sedum dendroideum</i>	Crassulaceae	Kaempferol 3-O- α -rhamnopyranosyl-(1 \rightarrow 2)- β -glucopyranoside-7-O- α -glucopyranoside, kaempferol 3-O- α -rhamnopyranosyl-(1 \rightarrow 2)- β -glucopyranoside-7-O- α -rhamnopyranoside, kaempferol 3,7-O- α -di-rhamnoside (kaempferitrin) and kaempferol 3-O- β -glucopyranoside-7-O- α -rhamnopyranoside	[302]
<i>Sedum sarmentosum</i>	Crassulaceae	Kaempferol 3- α -arabinopyranoside	[233]
<i>Senecio scandens</i>	Asteraceae	Kaempferol 3-O-rhamnoside	[685]
<i>Silphium perfoliatum</i>	Asteraceae	Kaempferol 3-O- β -D-apiofuranoside 7-O- α -L-rhamnosyl-(1" \rightarrow 6")-O- β -D-galactopyranoside and kaempferol 3-O- β -D-apiofuranoside-7-O- α -L-rhamnosyl-(1" \rightarrow 6")-O- β -D-(2"-O-E-caffeoyl)galactopyranoside)	[686]
<i>Simarouba versicolor</i>	Simaroubaceae	Kaempferol	[687]
<i>Siparuna apiosyce</i>	Monimiaceae	Kaempferol glycosides	[688, 689]
<i>Siraitia grosvenori</i>	Cucurbitaceae	Kaempferol 7-O- α -L-rhamnopyranoside and kaempferol 3,7-di-O- α -L-rhamnopyranoside	[690]
<i>Smilax bockii</i>	Smilacaceae	Kaempferol and kaempferol 7-O- β -D-glucopyranoside	[691]
<i>Smilax china</i>	Smilacaceae	Kaempferol 7-O- β -D-glucoside	[182, 187]
<i>Solanum lycopersicum</i>	Solanaceae	Kaempferol, kaempferol 3-O-sophoroside and kaempferol 3-O-rutinoside	[692-694]
<i>Solanum nigrum</i>	Solanaceae	Kaempferol	[695]
<i>Solenostemma argel</i>	Asclepiadaceae	Kaempferol 3-O- α -rhamnopyranosyl-(1 \rightarrow 2)- β -glucopyranoside	[696]
<i>Solidago altissima</i>	Asteraceae	Kaempferol 3-O- β -D-apiofuranosyl-(1 \rightarrow 6)- β -D-glucopyranoside	[697]
<i>Solidago virga-aurea</i>	Asteraceae	Kaempferol and kaempferol 3-O-rutinoside	[698]

(Table 1). Contd.....

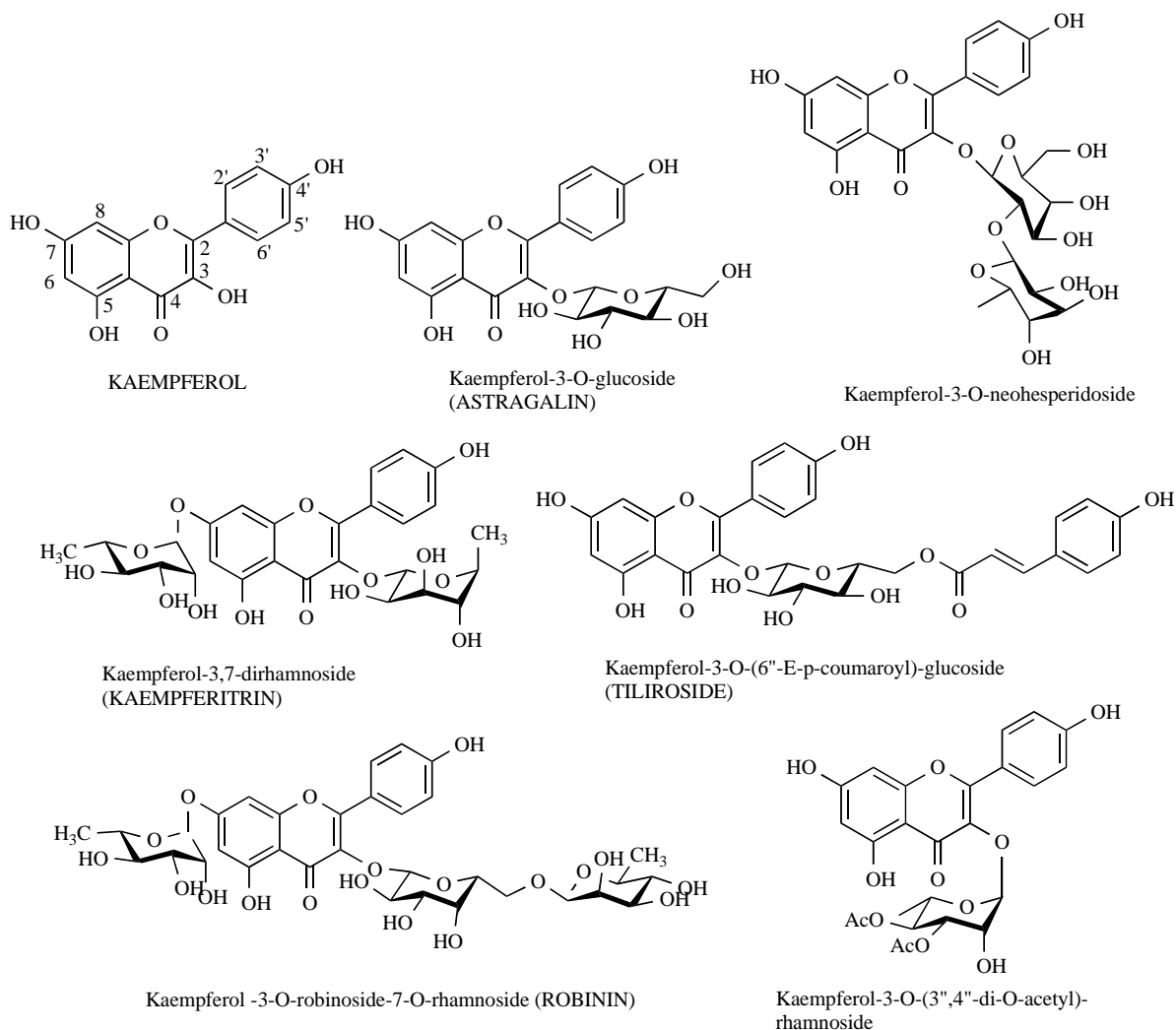
Species	Family	Compounds	Reference
<i>Sophora japonica</i>	Leguminosae	Kaempferol, kaempferol 3-O- α -L-rhamnopyranosyl-(1 \rightarrow 6)- β -D-glucopyranosyl-(1 \rightarrow 2)- β -D-glucopyranoside, kaempferol 3-O-[α -L-rhamnopyranosyl-(1 \rightarrow 6)]-[β -D-glucopyranosyl-(1 \rightarrow 2)]- β -D-glucopyranoside, kaempferol 3-O- β -D-glucopyranosyl-(1 \rightarrow 2)- β -D-glucopyranoside-7-O- α -L-rhamnopyranoside, kaempferol 3-O- β -rutinoside and kaempferol 3-O- α -L-rhamnopyranosyl(1 \rightarrow 6)- β -D-glucopyranosyl(1 \rightarrow 2)- β -D-glucopyranoside-7-O- α -L-rhamnopyranoside	[699-703]
<i>Spiraea canescens</i>	Rosaceae	6'-O-(4"-methoxy-trans-cinnamoyl)-kaempferol-3- β -D-glucopyranoside	[704]
<i>Spiranthes australis</i>	Orchidaceae	Kaempferol 3-O-[[O- β -D-xylopyranosyl]-(1 \rightarrow 2)- β -D-glucopyranosyl]-8-(p-hydroxy-benzyl) and kaempferol 3-O-[O-[2-O-(E)-p-coumaroyl- β -D-xylopyranosyl]-(1 \rightarrow 2)- β -D-glucopyranosyl]-8-(p-hydroxy-benzyl)	[705]
<i>Stenochlaena palustris</i>	Blechnaceae	Kaempferol 3-O-(3"-O-E-p-coumaroyl)-(6"-O-E-feruloyl)- β -D-glucopyranoside, kaempferol 3-O-(3",6"-di-O-E-p-coumaroyl)- β -D-glucopyranoside, kaempferol 3-O-(3"-O-E-p-coumaroyl)- β -D-glucopyranoside, kaempferol 3-O-(6"-O-E-p-coumaroyl)- β -D-glucopyranoside and kaempferol 3-O- β -D-glucopyranoside	[706]
<i>Stocksia brauhica</i>	Sapindaceae	Kaempferol 3-O-[(α -L-rhamnopyranosyl)oxy]-7-O-(acetyl)- β -D-glucopyranosyl-(1 \rightarrow 4)]-[6-O-(4-hydroxy-E-cinnamoyl)- β -D-glucopyranosyl-(1 \rightarrow 2)- α -L-rhamnopyranosyl-oxy], 3-O-[[2-O- β -D-glucopyranosyl]-3-[O- β -D-glucopyranosyl]-4-[(6-O-p-coumaroyl)-O- β -D-glucopyranosyl]]- α -L-rhamnopyranosyl-kaempferol-7-O- α -L-rhamnopyranoside and 3-O-{2-[(6-O-p-coumaroyl)-O- β -D-glucopyranosyl]-3-[O- β -D-glucopyranosyl]-4-[(6-O-p-coumaroyl)-O- β -D-glucopyranosyl]]- α -L-rhamnopyranosyl-kaempferol-7-O- α -L-rhamnopyranoside	[707, 708]
<i>Sutherlandia frutescens</i>	Leguminosae	Kaempferol 3-O- β -D-xylopyranosyl-(1 \rightarrow 2)-[6-O-(3-hydroxy-3-methylglutaroyl)]- β -D-glucopyranoside and kaempferol 3-O- β -D-apiofuranosyl-(1 \rightarrow 2)-[6-O-(3-hydroxy-3-methylglutaroyl)]- β -D-glucopyranoside	[709]
<i>Symplocarpus renifolius</i>	Araceae	Kaempferol 3-O- β -D-glucopyranosyl-(1 \rightarrow 2)- β -D-glucopyranosyl-7-O- β -D-glucopyranoside	[710]
<i>Syzygium aromaticum</i>	Myrtaceae	Kaempferol	[711]
<i>Tadehagi triquetrum</i>	Leguminosae	Kaempferol	[712]
<i>Tamarix nilotica</i>	Tamaricaceae	Kaempferol 4'-methyl ether	[713]
<i>Taxus baccata</i>	Taxaceae	Kaempferol and 7-O-glucosides kaempferol	[22]
<i>Tephrosia calophylla</i>	Leguminosae	Kaempferol 3-O- β -D-glucopyranoside	[714]
<i>Terminalia myriocarpa</i>	Combretaceae	Kaempferol 3-O- β -D-rutinoside	[715]
<i>Ternstroemia japonica</i>	Theaceae	Kaempferol derivative	[716]
<i>Tetrapanax papyriferus</i>	Araliaceae	Kaempferol 7-O-(2-E-p-coumaroyl- α -L-rhamnoside) and kaempferol 7-O-(2,3-di-E-p-coumaroyl- α -L-rhamnoside)	[717]
<i>Theobroma grandiflorum</i>	Sterculiaceae	Kaempferol	[718]
<i>Thesium chinense</i>	Santalaceae	Kaempferol and kaempferol 3-O-glucoside	[719]
<i>Thevetia peruviana</i>	Apocynaceae	Kaempferol 3-glucosyl-(1 \rightarrow 4)-[6"-sinapoylglucosyl]-(1 \rightarrow 2)-galactoside and 3-[2"-sinapoylglucosyl]-(1 \rightarrow 4)-[6"-sinapoylglucosyl]-(1 \rightarrow 2)-galactoside, kaempferol 3-[6"-sinapoylglucosyl]-(1 \rightarrow 2)-galactoside and kaempferol 3-glucosyl-(1 \rightarrow 2)-galactoside	[720]

(Table 1). Contd.....

Species	Family	Compounds	Reference
<i>Thyrocarpus glochidiatus</i>	Boraginaceae	Kaempferol 3-O- β -D-glucoside	[721]
<i>Tilia americana</i>	Tiliaceae	Kaempferol glycosides	[289]
<i>Tilia argentea</i>	Tiliaceae	Kaempferol 3,7-O- α -di-rhamnoside and kaempferol 3-O- β -D-(6"-E-p-coumaroyl)-glucopyranoside (tiliroside)	[86, 722]
<i>Tilia petiolaris</i>	Tiliaceae	Kaempferol 3-O-glucoside-7-O-rhamnoside	[292]
<i>Tilia tomentosa</i>	Tiliaceae	Kaempferol	[723]
<i>Tribulus terrestris</i>	Zygophyllaceae	Kaempferol 3-gentiobioside	[724]
<i>Trifolium alexandrinum</i>	Leguminosae	Kaempferol	[725]
<i>Trifolium resupinatum</i>	Leguminosae	Kaempferol glycosides	[726]
<i>Trigonella foenum-graecum</i>	Leguminosae	Kaempferol 3-O- β -D-glucosyl-(1 \rightarrow 2)- β -D-galactoside, kaempferol 3-O- β -D-glucosyl-(1 \rightarrow 2)- β -D-galactoside 7-O- β -D-glucoside and kaempferol 3-O- β -D-glucosyl-(1 \rightarrow 2)-(6"-O-acetyl)- β -D-galactoside-7-O- β -D-glucoside	[727]
<i>Tulipa gesneriana</i>	Liliaceae	Kaempferol 7-O-glucuronides	[728]
<i>Ullucus tuberosus</i>	Basellaceae	Kaempferol 3-O-(2",6"-di-O- α -L-rhamnopyranosyl)- β -D-glucopyranoside	[729]
<i>Uvaria tonkinensis</i>	Annonaceae	Kaempferol 3,7-O- α -L-di-rhamnoside	[730]
<i>Vaccinium vitis-idaea</i>	Ericaceae	Kaempferol-pentoside, kaempferol-deoxyhexoside and kaempferol 3-O-[4"-(3-hydroxy-3-methylglutaroyl)]- α -rhamnose	[731]
<i>Vahlia capensis</i>	Vahliaceae	Kaempferol	[732]
<i>Vernonia ferruginea</i>	Asteraceae	Kaempferol 3-O- β -D-apiofuranosyl-(1 \rightarrow 4)- β -D-glucopyranoside and kaempferol 4'-methyl ether 3-O- β -D-xylopyranoside	[733]
<i>Vernonia travancorica</i>	Asteraceae	Kaempferol 3-O- β -[β -(6"-acetyl)-D-glucopyranosyl-(1 \rightarrow 2)]-D-glucopyranosyl, 4'-methoxykaempferol, 3-O- β -D-glucopyranoside and kaempferol	[734]
<i>Vicia faba</i>	Leguminosae	Kaempferol 3-O-galactoside,7-O-rhamnoside	[735]
<i>Vinca minor</i>	Apocynaceae	Kaempferol glycosides	[736]
<i>Viola tricolor</i>	Violaceae	Kaempferol glycosides	[737]
<i>Vismia laurentii</i>	Clusiaceae	Kaempferol	[738]
<i>Vitis rotundifolia</i>	Vitaceae	Kaempferol glycosides	[739]
<i>Vitis vinifera</i>	Vitaceae	Kaempferol, kaempferol 3-glucoside, kaempferol 3-galactoside and kaempferol 3-glucuronide	[740]
<i>Waltheria indica</i>	Sterculiaceae	Kaempferol 3-O- β -D-(6"-O-coumaroyl)glucopyranoside (tiliroside)	[741]
<i>Warburgia stuhlmannii</i>	Canellaceae	Kaempferol, kaempferol 3-O- α -L-rhamnopyranoside and kaempferol 7-O- β -D-glucopyranoside	[742]
<i>Warburgia ugandensis</i>	Canellaceae	Kaempferol 3-O- α -rhamnoside-7,4'-di-O- β -galactoside, kaempferol 3,7,4'-tri-O- β -glucoside, kaempferol, kaempferol 3-rhamnoside, kaempferol 3-rutinoside, kaempferol 3-arabinoside, kaempferol 3-rhamnoside-4'-galactoside and kaempferol 3-glucoside	[743]
<i>Zelkova oregoniana</i>	Ulmaceae	Kaempferol	[744]
<i>Zingiber aromaticum</i>	Zingiberaceae	Kaempferol 3-O-(2,3-di-O-acetyl- α -L-rhamnopyranoside), kaempferol 3-O-(2,3,4-tri-O-acetyl- α -L-rhamnopyranoside) and kaempferol 3-O-methylether	[745, 746]

(Table 1). Contd.....

Species	Family	Compounds	Reference
<i>Zingiber zerumbet</i>	Zingiberaceae	Kaempferol 3,4',7-O-trimethylether, kaempferol 3-O-methylether, kaempferol 3,4'-O-dimethylether, 4''-O-acetylfazelin, kaempferol 3-O-(2,4-O-diacetyl- α -L-rhamnopyranoside) and kaempferol 3-O-(3,4-O-diacetyl- α -L-rhamnopyranoside)	[747, 748]
<i>Zollernia ilicifolia</i>	Leguminosae	Kaempferol 3-O- α -L-rhamnopyranosyl-(1 \rightarrow 2)-O-[α -L-rhamnopyranosyl-(1 \rightarrow 6)]-O- β -D-galactopyranoside-7-O- α -L-rhamnopyranoside	[749]

**Fig. (2).** Estructures of kaempferol and selected glycosides of kaempferol.

Liliaceae, Musaceae, Orchidaceae, Poaceae, Smilacaceae and Zingiberaceae (see Table 1 for references). In Dicotyledons, kaempferol has been identified in numerous families, including Actinidiaceae, Alangiaceae, Amaranthaceae, Anacardiaceae, Annonaceae, Apiaceae, Apocynaceae, Aquifoliaceae, Araliaceae, Asclepiadaceae, Asteraceae, Balsaminaceae, Basellaceae, Berberidaceae, Bignoniaceae, Boraginaceae, Brassicaceae, Cactaceae, Caesalpiniaceae, Campanulaceae, Canellaceae, Cannabaceae, Capparaceae, Caprifoliaceae, Caryophyllaceae,

Celastraceae, Chenopodiaceae, Chloranthaceae, Chrysobalanaceae, Cistaceae, Clusiaceae, Combretaceae, Convolvulaceae, Cornaceae, Corylaceae, Crassulaceae, Crypteroniaceae, Cucurbitaceae, Daphniphyllaceae, Dipsacaceae, Ebenaceae, Elaeagnaceae, Epacridaceae, Ericaceae, Euphorbiaceae, Fagaceae, Frankeniaceae, Gentianaceae, Geraniaceae, Grossulariaceae, Hydrangeaceae, Lamiaceae, Lauraceae, Lecythidaceae, Leguminosae, Loranthaceae, Lythraceae, Magnoliaceae, Malvaceae, Melastomataceae, Meliaceae, Monimiaceae,

Moraceae, Moringaceae, Myrsinaceae, Myrtaceae, Nelumbonaceae, Nepenthaceae, Nyctaginaceae, Nymphaeaceae, Ochnaceae, Oleaceae, Paeoniaceae, Papaveraceae, Phytolaccaceae, Platanaceae, Polygalaceae, Polygonaceae, Primulaceae, Punicaceae, Ranunculaceae, Resedaceae, Rhamnaceae, Rhizophoraceae, Rosaceae, Rubiaceae, Rutaceae, Salicaceae, Santalaceae, Sapindaceae, Saxifragaceae, Scrophulariaceae, Simaroubaceae, Solanaceae, Staphyleaceae, Sterculiaceae, Tamaricaceae, Theaceae, Thymelaeaceae, Tiliaceae, Ulmaceae, Vahliaceae, Violaceae, Vitaceae and Zygophyllaceae (see Table 1 for references).

Kaempferol has been identified in many edible plants. It has been found in *Allium ampeloprasum* (leek), *Allium cepa* (onion), *Allium schoenoprasum* (chives), *Amaranthus lividus* (amaranth), *Angelica keiskei* (ashitaba), *Armoracia rusticana* (horseradish), *Artemisia dracuncululus* (tarragon), *Atriplex hortensis* (orach), *Brassica campestris* (Chinese cabbage), *Brassica juncea* (mustard), *Brassica napobrassica* (rutabagas), *Brassica oleracea* (broccoli, brussels sprouts, green cabbage and kale), *Brassica rapa* (turnip greens), *Bunias orientalis* (Turkish rocket), *Camellia sinensis* (tea), *Capparis spinosa* (capers), *Celosia argentea* (feather cockscomb), *Cichorium endivia* (endive), *Citrus paradisi* (grapefruit), *Cnidioscolus aconitifolius* and *C. chayamansa* (tree spinach), *Coccinia grandis* (ivy gourd), *Cucumis sativus* (cucumber), *Cucurbita maxima* (squash), *Cyamopsis tetragonoloba* (cluster bean), *Diplotaxis eruroides* (wall rocket), *Diplotaxis tenuifolia* (wild rocket), *Eruca sativa* (rocket-salad), *Foeniculum vulgare* (fennel), *Fragaria vesca* (strawberry), *Houttuynia cordata* (fishwort), *Ipomoea batatas* (sweet potato), *Lactuca sativa* (lettuce), *Lepidium sativum* (cress), *Levisticum officinale* (lovage), *Lycium barbarum* and *L. chinense* (goji berries), *Malus domestica* (apple), *Momordica cochinchinensis* (gac), *Morinda citrifolia* (Indian mulberry), *Nasturtium officinale* (watercress), *Olea europaea* (olive oil), *Petroselinum crispum* (parsley), *Phaseolus vulgaris* (green beans), *Pistacia vera* (pistachio), *Prunus persica* (peach), *Raphanus sativus* (radishes), *Ribes uva-crispa* (gooseberries), *Rubus fruticosus* (blackberries), *Rubus idaeus* (raspberry), *Sambucus nigra* (elderberry), *Sauropus androgynus* (star gooseberry), *Sesbania grandiflora* (sesbania), *Solanum lycopersicum* (tomatoes), *Solanum nigrum* (nightshade), *Solanum tuberosum* (potatoes), *Spinacia oleracea* (spinach), *Vaccinium erythrocarpum*, *V. acrocarpon*, *V. microcarpum* and *V. oxycoccus* (cranberries), *Vaccinium vitis-idaea* (cowberries), *Vicia faba* (broadbeans) *Vigna unguiculata* (cowpea), *Vitis rotundifolia* (muscadine grapes) and *Vitis vinifera* (grapes) (for references, see Table 1, "USDA database for the flavonoid content of selected foods" or ref. [23, 24]).

Kaempferol has been identified in many plant species commonly used in traditional medicine. It has been found in *Acacia nilotica*, *Adansonia digitata*, *Albizia lebbek*, *Aloe vera*, *Amburana cearensis*, *Ammi majus*, *Angelica keiskei*, *Ardisia japonica*, *Bauhinia forficata*, *Bauhinia microstachya*, *Bunium persicum*, *Capparis spinosa*, *Cassia alata*, *Centella asiatica*, *Chromolaena odorata*, *Cissus sicyoides*, *Coccinia grandis*, *Crassocephalum crepidioides*,

Crocus sativus, *Cynanchum acutum*, *Cynanchum chinense*, *Dicliptera chinensis*, *Equisetum arvense*, *Euphorbia pekinensis*, *Ficaria verna*, *Foeniculum vulgare*, *Galega officinalis*, *Ginkgo biloba*, *Glycine max*, *Grindelia robusta*, *Gymnema sylvestris*, *Helleborus niger*, *Hippophae rhamnoides*, *Houttuynia cordata*, *Hypericum perforatum*, *Impatiens balsamina*, *Lamium album*, *Laurus nobilis*, *Lonicera japonica*, *Lycium barbarum*, *Lycium chinense*, *Lysimachia vulgaris*, *Malva parviflora*, *Peumus boldus*, *Phyllanthus emblica*, *Ribes nigrum*, *Rosmarinus officinalis*, *Sambucus nigra*, *Sanguisorba minor*, *Siraitia grosvenori*, *Solanum nigrum*, *Solenostemma argel*, *Solidago virgaurea*, *Sutherlandia frutescens*, *Symphytum officinale*, *Syzygium aromaticum*, *Tilia americana*, *Toona sinensis*, *Trigonella foenum-graecum*, *Tropaeolum majus*, *Vaccinium vitis-idaea*, *Warburgia ugandensis* and *Wedelia trilobata* (for references, see Table 1 or ref. [23]).

3. BIOLOGICAL ACTIVITIES OF KAEMPFEROL

3.1. Epidemiological Studies

The possible association between the consumption of foods containing kaempferol and a reduced risk of developing several disorders, including cancer and cardiovascular diseases, has been evaluated in several epidemiological studies. Some case-control (retrospective) and cohort (prospective) studies have evaluated the relationship between consumption of kaempferol-rich foods and the risk of developing several types of cancer. Cui *et al.* [25] conducted a case-control study of 558 lung cancer cases and a group of 837 controls, and observed that the consumption of kaempferol-rich foods (i.e. tea, beans, broccoli, spinach, apples and strawberries; approximately 2 mg kaempferol/day) was inversely associated with lung cancer risk (OR= 0.68; 95% CI: 0.51-0.90); this association was less clear among non-smokers. A case-control study conducted in Spain with 103 cases and 206 hospital controls also reported a positive association between high intake of kaempferol and a lower incidence of lung cancer (OR = 0.51; 95% CI: 0.22-1.17), although this association was not significant [26]. García-Closas *et al.* carried out a case-control study with 354 cases of gastric cancer and 354 controls and found that consumption of kaempferol-containing foods (i.e. onions, cruciferous, green beans, apples, grapes and strawberries) was associated with a reduced gastric cancer risk (OR = 0.48; p for trend = 0.04) [5]. Nöthlings *et al.* [27] conducted a cohort study to evaluate the possible association between consumption of kaempferol-containing foods (i.e. onions, tea, apples, cruciferous and other vegetables) and pancreatic cancer risk in 183518 people. After a follow-up period of 8 years, the authors found that kaempferol consumption was inversely associated with pancreatic cancer risk (RR = 0.78; 95% CI: 0.58-1.05; p trend: 0.017; the median intake of kaempferol was 3.89 mg/day). The correlation between intake of kaempferol-rich foods (i.e. broccoli, kale and tea) and incidence of epithelial ovarian cancer was evaluated in a cohort study in 66940 women. The study found that kaempferol was associated with a significant 40% decrease in epithelial ovarian cancer incidence (RR = 0.60; 95% CI: 0.42-0.87; p for trend: 0.002; median intake was 11 mg kaempferol/day) [28]. This association between kaempferol

intake and ovarian cancer risk, however, was not observed in a recent case-control study with 1141 cases and 1183 controls [29].

Other epidemiological studies did not find a positive correlation between consumption of kaempferol-containing foods and the risk of several other types of cancer. A case-control study carried out with 497 cases, 547 neighborhood controls and 566 hospital controls did not find a reduced bladder cancer risk in people consuming kaempferol-containing foods [30]. Likewise, McCann *et al.* [31] conducted a case-control study with 433 prostate cancer cases and 538 population-based controls and did not find a significant association between the consumption of kaempferol-containing foods (i.e. coffee, tea, green peppers, green beans, turnip greens, orange juice and apple juice) and a reduced prostate cancer risk (OR = 0.83; 95% CI: 0.58-1.18; p for trend: 0.80). A prospective study evaluated the association between flavonol-rich foods intake and breast cancer risk in 90630 women [32] and found a similar relative risk of developing breast cancer in the lower (0.80 mg kaempferol/day) and higher (12.9 mg kaempferol/day) quintile (MV RR = 1.01; 95% CI: 0.80-1.27; p for trend: 0.91). Lin *et al.* [33] evaluated the possible correlation between consumption of kaempferol-containing foods and the risk of colorectal cancer in a prospective study (71976 women and 35425 men) and found no association between increased intake of kaempferol and reduced colorectal cancer risk (RR = 1.12; 95% CI: 0.90-1.39; p for trend 0.25).

Several case-control and cohort studies have evaluated the relationship between consumption of kaempferol-rich foods and the risk of cardiovascular diseases. The possible correlation between flavonoid consumption (quercetin, kaempferol, myricetin, apigenin and luteolin) and mortality from coronary heart disease was studied in a cohort study in 805 men aged 65-84 years [1]. The study found that the consumption of flavonoid-containing foods (i.e. black tea, onions and apples) was associated with a reduced mortality from coronary heart disease (RR = 0.42; 95% CI; p for trend: 0.015) and with a reduced incidence of myocardial infarction (p trend: 0.08). Geleijnse *et al.* [34] conducted another cohort study in 4807 people followed-up for 5,6 years and observed that a high intake of tea (source of kaempferol) was inversely associated with myocardial infarction (RR = 0.57; 95% CI: 0.33-0.98). The correlation between intake of kaempferol-rich foods and the risk of coronary heart disease was also analyzed in a cohort study in 66360 women followed-up for 12 years [35]. This study found a weak risk reduction for coronary heart disease death among women with a higher intake of kaempferol-containing foods (i.e. broccoli, onions, grapes, apples and tea; RR = 0.66; 95% CI: 0.48-0.93; p trend: 0.04). In a cohort study with 361 men and 394 women aged 65-99 years and followed up for up to 10 years, kaempferol consumption was also associated with a significant decreased relative risk of acute myocardial infarction (RR = 0.481, 95% CI: 0.3-0.77; p trend: 0.002) [36]. The association between flavonoid-containing foods intake (i.e. apples, grapefruit, onions, white cabbage, berries, juices and oranges) and risk of several chronic diseases was assessed in a cohort study in 10054 people [37]. This study showed that the incidence of cerebrovascular disease was

lower at higher kaempferol intakes (RR = 0.70; 95% CI: 0.56-0.86; p: 0.003; median intake was 0.5 mg kaempferol/day). These data suggest that consumption of foods containing kaempferol may reduce the risk of developing some cardiovascular disorders.

The correlation between the consumption of kaempferol-containing foods and the occurrence of type 2 diabetes has been studied in a cohort study [38]. This study included 38018 women aged ≥ 45 and free of cardiovascular disease, cancer and diabetes. Although the authors found that none of the tested flavonoids (including kaempferol) was significantly associated with risk of type 2 diabetes, they observed a reduced relative risk of type 2 diabetes in people with an increased intake of the kaempferol-containing foods apples (RR: 0.72; p for trend: 0.006) and tea (RR: 0.73; p for trend: 0.06; 95% CI).

In summary, epidemiological data suggest that a high intake of kaempferol-containing foods may reduce the risk of developing several types of cancers (e.g. lung, gastric, pancreatic and ovarian cancer) and cardiovascular diseases. But the relatively low number of studies and the possible presence of other bioactive constituents in kaempferol-containing foods (e.g. vitamins, minerals and other phytochemicals) make these data insufficient to draw any conclusion regarding the possible protective effect of kaempferol in these diseases. Numerous experimental data have shown, however, that kaempferol possesses a wide range of biological activities involved in the prevention and treatment of these and other diseases. The biological activities of kaempferol are summarized and analyzed in the following sections.

3.2. Antioxidant Activity

We breathe because our cells need oxygen to generate energy in a mitochondrial process called oxidative phosphorylation (oxphos). In this process, the generation of energy in the form of ATP is coupled with a reaction in which oxygen (O_2) is reduced to H_2O . In this reaction, four electrons and four protons are added to O_2 to form two molecules of H_2O . But when a molecule of O_2 gains only one electron to form superoxide anion ($O_2^{\bullet -}$), this highly reactive oxygen species (ROS) tends to gain three more electrons and four protons to form H_2O ; this process involves several reactions and results in the production of other ROS such as hydrogen peroxide (H_2O_2), hydroxyl radical and peroxynitrite. The controlled production of ROS has an important physiological role [39]. A high production of ROS that is not counterbalanced by the cellular antioxidant defense, however, increases the cellular levels of ROS and originates oxidative stress. Oxidative stress has been proposed to play an important role in the pathogenesis of cancer, cardiovascular disease, atherosclerosis, hypertension, ischemia/reperfusion injury, diabetes mellitus, neurodegenerative disorders, rheumatoid arthritis, and ageing [40, 41]. Antioxidant agents can reduce oxidative stress and may therefore play a protective role in the development of these processes.

The antioxidant properties of flavonoids are widely acknowledged [42-44]. Numerous reports have shown that kaempferol, some glycosides of kaempferol, and several

kaempferol-containing plants have antioxidant activity not only *in vitro*, but also *in vivo* [45-50]. Several studies have shown that the presence of a double bond at C2-C3 in conjugation with an oxo group at C4, and the presence of hydroxyl groups at C3, C5 and C4', are important structural features involved in the antioxidant activity of kaempferol [42, 58].

Kaempferol has been found to be a potent superoxide scavenger, with an IC_{50} of 0.5 μ M [51]. The ability of kaempferol to decrease superoxide levels at low concentrations may play an important role in its antioxidant activity, as the formation of superoxide anion is required for the normal production of most reactive oxygen and nitrogen species involved in oxidative stress [52]. Superoxide anion is commonly converted into H_2O_2 by the enzymes superoxide dismutases. In the presence of reduced transition metals (e.g., ferrous or cuprous ions), H_2O_2 can be converted into the highly reactive hydroxyl radical. Superoxide anion can also react with nitric oxide to form peroxynitrite. Hydroxyl radical and peroxynitrite are highly reactive species known to cause damage to DNA, proteins and lipids. Interestingly, low concentrations of the flavonol kaempferol have been found to scavenge these two highly reactive species. Wang *et al.* [51] evaluated the antioxidant activity of several flavonoids and found that kaempferol was one of the strongest scavengers for the Fenton-generated hydroxyl radical, with an IC_{50} of 0.5 μ M. Likewise, Heijnen *et al.* [53] observed that kaempferol was a potent peroxynitrite scavenger, with an IC_{50} of 0.35 ± 0.5 μ M.

The ability of kaempferol to decrease superoxide anion, hydroxyl radical and peroxynitrite levels at submicromolar concentrations may play a key role in its antioxidant activity. Kaempferol has also been found to exert antioxidant effects through other mechanisms of action, although most of these effects have been observed at higher concentrations. Kaempferol can inhibit the activity of enzymes that generate ROS, such as the enzyme xanthine oxidase [51, 54, 55]. Like other flavonoids, kaempferol can also reduce the formation of hydroxyl radical through the Fenton's reaction by chelating ferrous or cuprous ions [56-58]. Kaempferol may also induce antioxidant effects by increasing the expression or activity of antioxidant enzymes such as superoxide dismutase, catalase, and heme oxygenase-1 [59-61]. Other studies have revealed that kaempferol and some glycosides of kaempferol prevent lipid peroxidation [58, 62-69]; the activity of kaempferol is higher than that of its glycosides because kaempferol has a higher lipophilia and capacity to penetrate into lipid bilayers to exert this activity. Kaempferol can also prevent the oxidation of low-density lipoprotein (LDL), which may play a protective role in atherosclerosis [70-74].

The antioxidant activity of kaempferol has been observed in several *in vivo* studies [45-50]. For instance, kaempferol was found to decrease intracellular ROS accumulation and increase the survival of *Caenorhabditis elegans* [45]. An extract of *Capparis spinosa*, which contained kaempferol derivatives, showed a significant antioxidant activity when applied topically in healthy human volunteers [46]. Sanz *et al.* [47] treated mice with kaempferol-3-O-galactoside after bromobenzene intoxication and observed that this flavonoid

reduced hepatic lipid peroxidation products and increased the levels of reduced glutathione. The intraperitoneal injection of an extract of *Crassocephalum crepidioides* that contained kaempferol glycosides showed a strong antioxidant effect and prevented galactosamine- and lipopolysaccharide-induced hepatotoxicity in rats [50]. A *Ginkgo biloba* extract, containing kaempferol and quercetin derivatives, has also been reported to exert antioxidants effects *in vivo* [48].

3.3. Anti-Inflammatory Activity

Inflammation is a physiological process in response to tissue damage resulting from microbial pathogen infection, chemical irritation, and/or wounding. After tissue injury, a multifactorial network of chemical signals initiates and maintains a host response designed to heal the damaged tissue. The activation and migration of leukocytes to the site of damage and the release of growth factors, cytokines and reactive oxygen and nitrogen species are known to play a crucial role in the inflammatory response. Inflammatory processes are required for immune surveillance, optimal repair, and regeneration after injury. When acute inflammation is not resolved, however, chronic inflammation occurs, which has a detrimental effect in several diseases including atherosclerosis, cancer, asthma and some neurological disorders, such as Alzheimer's disease and Parkinson's disease [75]. Like other flavonoids [76], kaempferol has anti-inflammatory properties. Numerous reports have shown that kaempferol, kaempferol glycosides and/or kaempferol-containing plants have anti-inflammatory activity not only *in vitro* but also *in vivo* [77-88].

The anti-inflammatory activity of kaempferol may be mediated by several mechanisms of action. The activation of the nuclear factor kappa B (NF- κ B) increases the expression of pro-inflammatory cytokines, chemokines and enzymes (e.g. TNF- α , IL-1, IL-6, IL8, COX-2, iNOS), and several works have shown that kaempferol inhibits NF- κ B activity [51, 77, 78, 90-93]. Several studies have also shown that kaempferol can inhibit TNF- α activity [94-99] and IL-1 β and IL-8 expression [94, 100]. Activator protein 1 (AP-1) is a transcriptional regulator composed of members of the Fos and Jun families that participates in the inflammatory response, and kaempferol has been shown to inhibit the activation of AP-1 [96, 101]. The enzymes cyclooxygenases (COX), lipoxygenases (LOX) and inducible nitric oxide synthase (iNOS) are known to play important roles in inflammation by participating in the synthesis of eicosanoids (e.g. prostaglandins, leukotrienes) and in the production of reactive species. Some reports have shown that kaempferol can inhibit COX-2 [90, 102-107], LOX [108, 109] and iNOS [89, 90, 105, 106, 110]. ROS are also known to participate in the inflammatory process and, as shown in the previous section, kaempferol can reduce the cellular levels of ROS. In summary, numerous *in vitro* and *in vivo* studies have revealed that kaempferol has anti-inflammatory activity and have shown several mechanisms that may participate in this activity.

3.4. Anticancer Activity

Epidemiological evidence (case-control and cohort studies) suggests that the consumption of kaempferol-rich

foods may reduce the risk of developing some types of cancer, including lung cancer [25, 26], gastric cancer [5], pancreatic cancer [27] and ovarian cancer [28]. Although few studies have evaluated the anti-carcinogenic activity of kaempferol in animal models [111], numerous preclinical studies have shown that this flavonoid has cancer preventive and therapeutic properties.

The most accepted view of carcinogenesis (the somatic mutation theory of cancer) considers that cancer is caused by DNA alterations [112], and several reports suggest that low concentrations of kaempferol may protect DNA from damage induced by different carcinogens [113-116]. It is widely accepted that the formation of a malignant tumor requires that tumor cells acquire several capabilities (the so-called hallmarks of cancer), such as apoptosis resistance, increased angiogenesis, or capacity of invasion and metastasis [118, 119]. The formation of a cancer requires that tumor cells develop apoptosis resistance, and it has been observed that kaempferol can induce apoptosis [120-127]. Angiogenesis (the generation of new blood vessels) is necessary for the formation of solid tumors; without vascular growth, the tumor mass is restricted to a tissue-diffusion distance of approximately 0.2 mm. Malignant tumors are known to activate angiogenesis, and kaempferol has been shown to inhibit angiogenesis *in vitro* [128-130]. It is recognized that the metastatic spread of primary tumors accounts for approximately 90% of all cancer deaths; the process by which cells from a localized tumor invade adjacent tissues and metastasize to distant organs may therefore be the most clinically relevant process involved in carcinogenesis [131]. Experimental data have revealed that kaempferol may inhibit this process *in vitro* and/or *in vivo* [93, 132-134].

Accumulating evidence suggests that reactive oxygen species (ROS) have an important role in the development of cancer. The key role of ROS in carcinogenesis is supported by experimental data showing that cancer cells commonly have increased levels of ROS, that ROS can induce cell malignant transformation and that the malignant phenotype of cancer cells can be reversed by reducing the cellular levels of ROS [135-138]. Because antioxidant agents can prevent the accumulation and/or reduce excessive cellular levels of ROS, they may play a protective role in cancer development. For instance, experimental data revealed that the expression of antioxidant enzymes in malignant cells decreased their cellular levels of ROS; these cells reverted to a normal appearance, their growth rate normalized, and they were no longer capable of producing tumors in athymic mice [137]. These data suggest that the extensively reported antioxidant activity of kaempferol may be a key mechanism by which this dietary flavonoid prevents cancer.

Several lines of research suggest that inflammation plays a significant role in cancer [139]. It is recognized that inflammatory diseases increase the risk of developing many types of cancer, including bladder, cervical, gastric, intestinal, esophageal, ovarian, prostate and thyroid cancer. Inflammatory cells, chemokines and cytokines are also present in the microenvironment of all tumors in experimental animal models and humans from the earliest stages of development. Transfer of inflammatory cells or

overexpression of inflammatory cytokines promotes the development of tumors. In addition, the targeting of inflammatory mediators (chemokines and cytokines, such as TNF- α and IL-1 β), transcription factors involved in inflammation (such as NF- κ B and STAT3) or inflammatory cells decreases the incidence and spread of cancer. Non-steroidal anti-inflammatory drugs are also known to reduce the risk of developing certain cancers (such as colon and breast cancer) and reduce the mortality caused by these cancers [139]. These data suggests that the anti-inflammatory properties of kaempferol may play an important role in the cancer preventive activity of this flavonoid.

Most chemical carcinogens need to be enzymatically activated to become genotoxic, and the cytochrome P450 (P450) enzymes are the most prominent enzymes involved in such activation [140]. Some studies have shown that kaempferol may inhibit P450 [141, 142] and therefore prevent the activation of carcinogenic agents. Kaempferol has been found to decrease the activation of the naturally occurring mycotoxin aflatoxin in rats [143]. This flavonoid has also been found to act as a potent inhibitor of the aryl hydrocarbon receptor (AhR); this receptor is activated by human carcinogens present in air pollution and cigarette smoke [144]. The authors observed that kaempferol also inhibited the ability of cigarette smoke condensate to induce growth of immortalized lung epithelial cells [144]. Mukai *et al.* found that oral administration of kaempferol or *Ginkgo biloba* extract (EGb) containing 24% flavonol at 100 mg/kg body weight suppressed AhR transformation induced by 3-methylcholanthrene in the liver of mice [145]. Kaempferol has also been described as a potent non-competitive inhibitor of sulfotransferase 1A1, an enzyme capable of bioactivating procarcinogens to reactive electrophiles [146]. In addition to inhibiting several phase I enzymes, kaempferol may induce phase II detoxifying enzymes, such as quinone reductase [147]. Kaempferol has also been found to reduce the cellular levels of the carcinogen 7,12-dimethylbenz(a)-anthracene by potently stimulating the P-glycoprotein-mediated efflux of this carcinogen [148].

Hypoxia-inducible factor 1 (HIF-1) is a key regulator of O₂ homeostasis. The activation of HIF-1 is known to play a vital role in the most relevant aspects of carcinogenesis, including cell survival, angiogenesis, invasion, metastasis, cellular immortalization and metabolic reprogramming [149-151]. HIF-1 activation is observed in most human cancers and has been associated with increased patient mortality. For instance, Zhong *et al.* [152] identified increased HIF-1 expression (relative to adjacent normal tissue) in 13 tumor types, including lung, prostate, breast, and colon carcinoma, which are the most common cancers in developed countries. These data suggest that HIF-1 activation is a key event in carcinogenesis and may therefore represent a key target for cancer chemoprevention [153]. Mylonis *et al.* [154] investigated the effect of kaempferol on activity, expression levels and localization of HIF-1 in hepatoma cancer cells and found that this dietary flavonoid inhibited this transcription factor in the low micromolar range. HIF-1 was also down-regulated by kaempferol in ovary cancer cells [155].

Kaempferol has shown other activities that may be relevant to cancer chemoprevention. The ribosomal S6 kinase 2 (RSK2), a member of the p90(RSK) family of proteins, is a widely expressed serine/threonine kinase that is activated by extracellular signal-regulated kinase 1/2 and phosphoinositide-dependent kinase 1 in response to many growth factors and peptide hormones. Its activation is known to be involved in proliferation and cell transformation induced by carcinogens. Kaempferol 3-*O*-(3",4"-di-*O*-acetyl- α -L-rhamnopyranoside), also known as compound SL0101 (Fig. 2), was identified as the first specific inhibitor of this kinase (IC_{50} = 89 nM) and was also found to suppress proliferation of MCF-7 breast cancer cells [156]. The related compound 3Ac-SL0101 also inhibited this kinase in LNCaP prostate cancer cells [157]. This activity has also been shown by its aglycone kaempferol [158, 159]. Kaempferol can also act as a weak estrogen receptor agonist and may cause estrogenic or antiestrogenic effects mainly depending on the concentration of endogenous estrogens. The antiestrogenic (and weak estrogenic) activity of kaempferol may result in inhibition of the growth of hormone-dependent cancers such as breast and prostate cancers [160-163]. Kaempferol may also induce differentiation in colon cancer cells by re-establishing gap junctional intercellular communications, which are commonly impaired in cancer [164].

In addition to having cancer chemopreventive properties, kaempferol has shown activities that may have relevance to cancer therapy. Numerous reports have shown that kaempferol and/or some kaempferol glycosides induce cell death in a variety of cancer cells from different tissues, including lung [124, 165-167], breast [120, 122, 127, 161, 168, 169], colon [159, 165, 170, 171], prostate [120, 172, 173], liver [154, 174], pancreas [126], blood/lymph [117, 123, 175-181], skin [159, 165, 167, 176, 182, 183], esophagus [184, 185], brain [125, 165, 186], uterus [182, 187], ovary [155, 165, 188, 189], thyroid [190] and bone [121]. Many of these reports have shown that kaempferol induces cell death through apoptosis, and the possible mechanisms involved in this process have been elucidated. Caspases are a family of cysteine proteases involved in the initiation and execution of apoptosis, and kaempferol has been found to induce the activation of caspase-3 [123, 125, 166, 170, 179, 184-186, 191], caspase-7 [122, 124] and caspase-9 [122, 123, 179, 184, 185]. This flavonoid can also activate the apoptosis-inducing factor (AIF), which is involved in the initiation of a caspase-independent pathway of apoptosis [166]. Several studies have shown that kaempferol can decrease the antiapoptotic proteins Bcl-2 [123-125, 191] and Bcl-X1 [124] and increase the proapoptotic proteins Bax and Bad [122-124]. This dietary agent can also down-regulate the antiapoptotic protein X-linked inhibitor of apoptosis protein (XIAP) [186]. Poly-ADP-ribose polymerase (PARP) activation, cytochrome c release and loss of mitochondrial membrane potential are important events in the induction of apoptosis, and kaempferol has been found to stimulate PARP [123, 125, 127, 179] and induce mitochondrial cytochrome c release and loss of mitochondrial membrane potential [123, 125, 170, 179, 186]. Filomeni *et al.* [192] have recently reported that kaempferol induces energetic failure due to inhibition of both glucose uptake and complex I of the mitochondrial

respiratory chain; this results in the activation of autophagy, which may prevent to some extent the capacity of kaempferol to induce apoptosis.

Several other mechanisms may participate in kaempferol-induced cancer cell death. Kaempferol-induced apoptosis in cancer cells has been associated with its ability to inhibit the activity of fatty acid synthase (FAS), a lipogenic enzyme that is activated in cancer cells [120, 193]. The enzymes DNA topoisomerases I and II are the target of several anticancer drugs commonly used in the clinic (e.g. etoposide, doxorubicin, topotecan and irinotecan), and kaempferol has been found to inhibit topoisomerase I [194] and topoisomerase II [195, 196]. Kaempferol can induce cell cycle arrest at G2/M [117, 122, 172, 183, 185, 191, 197], which may be mediated, for instance, by the inhibition of the activity of the cyclin-dependent kinase CDK1 [183]. Proteasomes are large protein complexes that degrade unneeded or damaged proteins by proteolysis; these complexes play an important function in regulating protein levels and are necessary for cell survival. Proteasome inhibitors are a new class of anticancer agents (e.g. bortezomib) and kaempferol has been found to act as a proteasome inhibitor in human leukemia cells [198]. The antiproliferative effects of kaempferol in cancer cells may also be mediated by the inhibition of the MAPK/ERK pathway [124, 127, 186]. Finally, although low concentrations of kaempferol can reduce the cellular levels of ROS and induce antioxidant effects, higher concentrations of this flavonoid are known to generate ROS [123, 125, 179, 186]. The induction of ROS in cancer cells is emerging as an important anticancer strategy [138, 199-201] and may play an important role in the cytotoxic effects of kaempferol [123, 125, 127].

Evidence suggests that kaempferol could be used in combination with several anticancer drugs to improve their therapeutic effects. It has been reported that kaempferol can sensitize cancer cells to the cytotoxic effects of cisplatin [189], 5-fluorouracil [126], cytarabine [202], doxorubicin [125], mitoxantrone and the active metabolite of irinotecan (SN-38) [203]. This flavonoid can also enhance the cytotoxic effects of tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) [204, 205]. These investigations suggest that kaempferol may have clinical applications as adjuvant therapy in the treatment of some cancers.

3.5. Antimicrobial Activity

Flavonoids are known to play a protective role against microbial invasion in plants that synthesize these polyphenols. This protective role involves the presence of flavonoids in plants as constitutive agents as well as their accumulation as phytoalexins in response to microbial attack [206]. It is not surprising, therefore, that plants rich in flavonoids have been used for many years in traditional medicine to treat infectious diseases [207]. Kaempferol and its glycosides have been isolated from plants used in popular medicine for their antimicrobial properties. Numerous papers have reported that kaempferol, its glycosides, or plants containing kaempferol have antibacterial, antiviral, antifungal and antiprotozoal activities.

The antibacterial activity of kaempferol has been widely reported. Table 2 shows bacteria species inhibited by kaempferol and/or by glycosides of kaempferol. The antibacterial activity of this flavonol has been observed *in vivo*. Four weeks after inoculating Mongolian gerbils with *Helicobacter pylori* orally, Kataoka *et al.* administered kaempferol orally twice a day for 10 days and observed that this flavonoid significantly decreased the number of *Helicobacter pylori* colonies in the stomach of the gerbils [208]. Habbu *et al.* [209] observed that the oral administration of kaempferol-7-O-methyl-3-sulphate (2 mg/Kg, daily for 7 days) significantly increased the survival of mice infected with the bacteria *Klebsiella pneumoniae* ($p < 0.001$). Kaempferol and its glycosides can also act synergistically with antibiotics (e.g. rifampicin, vancomycin, methicillin, erythromycin and clindamycin) against antibiotic-resistant bacteria [210-212], therefore suggesting that kaempferol could be used in combination with these drugs in cases of resistance.

Kaempferol has shown anti-viral activity against several viruses, including herpes simplex virus [213-215], cytomegalovirus [216], influenza virus [217], and human immunodeficiency virus (HIV) [218, 219]. Kaempferol can inhibit the enzyme reverse transcriptase [218, 220] and it has been suggested that the hydroxyl groups at C3 and C4' are necessary for the inhibition of this enzyme [220]. This activity, along with its ability to inhibit viral proteases and binding of gp120 to lymphocytes CD4, may account for the anti-HIV activity of this flavonoid [219]. Jeong *et al.* [217] observed that some compounds isolated from *Rhodiola rosea*, including kaempferol, inhibited neuraminidase activity and showed high inhibitory activity against the influenza viral strains H1N1 and H9N2. Mitrocotsa *et al.* [216] evaluated the antiviral activity of seven kaempferol derivatives against human cytomegalovirus and observed that the presence of acyl substituents markedly increased the activity; the most active compound was kaempferol-3-O- α -L-(2'',3''-di-E-p-acetylcoumaroyl)-rhamnopyranoside.

Some studies have found that kaempferol interferes with enzymes that are vital for growth or virulence of certain fungi [221-224]. Chitin is a structural polysaccharide of fungal cell walls, which is vital for its integrity and is synthesized by chitin synthase. Hwang *et al.* [221] carried out an *in vitro* study with several flavonoids and found that kaempferol and kaempferol-3-O-(2''-O-galloyl)- β -D-glucoside inhibited chitin synthase II and prevented fungal cell division. It has also been shown that kaempferol is able to inhibit melanin synthesis [223]; melanin is a virulence factor in pathogenic fungi and a known target for antimycotic compounds. Yordanov *et al.* [222] found that kaempferol affected extracellular enzymes of *Candida albicans*, which are claimed to be virulence factors responsible for penetration of the yeast into host cells. Kaempferol administration to animals with systemic *Candida albicans* infection was found to increase the number of survivors. The authors also showed that the application of kaempferol in cutaneous infection suppressed the symptoms and accelerated the elimination of the yeast from the site of inoculation [222].

Kaempferol has also been described as an antiprotozoal agent [225-228]. This flavonoid has shown activity against *Leishmania* spp [229, 230] and is also active against *Entamoeba histolytica* and *Giardia lamblia* [227], which are known to cause diarrhea. The anti-giardial activity of kaempferol has been observed *in vivo* in female CD-1 mice infected with *Giardia lamblia* [225].

3.6. Other Biological Activities

Several case-control and cohort studies evaluated the relationship between consumption of kaempferol-rich foods and the risk of developing or dying of cardiovascular diseases. These studies revealed that the consumption of kaempferol-containing foods was associated with a reduced mortality from coronary heart disease [1], with a weak risk reduction for coronary heart disease death [35], with a reduced incidence of myocardial infarction [1, 34, 36], and with a lower incidence of cerebrovascular disease [37]. The protective role of kaempferol in cardiovascular diseases may be mediated by different mechanisms. Because oxidative stress [231] and inflammation [232] are known cardiovascular risk factors, it makes sense to think that the known antioxidant and anti-inflammatory properties of kaempferol may play a critical role in this protective effect. The ability of kaempferol to prevent atherosclerosis may be mediated, for instance, by its capacity to prevent the oxidation of the lipoproteins LDL [70-74]. Kaempferol has also been shown to inhibit angiotensin converting enzyme (which converts angiotensin I to angiotensin II and causes an elevation of blood pressure) [233-236], to induce vasodilator effects [237-239], and to cause antiplatelet and antithrombotic effects [240, 241]. Kaempferol and some glycosides of kaempferol may also decrease triglycerides levels, cholesterol levels and/or reduce body weight [242-244].

Evidence suggests that some kaempferol glycosides and several kaempferol-containing plants have antidiabetic activity [245-254] and may prevent diabetic complications [255, 256]. For instance, studies with kaempferol-3,7-O- α -dirhamnoside (kaempferitrin) and kaempferol 3-neohesperidoside (Fig. 2), isolated from *Cyathea phalerata* stems, showed a significant hypoglycemic effect in diabetic rats [257]. This antidiabetic effect may be mediated by stimulation of glycogen synthesis, and Cazarolli *et al.* found that the phosphatidylinositol 3-kinase (PI3K)-glycogen synthase kinase 3 (GSK-3) pathway and the MAPK-protein phosphatase 1 (PP1) pathway were involved in the stimulatory effect of kaempferol 3-neohesperidoside on glycogen synthesis in rat soleus muscle [258]. This flavonoid was also found to induce a stimulatory effect on glucose uptake when the rat soleus muscle was incubated with very low concentrations of this kaempferol glycoside (1 and 100 nM) [259]. The authors also showed that 100 mg/kg of kaempferol 3-neohesperidoside administered by oral gavage was able to increase glycogen content in the muscle, and suggested that this flavonoid stimulates glucose uptake in the rat soleus muscle *via* the PI3K and PKC pathways [259]. The oral administration of kaempferitrin has also been found to induce a significant hypoglycemic effect in normal and in alloxan-induced diabetic rats [246]. A single oral administration of two extract from the aerial parts of

Table 2. Antibacterial Activity of Kaempferol and Glycosides of Kaempferol

Species	Compound	Reference
<i>Staphylococcus aureus</i>	Kaempferol 3-O-[3-O-acetyl-6-O-(E)-p-coumaroyl]- β -D-glucopyranoside and kaempferol 3-O- β -D-glucopyranoside (astragalol)	[678]
	Kaempferol 7-O methyl 3- sulphate	[209]
	Kaempferol 3-O- α -L-(2'',4''-di-E-p-coumaroyl)-rhamnoside and kaempferol 3-O- α -L-(2''-Z-p-coumaroyl-4''-E-p-coumaroyl)-rhamnoside	[211]
	Kaempferol 3-O-rutinoside	[577]
	Kaempferol	[212, 750]
	Kaempferol 3,7-O- α -L-di-rhamnoside	[224]
<i>Staphylococcus epidermidis</i>	Kaempferol 3-O-[3-O-acetyl-6-O-(E)-p-coumaroyl]- β -D-glucopyranoside and kaempferol 3-O- β -D-glucopyranoside (astragalol)	[678]
<i>Pseudomonas aeruginosa</i>	Kaempferol 3-O-[3-O-acetyl-6-O-(E)-p-coumaroyl]- β -d-glucopyranoside	[678]
<i>Enterococcus faecalis</i>	Kaempferol 7-O methyl 3- sulphate	[209]
	Kaempferol	[751]
	Kaempferol 3,7-O- α -L-di-rhamnoside	[224]
<i>Enterobacter cloacae</i>	Kaempferol 3-O-[3-O-acetyl-6-O-(E)-p-coumaroyl]- β -D-glucopyranoside and kaempferol 3-O- β -D-glucopyranoside (astragalol)	[678]
<i>Escherichia coli</i>	Kaempferol 3-O-[3-O-acetyl-6-O-(E)-p-coumaroyl]- β -D-glucopyranoside	[678]
	Kaempferol 7-O methyl 3- sulphate	[209]
	Kaempferol 3-O-(2''-O- β -D-glucopyranosyl)- α -L-rhamnopyranoside	[16]
	Kaempferol 3,7-O- α -L-di-rhamnoside	[224]
<i>Klebsiella pneumoniae</i>	Kaempferol 3-O-[3-O-acetyl-6-O-(E)-p-coumaroyl]- β -D-glucopyranoside and kaempferol 3-O- β -D-glucopyranoside (astragalol)	[678]
	Kaempferol 7-O methyl 3-sulphate	[209]
	Kaempferol 3,7-O- α -L-di-rhamnoside	[224]
<i>Mycobacterium tuberculosis</i>	Kaempferol 7-O methyl 3-sulphate	[209]
<i>Acinetobacter baumannii</i>	Kaempferol 3,7-O- α -L-di-rhamnoside	[224]
<i>Bacillus subtilis</i>	Kaempferol 3,7-O- α -L-di-rhamnoside	[224]
<i>Porphyromonas gingivalis</i>	Kaempferol	[711]
<i>Prevotella intermedia</i>	Kaempferol	[711]
<i>Helicobacter pylori</i>	Kaempferol	[208, 671]
<i>Clostridium perfringens</i>	Kaempferol 3-O- α -(6'''-p-coumaroyl)glucosyl- β -1,4-rhamnoside)	[16]
<i>Vibrio cholerae</i>	Kaempferol	[751]
<i>Propionibacterium acnes</i>	Kaempferol	[210]

Equisetum myriochaetum also reduced blood glucose levels in diabetic rats; the authors proposed that kaempferol-3-sophoroside-4'-O- β -D-glucoside was responsible for this activity [248]. The hypoglycemic effect of a water extract from the aerial parts of *Equisetum myriochaetum* was later evaluated in eleven type 2 diabetic patients [249]. The authors found that the oral administration of the extract (0.33

g/kg) significantly reduced blood glucose levels in these diabetic patients without significantly affecting insulin levels [249]. Fang *et al.* observed that kaempferol improved insulin-stimulated glucose uptake in mature adipocytes and suggested that this flavonoid could potentially act on multiple targets to ameliorate hyperglycemia, including the peroxisome proliferator-agonist receptor γ (PPAR γ) [260].

Kaempferol may also prevent the onset of diabetes by preventing oxidative damage in pancreatic β cells [261]. All these studies support the idea that kaempferol, several glycosides of kaempferol and/or some kaempferol-containing plants have potential to be developed as antidiabetic agents.

Phytoestrogens are a group of compounds from plant origin that share a similar structure with the estrogenic compound estradiol. These compounds can bind to the estrogen receptors and produce estrogenic effects, although these effects are weaker than those of 17β -estradiol. Because phytoestrogens compete with estrogens for the same receptors, phytoestrogens can act as estrogenic or antiestrogenic compounds mainly depending on the concentration of endogenous estrogens. When the levels of estrogens are low (e.g. after menopause), phytoestrogen act as estrogenic compounds and may relieve the menopause symptoms and cause beneficial effects on cardiovascular disease, osteoporosis, Alzheimer's disease, etc. When the levels of estrogens are not decreased, phytoestrogens act as antiestrogenic compounds and may be beneficial for preventing the development of some cancers that need estrogens for their growth, such as some breast and prostate cancers. Several reports have shown that kaempferol is a phytoestrogen that cause both estrogenic and antiestrogenic effects [161, 162, 262]. This flavonoid can bind to both subtypes of the estrogen receptor (ER), ER- α and ER- β , although it has been observed that the affinity of kaempferol for the ER- β receptor is higher than for the ER- α receptor [161, 162, 262-265]. The hydroxyl group at C-4' seems crucial for the estrogenic activity of kaempferol [266]. The antiestrogenic (and weak estrogenic) activity of kaempferol may result in inhibition of the growth of hormone-dependent cancers such as breast and prostate cancers [160-163]. The estrogenic effects of kaempferol may be beneficial for women after menopause, when estrogen levels fall. It has been reported that kaempferol potentiated the uterotrophic effect of 17β -estradiol in immature and ovariectomized rats [267] and increased sensitivity of the uterus to estrogens in immature female mice [268].

Kaempferol may exert a protective effect against post-menopausal bone loss, as it has been found to promote osteoblast mineralization *in vitro* and bone formation *in vivo* [269-274]. Wattel *et al.* [269] observed that kaempferol, at concentrations ranging from 100 nM to 100 μ M reduced bone resorption in a time and dose-dependent manner. The authors observed significant inhibitory effects at concentrations as low as 100 nM and reported data suggesting that the estrogenic activity of kaempferol could be involved in the inhibition of bone resorption [269]. Trivedi *et al.* [270] reported that kaempferol increased mineralized nodules in rat primary osteoblasts at concentrations from 0.2 to 5 μ M. Daily oral administrations of kaempferol (5 mg/kg body weight) for 10 weeks to ovariectomized rats resulted in significantly higher bone mineral density (BMD) in the trabecular regions (femur neck, proximal tibia and vertebrae) and lower serum ALP (a bone turnover marker). Kaempferol treatment also caused an increase in osteoprogenitor cells as well as a reduction of adipocyte differentiation from bone marrow cells. The

authors concluded that kaempferol exerts bone anabolic activity with attendant inhibition of bone marrow adipogenesis [270]. An extract of *Ginkgo biloba*, which contains kaempferol glycosides, was also found to restore bone mass in aged ovariectomized rats [271].

Some *in vitro* and *in vivo* studies suggest that kaempferol, some glycosides of kaempferol and plants containing this flavonoid may have neuroprotective activity and play a protective role in the development of Alzheimer's disease, Parkinson's disease or Huntington's disease [275-283]. For instance, Lopez-Sanchez *et al.* [275] carried out an *in vivo* study to evaluate the neuroprotective effect of kaempferol and found that the intravenous administration of this dietary agent decreased ischemia-induced brain damage in rats. This protective effect was associated with the capacity of kaempferol to reduce metalloproteinase activation, to prevent protein nitrotyrosines accumulation and to protect against apoptotic cell death caused by oxidative stress [275]. Because oxidative stress is known to play an important role in some neurodegenerative disorders [40, 41] it makes sense to think that the antioxidant activity of kaempferol participates in its neuroprotective activity [284]. Other mechanisms, such as reduction in amyloid β protein, may participate in the possible protective effect of kaempferol against Alzheimer's disease [279, 285-287].

Several reports suggest that kaempferol, glycosides of kaempferol such as tiliroside (Fig. 2) and several kaempferol-containing plants (e.g. *Tilia* species) have anxiolytic activity *in vivo* [288-292]. *In vitro* and *in vivo* studies have also reported that kaempferol have antiallergic and antiasthmatic activity [84, 97, 293-298], which may be mediated, at least in part, by inhibition of histamine release [97, 293, 294]. Analgesic activity has also been observed with several kaempferol glycosides and kaempferol-containing plants in numerous *in vivo* studies [79, 83, 85-88, 299-302].

Okamoto *et al.* [303] reported data suggesting that kaempferol could be useful for the treatment of cell mediated immune diseases, such as acute graft-versus-host disease (GVHD). The authors found that kaempferol acted directly on T cells and inhibited Th1 cytokine production, thus suppressing the expansion and generation of CD8⁺ cytotoxic T-lymphocytes *in vitro*. They also observed that C57BL/6-into-BDF1 mice treated with kaempferol for 1 week after acute GVHD induction showed reduced GVHD-associated anti-host CTL activity, induction of Th2 cell development, and host repopulation with donor lymphocytes, resulting in early restoration of body weight and increased survival [303]. Cortés *et al.* found that kaempferol inhibited IL-4-induced STAT6 activation by specifically targeting the tyrosine kinase JAK3 [304]. The authors discussed that this inhibitory effects of kaempferol could be useful in controlling T cell-dependent responses mediated by JAK3-dependent cytokines, and that the specific inhibition of JAK3 would result in a T cell immune suppression that could be useful in lymphoproliferative diseases with JAK3 abnormalities and in transplant rejection [304].

4. PHARMACOKINETICS

Numerous *in vitro* studies have revealed that kaempferol has a wide range of biological activities. Some of these

activities, however, may not be relevant in an *in vivo* setting if this flavonoid does not reach the target tissues at specific concentrations for a particular period of time. In other words, a low bioavailability and/or a high metabolism of kaempferol may be an obstacle for kaempferol to induce some bioactivities *in vivo*. Since kaempferol is a common dietary constituent and since the oral route is the preferred route of administration for most drugs, it is important to understand the fate of this flavonoid in the body when taken orally.

The pharmacokinetics of kaempferol has been studied *in vitro* and *in vivo*, both in rats and humans. An overview of the pharmacokinetics of kaempferol is illustrated in Fig. (3). Flavonols such as kaempferol are commonly ingested as glycosides. It is recognized that the high polarity of glycosides hinders their absorption, whereas the intermediate polarity of aglycones facilitates it. Although this suggests that the absorption of glycosides needs the previous hydrolysis to absorbable aglycones, studies have shown that glycosides can be absorbed without hydrolysis [305-307]. Like other flavonoids, kaempferol is mainly absorbed in the small intestine. The lipophilicity of kaempferol facilitates its absorption by passive diffusion, but evidence suggests that it can also be absorbed by facilitated diffusion or active transport [308]. Kaempferol can be metabolized in the small intestine (to glucuronides and sulfoconjugates) by intestinal conjugation enzymes [308]. Like other flavonoids, kaempferol glycosides and kaempferol are extensively metabolized by the colon microflora. The colonic bacteria can both hydrolyze the glycosides to aglycones and break the C3 ring of aglycones to form simple phenolic compounds such as 4-hydroxyphenylacetic acid, phloroglucinol and 4-methylphenol, which can either be absorbed or excreted in feces [307, 309-314]. After absorption, kaempferol is extensively metabolized in the liver to form glucurono- and sulfo-conjugated forms [308, 315, 316]. These conjugated forms of kaempferol, some phenolic compounds produced by the colon microflora, kaempferol and some kaempferol glycosides can reach systemic circulation and tissues and are then excreted in urine [270, 317-323]. The percentage of kaempferol excreted in urine has been found to be 1.9% [318] and 2.5% [321] of the total amount of kaempferol ingested.

Several human studies have demonstrated that, after oral ingestion, kaempferol is detected in plasma at nanomolar concentrations. Eight healthy volunteers received endive as a source of kaempferol (246 mg kaempferol/kg endive) [318]. A portion of endive, which contained 8.65 mg kaempferol in the form of kaempferol-3-glucuronide (79%), kaempferol-3-glucoside (14%) and kaempferol-3-(6-malonyl)-glucoside (7%), led to a mean maximum plasma concentration of 100 nM 5.8 h after oral ingestion. Radket *et al.* [322] measured the plasma concentrations of several flavonoids in 48 healthy women and found that the mean intake of kaempferol was 4.7 mg/day and that the corresponding mean plasma concentration was 10.7 nM. Cao *et al.* [324] undertook a dietary intervention study and reported that an estimated intake of 14.97 mg kaempferol/day led to a plasma concentration of 57.86 nM. An intake of 27 mg kaempferol from tea has also been reported to lead to a plasma concentration of 15 ng/ml (52 nM) [321].

Some studies have found interconversions between kaempferol (3,5,7,4'-tetrahydroxyflavone) and the flavonoids galangin (3,5,7-trihydroxyflavone) and quercetin (3,5,7,3',4'-pentahydroxyflavone). Galangin can be transformed to kaempferol by rat cytochrome P450 (CYP) *via* hydroxylation of the B-ring at C3' [325]. Nielsen *et al.* [326] found that kaempferol was metabolized to quercetin and that this effect was probably mediated by the enzymes CYP1A. Similar findings were reported by Breinholt *et al.* [327]. These data suggest that the biological activities of kaempferol could be observed after galangin ingestion, and that some of the effects induced by kaempferol *in vivo* might be mediated in part by quercetin, which is known to have a wide range of biological activities [328, 329].

As discussed previously, rodent and human studies have shown that the concentrations of kaempferol achieved in plasma and tissues after oral ingestion are in the nanomolar range. Although kaempferol has shown biological activities in this range, most of the effects described in the literature come from *in vitro* studies in which kaempferol was tested at micromolar concentrations. This suggests that, outside of the gastrointestinal tract, these effects may not be relevant in an *in vivo* setting when kaempferol is taken orally. Many *in vivo* studies discussed in this review have found, however, that kaempferol has different biological activities outside of the gastrointestinal tract when administered orally (e.g. antioxidant, anti-inflammatory, analgesic, antidiabetic, anticancer, etc). For instance, a recent study by Huang *et al.* [121] found that kaempferol inhibited the growth and induced apoptosis in human osteosarcoma U-2 OS cells *in vitro*, with an IC₅₀ of approximately 150 μ M. The authors then observed that the oral administration of 25 or 50 mg/kg of kaempferol significantly reduced tumor weights and volumes in BALB/c (nu/nu) mice subcutaneously injected with U-2 OS cells [121]. Because the oral bioavailability of kaempferol is low, it is unlikely that the oral administration of kaempferol can lead to a concentration close to 150 μ M in the U-2 OS cancer cells placed under the skin of the mice. This suggests that the tumor growth inhibition observed in mice after the oral administration of kaempferol [121] may be caused by metabolites of kaempferol or by an unknown indirect effect. The low oral bioavailability of kaempferol may be overcome by using alternative routes of administration, such as the i.v. route [275].

5. SAFETY

There are conflicting reports regarding the safety of kaempferol. For example, although some studies have shown kaempferol to induce antimutagenic activity [114, 115, 330-332], other reports have revealed that this flavonoid may induce genotoxic effects [117, 333-341]. The prooxidant activity of kaempferol may play an important role in these genotoxic effects. Although flavonoids are known antioxidants, these compounds can also act as prooxidant agents under specific conditions [56]. By hydrogen atom donation, a flavonoid can reduce a free radical and be transformed in a phenoxyl radical. This phenoxyl radical may either react with a second radical becoming stable (antioxidant effect), or interact with oxygen resulting in the generation of ROS (prooxidant effect) [44]. The prooxidant effects of flavonoids have also been attributed to their

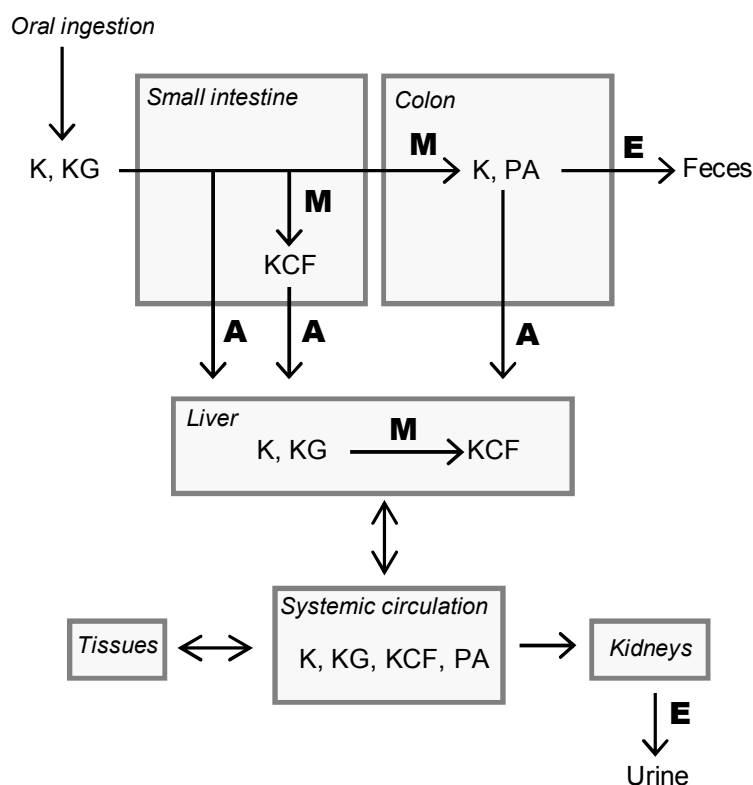


Fig. (3). Overview of the pharmacokinetics of kaempferol (see text for details). K: kaempferol; KG: kaempferol glycosides; KCF: kaempferol conjugated forms; PA: phenolic acids; A: absorption, M: metabolism; E: excretion.

capacity of reducing iron and copper ions; these reduced metals may participate in the formation of hydroxyl radicals (e.g. Fenton reaction) and in lipid peroxidation [56, 66, 339]. Although the prooxidant effects of flavonoids have also been attributed to their capacity to autooxidize when they are dissolved in aqueous buffers, kaempferol did not show this capacity in a study performed by Canada *et al.* [342]. The prooxidant activity of kaempferol may also be mediated by its ability to modulate the levels or activity of antioxidant and prooxidant enzymes [340, 343]. Some reports suggest that the mutagenicity of kaempferol may also be caused by its transformation, by CYP 1A1 enzymes, to the more potent genotoxicant quercetin [337, 338]. Although numerous *in vitro* studies suggest that kaempferol and quercetin may be genotoxic and carcinogenic, these effects have not been demonstrated in animal studies [344-346]. For instance, Takanashi *et al.* [346] observed that the oral administration of quercetin (0.1%) and kaempferol (0.04%) in the diet for 540 days did not increase the incidences of tumors in rats [346]. The low oral bioavailability of kaempferol may prevent this flavonoid from reaching genotoxic concentrations *in vivo*.

Several reports suggest that kaempferol may cause other unwanted effects under specific conditions. Kaempferol can react with iron and reduce its bioavailability [347] or may decrease the cellular uptake of folic acid [348]; these effects may cause problems in people with iron or folic acid deficiency. Kaempferol may also increase the bioavailability of the anticancer drug etoposide and increase its toxicity [349] or reduce the anticancer activity of

cisplatin [350]. It is important to mention that, like other antioxidant/prooxidant polyphenols [351], kaempferol can potentially increase or decrease the activity and toxicity of anticancer drugs with a prooxidant mechanism of action. At low concentrations, kaempferol may exert an antioxidant effect and reduce the activity of drugs that need to generate ROS to kill cancer cells efficiently. At high concentrations, kaempferol may generate ROS and increase the activity and toxicity of anticancer agents with a prooxidant mechanism. Because kaempferol has a low oral bioavailability, the ingestion of relatively high amounts of kaempferol leads to low concentrations of this flavonoid in plasma and tissues. These concentrations of kaempferol may cause antioxidant effects and can potentially reduce the activity of anticancer agents that need to generate ROS to exert their therapeutic effect.

6. CONCLUSIONS

Kaempferol is a flavonoid widely distributed in the plant kingdom. Numerous edible plants contain kaempferol, and it has been estimated that the human dietary intake of this polyphenol may be up to approximately 10 mg/day [28, 32, 352]. Epidemiological studies have found a positive association between the consumption of kaempferol-containing foods and a reduced risk of developing cardiovascular diseases and some types of cancer. Numerous *in vitro* and some animal studies support a role of kaempferol in the prevention and/or treatment of these and other diseases, such as neurodegenerative diseases, infectious diseases, diabetes, osteoporosis, anxiety, allergies,

inflammation and pain. However, many of these studies have been conducted at doses higher than those documented in humans and, therefore, it is difficult to predict from these results the effects of kaempferol intake on the prevention of these diseases. Likewise, most *in vitro* studies have been carried out at concentrations far beyond those achieved in plasma and tissues after the oral administration of kaempferol. This means that some biological effects induced by kaempferol *in vitro*, including some toxic effects, may not be relevant *in vivo* when this flavonoid is taken orally. The low oral bioavailability and high metabolism of kaempferol in humans should be overcome (e.g. by using alternative routes and forms of administration) to maximize some therapeutic properties of kaempferol. Additional animal studies and clinical trials are needed to better understand the possible health effects of kaempferol and to further evaluate its potential as a new drug.

REFERENCES

- [1] Hertog, M.G.; Feskens, E.J.; Hollman, P.C.; Katan, M.B.; Kromhout, D. Dietary antioxidant flavonoids and risk of coronary heart disease: the Zutphen Elderly Study. *Lancet*, **1993**, *342*, 1007-11.
- [2] Neuhouser, M.L. Dietary flavonoids and cancer risk: evidence from human population studies. *Nutr. Cancer*, **2004**, *50*, 1-7.
- [3] Le Marchand, L. Cancer preventive effects of flavonoids--a review. *Biomed. Pharmacother.*, **2002**, *56*, 296-301.
- [4] Maron, D.J. Flavonoids for reduction of atherosclerotic risk. *Curr. Atheroscler. Rep.*, **2004**, *6*, 73-8.
- [5] Garcia-Closas, R.; Gonzalez, C.A.; Agudo, A.; Riboli, E. Intake of specific carotenoids and flavonoids and the risk of gastric cancer in Spain. *Cancer Causes Control*, **1999**, *10*, 71-5.
- [6] Middleton E Jr; Kandaswami, C.; Theoharides, T.C. The effects of plant flavonoids on mammalian cells: implications for inflammation, heart disease, and cancer. *Pharmacol. Rev.*, **2000**, *52*, 673-751.
- [7] Wang, H.K. The therapeutic potential of flavonoids. *Expert. Opin. Investig. Drugs*, **2000**, *9*, 2103-19.
- [8] Lopez-Lazaro, M. Flavonoids as anticancer agents: structure-activity relationship study. *Curr. Med. Chem. Anticancer Agents*, **2002**, *2*, 691-714.
- [9] Li, Y.; Fang, H.; Xu, W. Recent advance in the research of flavonoids as anticancer agents. *Mini Rev. Med. Chem.*, **2007**, *7*, 663-78.
- [10] Lopez-Lazaro, M. Distribution and biological activities of the flavonoid luteolin. *Mini Rev. Med. Chem.*, **2009**, *9*, 31-59.
- [11] Ren, W.; Qiao, Z.; Wang, H.; Zhu, L.; Zhang, L. Flavonoids: promising anticancer agents. *Med. Res. Rev.*, **2003**, *23*, 519-34.
- [12] Hoensch, H.P. and Kirch, W. Potential role of flavonoids in the prevention of intestinal neoplasia: a review of their mode of action and their clinical perspectives. *Int. J. Gastrointest. Cancer*, **2005**, *35*, 187-95.
- [13] Winkel-Shirley, B. Flavonoid biosynthesis. A colorful model for genetics, biochemistry, cell biology, and biotechnology. *Plant Physiol.*, **2001**, *126*, 485-93.
- [14] Winkel-Shirley, B. Biosynthesis of flavonoids and effects of stress. *Curr. Opin. Plant Biol.*, **2002**, *5*, 218-23.
- [15] Yoon, K.D.; Jeong, D.G.; Hwang, Y.H.; Ryu, J.M.; Kim, J. Inhibitors of osteoclast differentiation from *Cephalotaxus koreana*. *J. Nat. Prod.*, **2007**, *70*, 2029-32.
- [16] Lee, H.S. and Kim, M.J. Selective responses of three *Ginkgo biloba* leaf-derived constituents on human intestinal bacteria. *J. Agric. Food Chem.*, **2002**, *50*, 1840-4.
- [17] Markham, K.R.; Geiger, H.; Jaggy, H. Kaempferol-3-O-glucosyl-(1-2)rhannoside from *Ginkgo biloba* and a reappraisal of other gluco(1-2, 1-3 and 1-4)rhannoside structures. **1992**, *31*, 1009-11.
- [18] Tang, Y.; Lou, F.; Wang, J.; Li, Y.; Zhuang, S. Coumaroyl flavonol glycosides from the leaves of *Ginkgo biloba*. *Phytochemistry*, **2001**, *58*, 1251-6.
- [19] Zheng, W. and Wang, S.Y. Antioxidant activity and phenolic compounds in selected herbs. *J. Agric. Food Chem.*, **2001**, *49*, 5165-70.
- [20] von Moltke, L.L.; Weemhoff, J.L.; Bedir, E.; Khan, I.A.; Harmatz, J.S.; Goldman, P.; Greenblatt, D.J. Inhibition of human cytochromes P450 by components of *Ginkgo biloba*. *J. Pharm. Pharmacol.*, **2004**, *56*, 1039-44.
- [21] Kwon, S.H.; Nam, J.I.; Kim, S.H.; Kim, J.H.; Yoon, J.H.; Kim, K.S. Kaempferol and quercetin, essential ingredients in *Ginkgo biloba* extract, inhibit interleukin-1beta-induced MUC5AC gene expression in human airway epithelial cells. *Phytother. Res.*, **2009**.
- [22] Krauze-Baranowska, M. Flavonoids from the genus *Taxus*. *Z. Naturforsch. [C. J.]*, **2004**, *59*, 43-7.
- [23] Yang, R.Y.; Lin, S.; Kuo, G. Content and distribution of flavonoids among 91 edible plant species. *Asia Pac. J. Clin. Nutr.*, **2008**, *17* Suppl 1, 275-9.
- [24] Franke, A.A.; Custer, L.J.; Arakaki, C.; Murphy, S.P. Vitamin C and flavonoid levels of fruits and vegetables consumed in Hawaii. *J. Food Compos. Anal.*, **2004**, *17*, 1-35.
- [25] Cui, Y.; Morgenstern, H.; Greenland, S.; Tashkin, D.P.; Mao, J.T.; Cai, L.; Cozen, W.; Mack, T.M.; Lu, Q.Y.; Zhang, Z.F. Dietary flavonoid intake and lung cancer-A population-based case-control study. *Cancer*, **2008**, *112*, 2241-8.
- [26] Garcia-Closas, R.; Agudo, A.; Gonzalez, C.A.; Riboli, E. Intake of specific carotenoids and flavonoids and the risk of lung cancer in women in Barcelona, Spain. *Nutr. Cancer*, **1998**, *32*, 154-8.
- [27] Nothlings, U.; Murphy, S.P.; Wilkens, L.R.; Henderson, B.E.; Kolonel, L.N. Flavonols and pancreatic cancer risk: the multiethnic cohort study. *Am. J. Epidemiol.*, **2007**, *166*, 924-31.
- [28] Gates, M.A.; Tworoger, S.S.; Hecht, J.L.; De, V., I.; Rosner, B.; Hankinson, S.E. A prospective study of dietary flavonoid intake and incidence of epithelial ovarian cancer. *Int. J. Cancer*, **2007**, *121*, 2225-32.
- [29] Gates, M.A.; Vitonis, A.F.; Tworoger, S.S.; Rosner, B.; Titus-Ernstoff, L.; Hankinson, S.E.; Cramer, D.W. Flavonoid intake and ovarian cancer risk in a population-based case-control study. *Int. J. Cancer*, **2009**, *124*, 1918-25.
- [30] Garcia, R.; Gonzalez, C.A.; Agudo, A.; Riboli, E. High intake of specific carotenoids and flavonoids does not reduce the risk of bladder cancer. *Nutr. Cancer*, **1999**, *35*, 212-4.
- [31] McCann, S.E.; Ambrosone, C.B.; Moysich, K.B.; Brasure, J.; Marshall, J.R.; Freudenheim, J.L.; Wilkinson, G.S.; Graham, S. Intakes of selected nutrients, foods, and phytochemicals and prostate cancer risk in western New York. *Nutr. Cancer*, **2005**, *53*, 33-41.
- [32] Adebamowo, C.A.; Cho, E.; Sampson, L.; Katan, M.B.; Spiegelman, D.; Willett, W.C.; Holmes, M.D. Dietary flavonols and flavonol-rich foods intake and the risk of breast cancer. *Int. J. Cancer*, **2005**, *20*:114, 628-33.
- [33] Lin, J.; Zhang, S.M.; Wu, K.; Willett, W.C.; Fuchs, C.S.; Giovannucci, E. Flavonoid intake and colorectal cancer risk in men and women. *Am. J. Epidemiol.*, **2006**, *164*, 644-51.
- [34] Geleijnse, J.M.; Launer, L.J.; Van der Kuip, D.A.; Hofman, A.; Witteman, J.C. Inverse association of tea and flavonoid intakes with incident myocardial infarction: the Rotterdam Study. *Am. J. Clin. Nutr.*, **2002**, *75*, 880-6.
- [35] Lin, J.; Rexrode, K.M.; Hu, F.; Albert, C.M.; Chae, C.U.; Rimm, E.B.; Stampfer, M.J.; Manson, J.E. Dietary intakes of flavonols and flavones and coronary heart disease in US women. *Am. J. Epidemiol.*, **2007**, *165*, 1305-13.
- [36] Marniemi, J.; Alanen, E.; Impivaara, O.; Seppanen, R.; Hakala, P.; Rajala, T.; Ronnema, T. Dietary and serum vitamins and minerals as predictors of myocardial infarction and stroke in elderly subjects. *Nutr. Metab. Cardiovasc. Dis.*, **2005**, *15*, 188-97.
- [37] Knekt, P.; Kumpulainen, J.; Jarvinen, R.; Rissanen, H.; Heliovaara, M.; Reunanen, A.; Hakulinen, T.; Aromaa, A. Flavonoid intake and risk of chronic diseases. *Am. J. Clin. Nutr.*, **2002**, *76*, 560-8.
- [38] Song, Y.; Manson, J.E.; Buring, J.E.; Sesso, H.D.; Liu, S. Associations of dietary flavonoids with risk of type 2 diabetes, and markers of insulin resistance and systemic inflammation in women: a prospective study and cross-sectional analysis. *J. Am. Coll. Nutr.*, **2005**, *24*, 376-84.
- [39] Droge, W. Free radicals in the physiological control of cell function. *Physiol Rev.*, **2002**, *82*, 47-95.

- [40] Finkel, T. Radical medicine: treating ageing to cure disease. *Nat. Rev. Mol. Cell Biol.*, **2005**, *6*, 971-6.
- [41] Valko, M.; Leibfritz, D.; Moncol, J.; Cronin, M.T.; Mazur, M.; Telser, J. Free radicals and antioxidants in normal physiological functions and human disease. *Int. J. Biochem. Cell Biol.*, **2007**, *39*, 44-84.
- [42] Rice-Evans, C. Flavonoid antioxidants. *Curr. Med. Chem.*, **2001**, *8*, 797-807.
- [43] Rice-Evans, C.A.; Miller, N.J.; Paganga, G. Structure-antioxidant activity relationships of flavonoids and phenolic acids. *Free Radic. Biol. Med.*, **1996**, *20*, 933-56.
- [44] Pietta, P.G. Flavonoids as antioxidants. *J. Nat. Prod.*, **2000**, *63*, 1035-42.
- [45] Kampkötter, A.; Gombitang, N.C.; Zurawski, R.F.; Timpel, C.; Chovolou, Y.; Watjen, W.; Kahl, R. Effects of the flavonoids kaempferol and fisetin on thermotolerance, oxidative stress and FoxO transcription factor DAF-16 in the model organism *Caenorhabditis elegans*. *Arch. Toxicol.*, **2007**, *81*, 849-58.
- [46] Bonina, F.; Puglia, C.; Ventura, D.; Aquino, R.; Tortora, S.; Sacchi, A.; Saija, A.; Tomaino, A.; Pellegrino, M.L.; de Caprariis, P. *In vitro* antioxidant and *in vivo* photoprotective effects of a lyophilized extract of *Capparis spinosa* L buds. *J. Cosmet. Sci.*, **2002**, *53*, 321-35.
- [47] Sanz, M.J.; Ferrandiz, M.L.; Cejudo, M.; Terencio, M.C.; Gil, B.; Bustos, G.; Ubeda, A.; Gunasegaran, R.; Alcaraz, M.J. Influence of a series of natural flavonoids on free radical generating systems and oxidative stress. *Xenobiotica*, **1994**, *24*, 689-99.
- [48] Hibatallah, J.; Carduner, C.; Poelman, M.C. In-vivo and in-vitro assessment of the free-radical-scavenger activity of Ginkgo flavone glycosides at high concentration. *J. Pharm. Pharmacol.*, **1999**, *51*, 1435-40.
- [49] Verma, A.R.; Vijayakumar, M.; Mathela, C.S.; Rao, C.V. *In vitro* and *in vivo* antioxidant properties of different fractions of Moringa oleifera leaves. *Food Chem. Toxicol.*, **2009**, *47*, 2196-201.
- [50] Aniya, Y.; Koyama, T.; Miyagi, C.; Miyahira, M.; Inomata, C.; Kinoshita, S.; Ichiba, T. Free radical scavenging and hepatoprotective actions of the medicinal herb, *Crassocephalum crepidioides* from the Okinawa Islands. *Biol. Pharm. Bull.*, **2005**, *28*, 19-23.
- [51] Wang, L.; Tu, Y.C.; Lian, T.W.; Hung, J.T.; Yen, J.H.; Wu, M.J. Distinctive antioxidant and antiinflammatory effects of flavonols. *J. Agric. Food Chem.*, **2006**, *54*, 9798-804.
- [52] Klaunig, J.E. and Kamendulis, L.M. The role of oxidative stress in carcinogenesis. *Annu. Rev. Pharmacol. Toxicol.*, **2004**, *44*, 239-67.
- [53] Heijnen, C.G.; Haenen, G.R.; van Acker, F.A.; van der Vijgh, W.J.; Bast, A. Flavonoids as peroxynitrite scavengers: the role of the hydroxyl groups. *Toxicol. In vitro*, **2001**, *15*, 3-6.
- [54] Nagao, A.; Seki, M.; Kobayashi, H. Inhibition of xanthine oxidase by flavonoids. *Biosci. Biotechnol. Biochem.*, **1999**, *63*, 1787-90.
- [55] Ozyurek, M.; Bektasoglu, B.; Guclu, K.; Apak, R. Measurement of xanthine oxidase inhibition activity of phenolics and flavonoids with a modified cupric reducing antioxidant capacity (CUPRAC) method. *Anal. Chim. Acta*, **2009**, *636*, 42-50.
- [56] Mira, L.; Fernandez, M.T.; Santos, M.; Rocha, R.; Florencio, M.H.; Jennings, K.R. Interactions of flavonoids with iron and copper ions: a mechanism for their antioxidant activity. *Free Radic. Res.*, **2002**, *36*, 1199-208.
- [57] Ren, J.; Meng, S.; Lekka, C.; Kaxiras, E. Complexation of flavonoids with iron: structure and optical signatures. *J. Phys. Chem. B.*, **2008**, *112*, 1845-50.
- [58] van Acker, S.A.; van den Berg, D.J.; Tromp, M.N.; Griffioen, D.H.; van Bennekom, W.P.; van der Vijgh, W.J.; Bast, A. Structural aspects of antioxidant activity of flavonoids. *Free Radic. Biol. Med.*, **1996**, *20*, 331-42.
- [59] Doronicheva, N.; Yasui, H.; Sakurai, H. Chemical structure-dependent differential effects of flavonoids on the catalase activity as evaluated by a chemiluminescent method. *Biol. Pharm. Bull.*, **2007**, *30*, 213-7.
- [60] Hong, J.T.; Yen, J.H.; Wang, L.; Lo, Y.H.; Chen, Z.T.; Wu, M.J. Regulation of heme oxygenase-1 expression and MAPK pathways in response to kaempferol and rhamnocitrin in PC12 cells. *Toxicol. Appl. Pharmacol.*, **2009**, *237*, 59-68.
- [61] Lin, H.Y.; Juan, S.H.; Shen, S.C.; Hsu, F.L.; Chen, Y.C. Inhibition of lipopolysaccharide-induced nitric oxide production by flavonoids in RAW264.7 macrophages involves heme oxygenase-1. *Biochem. Pharmacol.*, **2003**, *66*, 1821-32.
- [62] Maridonneau-Parini, I.; Braquet, P.; Garay, R.P. Heterogeneous effect of flavonoids on K⁺ loss and lipid peroxidation induced by oxygen-free radicals in human red cells. *Pharmacol. Res. Commun.*, **1986**, *18*, 61-72.
- [63] Ozgova, S.; Hermanek, J.; Gut, I. Different antioxidant effects of polyphenols on lipid peroxidation and hydroxyl radicals in the NADPH-, Fe-ascorbate- and Fe-microsomal systems. *Biochem. Pharmacol.*, **2003**, *66*, 1127-37.
- [64] Silva, M.M.; Santos, M.R.; Caroco, G.; Rocha, R.; Justino, G.; Mira, L. Structure-antioxidant activity relationships of flavonoids: a re-examination. *Free Radic. Res.*, **2002**, *36*, 1219-27.
- [65] Cos, P.; Calomme, M.; Sindambiwe, J.B.; De Bruyne, T.; Cimanga, K.; Pieters, L.; Vlietinck, A.J.; Vanden Berghe, D. Cytotoxicity and lipid peroxidation-inhibiting activity of flavonoids. *Planta Med.*, **2001**, *67*, 515-9.
- [66] Sugihara, N.; Arakawa, T.; Ohnishi, M.; Furuno, K. Anti- and pro-oxidative effects of flavonoids on metal-induced lipid hydroperoxide-dependent lipid peroxidation in cultured hepatocytes loaded with alpha-linolenic acid. *Free Radic. Biol. Med.*, **1999**, *27*, 1313-23.
- [67] Filipe, P.; Lanca, V.; Silva, J.N.; Morliere, P.; Santus, R.; Fernandes, A. Flavonoids and urate antioxidant interplay in plasma oxidative stress. *Mol. Cell Biochem.*, **2001**, *221*, 79-87.
- [68] Vareed, S.K.; Schutzki, R.E.; Nair, M.G. Lipid peroxidation, cyclooxygenase enzyme and tumor cell proliferation inhibitory compounds in *Cornus kousa* fruits. *Phytomedicine*, **2007**, *14*, 706-9.
- [69] Hou, L.; Zhou, B.; Yang, L.; Liu, Z.L. Inhibition of free radical initiated peroxidation of human erythrocyte ghosts by flavonols and their glycosides. *Org. Biomol. Chem.*, **2004**, *2*, 1419-23.
- [70] Tu, Y.C.; Lian, T.W.; Yen, J.H.; Chen, Z.T.; Wu, M.J. Antiatherogenic effects of kaempferol and rhamnocitrin. *J. Agric. Food Chem.*, **2007**, *55*, 9969-76.
- [71] Yu, J.; Wang, L.; Walzem, R.L.; Miller, E.G.; Pike, L.M.; Patil, B.S. Antioxidant activity of citrus limonoids, flavonoids, and coumarins. *J. Agric. Food Chem.*, **2005**, *53*, 2009-14.
- [72] Hou, L.; Zhou, B.; Yang, L.; Liu, Z.L. Inhibition of human low density lipoprotein oxidation by flavonols and their glycosides. *Chem. Phys. Lipids*, **2004**, *129*, 209-19.
- [73] Hirano, R.; Sasamoto, W.; Matsumoto, A.; Itakura, H.; Igarashi, O.; Kondo, K. Antioxidant ability of various flavonoids against DPPH radicals and LDL oxidation. *J. Nutr. Sci. Vitaminol. (Tokyo)*, **2001**, *47*, 357-62.
- [74] Brown, J.E.; Khodr, H.; Hider, R.C.; Rice-Evans, C.A. Structural dependence of flavonoid interactions with Cu²⁺ ions: implications for their antioxidant properties. *Biochem. J.*, **1998**, *330*, 1173-8.
- [75] Serhan, C.N.; Chiang, N.; Van Dyke, T.E. Resolving inflammation: dual anti-inflammatory and pro-resolution lipid mediators. *Nat. Rev. Immunol.*, **2008**, *8*, 349-61.
- [76] Kim, H.P.; Son, K.H.; Chang, H.W.; Kang, S.S. Anti-inflammatory plant flavonoids and cellular action mechanisms. *J. Pharmacol. Sci.*, **2004**, *96*, 229-45.
- [77] Park, M.J.; Lee, E.K.; Heo, H.S.; Kim, M.S.; Sung, B.; Kim, M.K.; Lee, J.; Kim, N.D.; Anton, S.; Choi, J.S.; Yu, B.P.; Chung, H.Y. The anti-inflammatory effect of kaempferol in aged kidney tissues: the involvement of nuclear factor-kappaB via nuclear factor-inducing kinase/IkappaB kinase and mitogen-activated protein kinase pathways. *J. Med. Food.*, **2009**, *12*, 351-8.
- [78] Kim, J.M.; Lee, E.K.; Kim, D.H.; Yu, B.P.; Chung, H.Y. Kaempferol modulates pro-inflammatory NF-kappaB activation by suppressing advanced glycation endproducts-induced NADPH oxidase. *Age (Dordr.)*, **2010**, *32*, 197-208.
- [79] De Melo, G.O.; Malvar, D.C.; Vanderlinde, F.A.; Rocha, F.F.; Pires, P.A.; Costa, E.A.; de Matos, L.G.; Kaiser, C.R.; Costa, S.S. Antinociceptive and anti-inflammatory kaempferol glycosides from *Sedum dendroideum*. *J. Ethnopharmacol.*, **2009**, *124*, 228-32.
- [80] Della, L.R.; Ragazzi, E.; Tubaro, A.; Fassina, G.; Vertua, R. Anti-inflammatory activity of benzopyrones that are inhibitors of cyclo-oxygenase. *Pharmacol. Res. Commun.*, **1988**, *20* Suppl 5, 91-4.
- [81] Gil, B.; Sanz, M.J.; Terencio, M.C.; Ferrandiz, M.L.; Bustos, G.; Paya, M.; Gunasegaran, R.; Alcaraz, M.J. Effects of flavonoids on

- Naja naja and human recombinant synovial phospholipases A2 and inflammatory responses in mice. *Life Sci.*, **1994**, *54*, L333-L338.
- [82] Innocenti, G.; Dall'Acqua, S.; Sosa, S.; Altinier, G.; Della, L.R. Topical anti-inflammatory activity of *Solenostemma argel* leaves. *J. Ethnopharmacol.*, **2005**, *102*, 307-10.
- [83] Kupeli, E.; Tatli, I.I.; Akdemir, Z.S.; Yesilada, E. Estimation of antinociceptive and anti-inflammatory activity on *Geranium pratense* subsp. *finitimum* and its phenolic compounds. *J. Ethnopharmacol.*, **2007**, *114*, 234-40.
- [84] Medeiros, K.C.; Faustino, L.; Borduchi, E.; Nascimento, R.J.; Silva, T.M.; Gomes, E.; Piuvezam, M.R.; Russo, M. Preventive and curative glycoside kaempferol treatments attenuate the TH2-driven allergic airway disease. *Int. Immunopharmacol.*, **2009**, *9*, 1540-8.
- [85] Orhan, I.; Kupeli, E.; Terzioglu, S.; Yesilada, E. Bioassay-guided isolation of kaempferol-3-O-beta-D-galactoside with anti-inflammatory and antinociceptive activity from the aerial part of *Calluna vulgaris* L. *J. Ethnopharmacol.*, **2007**, *114*, 32-7.
- [86] Toker, G.; Kupeli, E.; Memisoglu, M.; Yesilada, E. Flavonoids with antinociceptive and anti-inflammatory activities from the leaves of *Tilia argentea* (silver linden). *J. Ethnopharmacol.*, **2004**, *95*, 393-7.
- [87] Ahmed, M.S.; El Tanbouly, N.D.; Islam, W.T.; Sleem, A.A.; El Senousy, A.S. Antiinflammatory flavonoids from *Opuntia dillenii* (Ker-Gawl) Haw. flowers growing in Egypt. *Phytother. Res.*, **2005**, *19*, 807-9.
- [88] Dongmo, A.B.; Kamanyi, A.; Dzikouk, G.; Nkeh, B.C.; Tan, P.V.; Nguelefack, T.; Nole, T.; Bopellet, M.; Wagner, H. Anti-inflammatory and analgesic properties of the stem bark extract of *Mitragyna ciliata* (Rubiaceae) Aubrev. & Pellegr. *J. Ethnopharmacol.*, **2003**, *84*, 17-21.
- [89] Olszanecki, R.; Gebcka, A.; Kozlovski, V.I.; Gryglewski, R.J. Flavonoids and nitric oxide synthase. *J. Physiol Pharmacol.*, **2002**, *53*, 571-84.
- [90] Garcia-Mediavilla, V.; Crespo, I.; Collado, P.S.; Esteller, A.; Sanchez-Campos, S.; Tunon, M.J.; Gonzalez-Gallego, J. The anti-inflammatory flavones quercetin and kaempferol cause inhibition of inducible nitric oxide synthase, cyclooxygenase-2 and reactive C-protein, and down-regulation of the nuclear factor kappaB pathway in Chang Liver cells. *Eur. J. Pharmacol.*, **2007**, *557*, 221-9.
- [91] Jung, C.H.; Kim, J.H.; Hong, M.H.; Seog, H.M.; Oh, S.H.; Lee, P.J.; Kim, G.J.; Kim, H.M.; Um, J.Y.; Ko, S.G. Phenolic-rich fraction from *Rhus verniciflua* Stokes (RVS) suppress inflammatory response via NF-kappaB and JNK pathway in lipopolysaccharide-induced RAW 264.7 macrophages. *J. Ethnopharmacol.*, **2007**, *110*, 490-7.
- [92] Hamalainen, M.; Nieminen, R.; Vuorela, P.; Heinonen, M.; Moilanen, E. Anti-inflammatory effects of flavonoids: genistein, kaempferol, quercetin, and daidzein inhibit STAT-1 and NF-kappaB activations, whereas flavone, isorhamnetin, naringenin, and pelargonidin inhibit only NF-kappaB activation along with their inhibitory effect on iNOS expression and NO production in activated macrophages. *Mediators Inflamm.*, **2007**, *2007*, 45673-45673.
- [93] Lin, C.W.; Shen, S.C.; Chien, C.C.; Yang, L.Y.; Shia, L.T.; Chen, Y.C. 12-O-tetradecanoylphorbol-13-acetate-induced invasion/migration of glioblastoma cells through activating PKCalpha/ERK/NF-kappaB-dependent MMP-9 expression. *J. Cell Physiol.*, **2010**, *225*, 472-81.
- [94] Kowalski, J.; Samoiedny, A.; Paul, M.; Pietsch, G.; Wilczok, T. Effect of apigenin, kaempferol and resveratrol on the expression of interleukin-1beta and tumor necrosis factor-alpha genes in J774.2 macrophages. *Pharmacol. Rep.*, **2005**, *57*, 390-4.
- [95] Takano-Ishikawa, Y.; Goto, M.; Yamaki, K. Inhibitory effects of several flavonoids on E-selectin expression on human umbilical vein endothelial cells stimulated by tumor necrosis factor-alpha. *Phytother. Res.*, **2003**, *17*, 1224-7.
- [96] Chen, C.C.; Chow, M.P.; Huang, W.C.; Lin, Y.C.; Chang, Y.J. Flavonoids inhibit tumor necrosis factor-alpha-induced up-regulation of intercellular adhesion molecule-1 (ICAM-1) in respiratory epithelial cells through activator protein-1 and nuclear factor-kappaB: structure-activity relationships. *Mol. Pharmacol.*, **2004**, *66*, 683-93.
- [97] Kempuraj, D.; Madhappan, B.; Christodoulou, S.; Boucher, W.; Cao, J.; Papadopoulou, N.; Cetrulo, C.L.; Theoharides, T.C. Flavonols inhibit proinflammatory mediator release, intracellular calcium ion levels and protein kinase C theta phosphorylation in human mast cells. *Br. J. Pharmacol.*, **2005**, *145*, 934-44.
- [98] Kim, H.K.; Park, H.R.; Lee, J.S.; Chung, T.S.; Chung, H.Y.; Chung, J. Down-regulation of iNOS and TNF-alpha expression by kaempferol via NF-kappaB inactivation in aged rat gingival tissues. *Biogerontology*, **2007**, *8*, 399-408.
- [99] Rao, Y.K.; Fang, S.H.; Tzeng, Y.M. Antiinflammatory activities of flavonoids and a triterpene caffeate isolated from *Bauhinia variegata*. *Phytother. Res.*, **2008**, *22*, 957-62.
- [100] Lee, S.; Kim, Y.J.; Kwon, S.; Lee, Y.; Choi, S.Y.; Park, J.; Kwon, H.J. Inhibitory effects of flavonoids on TNF-alpha-induced IL-8 gene expression in HEK 293 cells. *BMB. Rep.*, **2009**, *42*, 265-70.
- [101] Gopalakrishnan, A.; Xu, C.J.; Nair, S.S.; Chen, C.; Hebbbar, V.; Kong, A.N. Modulation of activator protein-1 (AP-1) and MAPK pathway by flavonoids in human prostate cancer PC3 cells. *Arch. Pharm. Res.*, **2006**, *29*, 633-44.
- [102] Mahat, M.Y.; Kulkarni, N.M.; Vishwakarma, S.L.; Khan, F.R.; Thippeswamy, B.S.; Hebbballi, V.; Adhyapak, A.A.; Benade, V.S.; Ashfaq, S.M.; Tubachi, S.; Patil, B.M. Modulation of the cyclooxygenase pathway via inhibition of nitric oxide production contributes to the anti-inflammatory activity of kaempferol. *Eur. J. Pharmacol.*, **2010**, *642*, 169-76.
- [103] Mi, L.K.; Lee, K.W.; Jung, S.K.; Lee, E.J.; Heo, Y.S.; Bode, A.M.; Lubet, R.A.; Lee, H.J.; Dong, Z. Kaempferol inhibits UVB-induced COX-2 expression by suppressing Src kinase activity. *Biochem. Pharmacol.*, **2010**, *80*, 2042-9.
- [104] Kim, S.K.; Kim, H.J.; Choi, S.E.; Park, K.H.; Choi, H.K.; Lee, M.W. Anti-oxidative and inhibitory activities on nitric oxide (NO) and prostaglandin E2 (COX-2) production of flavonoids from seeds of *Prunus tomentosa* Thunberg. *Arch. Pharm. Res.*, **2008**, *31*, 424-8.
- [105] Liang, Y.C.; Huang, Y.T.; Tsai, S.H.; Lin-Shiau, S.Y.; Chen, C.F.; Lin, J.K. Suppression of inducible cyclooxygenase and inducible nitric oxide synthase by apigenin and related flavonoids in mouse macrophages. *Carcinogenesis*, **1999**, *20*, 1945-52.
- [106] Liang, Y.C.; Tsai, S.H.; Tsai, D.C.; Lin-Shiau, S.Y.; Lin, J.K. Suppression of inducible cyclooxygenase and nitric oxide synthase through activation of peroxisome proliferator-activated receptor-gamma by flavonoids in mouse macrophages. *FEBS Lett.*, **2001**, *496*, 12-8.
- [107] Mutoh, M.; Takahashi, M.; Fukuda, K.; Matsushima-Hibiya, Y.; Mutoh, H.; Sugimura, T.; Wakabayashi, K. Suppression of cyclooxygenase-2 promoter-dependent transcriptional activity in colon cancer cells by chemopreventive agents with a resorcin-type structure. *Carcinogenesis*, **2000**, *21*, 959-63.
- [108] Nakadate, T.; Yamamoto, S.; Aizu, E.; Kato, R. Effects of flavonoids and antioxidants on 12-O-tetradecanoyl-phorbol-13-acetate-caused epidermal ornithine decarboxylase induction and tumor promotion in relation to lipoxigenase inhibition by these compounds. *Gann.*, **1984**, *75*, 214-22.
- [109] Deng, S.; Palu, K.; West, B.J.; Su, C.X.; Zhou, B.N.; Jensen, J.C. Lipoxigenase inhibitory constituents of the fruits of noni (*Morinda citrifolia*) collected in Tahiti. *J. Nat. Prod.*, **2007**, *70*, 859-62.
- [110] Rostoka, E.; Baumane, L.; Isajevs, S.; Line, A.; Dzintare, M.; Svirina, D.; Sharipova, J.; Silina, K.; Kalvinsh, I.; Sjakste, N. Effects of Kaempferol and Myricetin on Inducible Nitric Oxide Synthase Expression and Nitric Oxide Production in Rats. *Basic Clin. Pharmacol. Toxicol.*, **2010**, *106*, 461-6.
- [111] Yasukawa, K.; Takido, M.; Takeuchi, M.; Sato, Y.; Nitta, K.; Nakagawa, S. Inhibitory effects of flavonol glycosides on 12-O-tetradecanoylphorbol-13-acetate-induced tumor promotion. *Chem. Pharm. Bull. (Tokyo)*, **1990**, *38*, 774-6.
- [112] Vogelstein, B. and Kinzler, K.W. Cancer genes and the pathways they control. *Nat. Med.*, **2004**, *10*, 789-99.
- [113] Cemeli, E.; Schmid, T.E.; Anderson, D. Modulation by flavonoids of DNA damage induced by estrogen-like compounds. *Environ. Mol. Mutagen.*, **2004**, *44*, 420-6.
- [114] Francis, A.R.; Shetty, T.K.; Bhattacharya, R.K. Modulating effect of plant flavonoids on the mutagenicity of N-methyl-N'-nitro-N-nitrosoguanidine. *Carcinogenesis*, **1989**, *10*, 1953-5.
- [115] Francis, A.R.; Shetty, T.K.; Bhattacharya, R.K. Modifying role of dietary factors on the mutagenicity of aflatoxin B1: *in vitro* effect of plant flavonoids. *Mutat. Res.*, **1989**, *222*, 393-401.

- [116] Anderson, D.; Basaran, N.; Dobrzynska, M.M.; Basaran, A.A.; Yu, T.W. Modulating effects of flavonoids on food mutagens in human blood and sperm samples in the comet assay. *Teratog. Carcinog. Mutagen.*, **1997**, *17*, 45-58.
- [117] Bestwick, C.S.; Milne, L.; Pirie, L.; Duthie, S.J. The effect of short-term kaempferol exposure on reactive oxygen levels and integrity of human (HL-60) leukaemic cells. *Biochim. Biophys. Acta*, **2005**, *1740*, 340-9.
- [118] Hanahan, D. and Weinberg, R.A. The hallmarks of cancer. *Cell*, **2000**, *100*, 57-70.
- [119] Hahn, W.C. and Weinberg, R.A. Rules for making human tumor cells. *N. Engl. J. Med.*, **2002**, *347*, 1593-603.
- [120] Brusselmans, K.; Vrolix, R.; Verhoeven, G.; Swinnen, J.V. Induction of cancer cell apoptosis by flavonoids is associated with their ability to inhibit fatty acid synthase activity. *J. Biol. Chem.*, **2005**, *280*, 5636-45.
- [121] Huang, W.W.; Chiu, Y.J.; Fan, M.J.; Lu, H.F.; Yeh, H.F.; Li, K.H.; Chen, P.Y.; Chung, J.G.; Yang, J.S. Kaempferol induced apoptosis via endoplasmic reticulum stress and mitochondria-dependent pathway in human osteosarcoma U-2 OS cells. *Mol. Nutr. Food Res.*, **2010**, *54*, 1585-95.
- [122] Kang, G.Y.; Lee, E.R.; Kim, J.H.; Jung, J.W.; Lim, J.; Kim, S.K.; Cho, S.G.; Kim, K.P. Downregulation of PLK-1 expression in kaempferol-induced apoptosis of MCF-7 cells. *Eur. J. Pharmacol.*, **2009**, *611*, 17-21.
- [123] Marfe, G.; Tafani, M.; Indelicato, M.; Sinibaldi-Salimei, P.; Reali, V.; Pucci, B.; Fini, M.; Russo, M.A. Kaempferol induces apoptosis in two different cell lines via Akt inactivation, Bax and SIRT3 activation, and mitochondrial dysfunction. *J. Cell Biochem.*, **2009**, *106*, 643-50.
- [124] Nguyen, T.T.; Tran, E.; Ong, C.K.; Lee, S.K.; Do, P.T.; Huynh, T.T.; Nguyen, T.H.; Lee, J.J.; Tan, Y.; Ong, C.S.; Huynh, H. Kaempferol-induced growth inhibition and apoptosis in A549 lung cancer cells is mediated by activation of MEK-MAPK. *J. Cell Physiol.*, **2003**, *197*, 110-21.
- [125] Sharma, V.; Joseph, C.; Ghosh, S.; Agarwal, A.; Mishra, M.K.; Sen, E. Kaempferol induces apoptosis in glioblastoma cells through oxidative stress. *Mol. Cancer Ther.*, **2007**, *6*, 2544-53.
- [126] Zhang, Y.; Chen, A.Y.; Li, M.; Chen, C.; Yao, Q. Ginkgo biloba extract kaempferol inhibits cell proliferation and induces apoptosis in pancreatic cancer cells. *J. Surg. Res.*, **2008**, *148*, 17-23.
- [127] Kim, B.W.; Lee, E.R.; Min, H.M.; Jeong, H.S.; Ahn, J.Y.; Kim, J.H.; Choi, H.Y.; Choi, H.; Kim, E.Y.; Park, S.P.; Cho, S.G. Sustained ERK activation is involved in the kaempferol-induced apoptosis of breast cancer cells and is more evident under 3-D culture condition. *Cancer Biol. Ther.*, **2008**, *7*, 1080-9.
- [128] Ahn, M.R.; Kunimasa, K.; Kumazawa, S.; Nakayama, T.; Kaji, K.; Uto, Y.; Hori, H.; Nagasawa, H.; Ohta, T. Correlation between antiangiogenic activity and antioxidant activity of various components from propolis. *Mol. Nutr. Food Res.*, **2009**, *53*, 643-51.
- [129] Schindler, R. and Mentlein, R. Flavonoids and vitamin E reduce the release of the angiogenic peptide vascular endothelial growth factor from human tumor cells. *J. Nutr.*, **2006**, *136*, 1477-82.
- [130] Kim, J.D.; Liu, L.; Guo, W.; Meydani, M. Chemical structure of flavonols in relation to modulation of angiogenesis and immune-endothelial cell adhesion. *J. Nutr. Biochem.*, **2006**, *17*, 165-76.
- [131] Christofori, G. New signals from the invasive front. *Nature*, **2006**, *441*, 444-50.
- [132] Phromnoi, K.; Yodkeeree, S.; Anuchapreeda, S.; Limtrakul, P. Inhibition of MMP-3 activity and invasion of the MDA-MB-231 human invasive breast carcinoma cell line by bioflavonoids. *Acta Pharmacol. Sin.*, **2009**, *30*, 1169-76.
- [133] Shen, S.C.; Lin, C.W.; Lee, H.M.; Chien, L.L.; Chen, Y.C. Lipopolysaccharide plus 12-o-tetradecanoylphorbol 13-acetate induction of migration and invasion of glioma cells *in vitro* and *in vivo*: Differential inhibitory effects of flavonoids. *Neuroscience*, **2006**, *140*, 477-89.
- [134] Labbe, D.; Provençal, M.; Lamy, S.; Boivin, D.; Gingras, D.; Beliveau, R. The flavonols quercetin, kaempferol, and myricetin inhibit hepatocyte growth factor-induced medulloblastoma cell migration. *J. Nutr.*, **2009**, *139*, 646-52.
- [135] Szatrowski, T.P. and Nathan, C.F. Production of large amounts of hydrogen peroxide by human tumor cells. *Cancer Res.*, **1991**, *51*, 794-8.
- [136] Suh, Y.A.; Arnold, R.S.; Lassegue, B.; Shi, J.; Xu, X.; Sorescu, D.; Chung, A.B.; Griendling, K.K.; Lambeth, J.D. Cell transformation by the superoxide-generating oxidase Mox1. *Nature*, **1999**, *401*, 79-82.
- [137] Arnold, R.S.; Shi, J.; Murad, E.; Whalen, A.M.; Sun, C.Q.; Polavarapu, R.; Parthasarathy, S.; Petros, J.A.; Lambeth, J.D. Hydrogen peroxide mediates the cell growth and transformation caused by the mitogenic oxidase Nox1. *Proc. Natl. Acad. Sci. U. S. A.*, **2001**, *98*, 5550-5.
- [138] Lopez-Lazaro, M. A new view of carcinogenesis and an alternative approach to cancer therapy. *Mol. Med.*, **2010**, *16*, 144-53.
- [139] Mantovani, A.; Allavena, P.; Sica, A.; Balkwill, F. Cancer-related inflammation. *Nature*, **2008**, *454*, 436-44.
- [140] Guengerich, F.P. and Shimada, T. Activation of procarcinogens by human cytochrome P450 enzymes. *Mutat. Res.*, **1998**, *400*, 201-13.
- [141] Chaudhary, A. and Willett, K.L. Inhibition of human cytochrome CYP 1 enzymes by flavonoids of St. John's wort. *Toxicology*, **2006**, *217*, 194-205.
- [142] Chang, T.K.; Chen, J.; Yeung, E.Y. Effect of Ginkgo biloba extract on procarcinogen-bioactivating human CYP1 enzymes: identification of isorhamnetin, kaempferol, and quercetin as potent inhibitors of CYP1B1. *Toxicol. Appl. Pharmacol.*, **2006**, *213*, 18-26.
- [143] Aboobaker, V.S.; Balgi, A.D.; Bhattacharya, R.K. *In vivo* effect of dietary factors on the molecular action of aflatoxin B1: role of non-nutrient phenolic compounds on the catalytic activity of liver fractions. *In vivo*, **1994**, *8*, 1095-8.
- [144] Puppala, D.; Gairola, C.G.; Swanson, H.I. Identification of kaempferol as an inhibitor of cigarette smoke-induced activation of the aryl hydrocarbon receptor and cell transformation. *Carcinogenesis*, **2007**, *28*, 639-47.
- [145] Mukai, R.; Satsu, H.; Shimizu, M.; Ashida, H. Inhibition of P-glycoprotein enhances the suppressive effect of kaempferol on transformation of the aryl hydrocarbon receptor. *Biosci. Biotechnol. Biochem.*, **2009**, *73*, 1635-9.
- [146] Mesia-Vela, S. and Kauffman, F.C. Inhibition of rat liver sulfotransferases SULT1A1 and SULT2A1 and glucuronosyltransferase by dietary flavonoids. *Xenobiotica*, **2003**, *33*, 1211-20.
- [147] Uda, Y.; Price, K.R.; Williamson, G.; Rhodes, M.J. Induction of the anticarcinogenic marker enzyme, quinone reductase, in murine hepatoma cells *in vitro* by flavonoids. *Cancer Lett.*, **1997**, *120*, 213-6.
- [148] Phang, J.M.; Poore, C.M.; Lopaczynska, J.; Yeh, G.C. Flavonol-stimulated efflux of 7,12-dimethylbenz(a)anthracene in multidrug-resistant breast cancer cells. *Cancer Res.*, **1993**, *53*, 5977-81.
- [149] Semenza, G.L. Development of novel therapeutic strategies that target HIF-1. *Expert. Opin. Ther. Targets.*, **2006**, *10*, 267-80.
- [150] Semenza, G.L. Life with oxygen. *Science*, **2007**, *318*, 62-4.
- [151] Semenza, G.L. Evaluation of HIF-1 inhibitors as anticancer agents. *Drug Discov. Today.*, **2007**, *12*, 853-9.
- [152] Zhong, H.; De Marzo, A.M.; Laughner, E.; Lim, M.; Hilton, D.A.; Zagzag, D.; Buechler, P.; Isaacs, W.B.; Semenza, G.L.; Simons, J.W. Overexpression of hypoxia-inducible factor 1alpha in common human cancers and their metastases. *Cancer Res.*, **1999**, *59*, 5830-5.
- [153] Lopez-Lazaro, M. Hypoxia-inducible factor 1 as a possible target for cancer chemoprevention. *Cancer Epidemiol. Biomarkers Prev.*, **2006**, *15*, 2332-5.
- [154] Mylonis, I.; Lakka, A.; Tsakalof, A.; Simos, G. The dietary flavonoid kaempferol effectively inhibits hif-1 activity and hepatoma cancer cell viability under hypoxic conditions. *Biochem. Biophys. Res. Commun.*, **2010**, *398*, 74-8.
- [155] Luo, H.; Rankin, G.O.; Liu, L.; Daddysman, M.K.; Jiang, B.H.; Chen, Y.C. Kaempferol inhibits angiogenesis and VEGF expression through both HIF dependent and independent pathways in human ovarian cancer cells. *Nutr. Cancer*, **2009**, *61*, 554-63.
- [156] Smith, J.A.; Poteet-Smith, C.E.; Xu, Y.; Errington, T.M.; Hecht, S.M.; Lannigan, D.A. Identification of the first specific inhibitor of p90 ribosomal S6 kinase (RSK) reveals an unexpected role for RSK in cancer cell proliferation. *Cancer Res.*, **2005**, *65*, 1027-34.
- [157] Clark, D.E.; Errington, T.M.; Smith, J.A.; Frierson, H.F., Jr.; Weber, M.J.; Lannigan, D.A. The serine/threonine protein kinase, p90 ribosomal S6 kinase, is an important regulator of prostate cancer cell proliferation. *Cancer Res.*, **2005**, *65*, 3108-16.

- [158] Cho, Y.Y.; Yao, K.; Kim, H.G.; Kang, B.S.; Zheng, D.; Bode, A.M.; Dong, Z. Ribosomal S6 kinase 2 is a key regulator in tumor promoter induced cell transformation. *Cancer Res.*, **2007**, *67*, 8104-12.
- [159] Cho, Y.Y.; Yao, K.; Pugliese, A.; Malakhova, M.L.; Bode, A.M.; Dong, Z. A regulatory mechanism for RSK2 NH(2)-terminal kinase activity. *Cancer Res.*, **2009**, *69*, 4398-406.
- [160] Harris, D.M.; Besselink, E.; Henning, S.M.; Go, V.L.; Heber, D. Phytoestrogens induce differential estrogen receptor alpha- or Beta-mediated responses in transfected breast cancer cells. *Exp. Biol. Med.*, **2005**, *230*, 558-68.
- [161] Oh, S.M.; Kim, Y.P.; Chung, K.H. Biphasic effects of kaempferol on the estrogenicity in human breast cancer cells. *Arch. Pharm. Res.*, **2006**, *29*, 354-62.
- [162] Tang, X.; Zhu, X.; Liu, S.; Nicholson, R.C.; Ni, X. Phytoestrogens induce differential estrogen receptor beta-mediated responses in transfected MG-63 cells. *Endocrine*, **2008**, *34*, 29-35.
- [163] Wang, J.; Fang, F.; Huang, Z.; Wang, Y.; Wong, C. Kaempferol is an estrogen-related receptor alpha and gamma inverse agonist. *FEBS Lett.*, **2009**.
- [164] Nakamura, Y.; Chang, C.C.; Mori, T.; Sato, K.; Ohtsuki, K.; Upham, B.L.; Trosko, J.E. Augmentation of differentiation and gap junction function by kaempferol in partially differentiated colon cancer cells. *Carcinogenesis*, **2005**, *26*, 665-71.
- [165] Kim, Y.K.; Kim, Y.S.; Choi, S.U.; Ryu, S.Y. Isolation of flavonol rhamnosides from *Loranthus tanakae* and cytotoxic effect of them on human tumor cell lines. *Arch. Pharm. Res.*, **2004**, *27*, 44-7.
- [166] Leung, H.W.; Lin, C.J.; Hour, M.J.; Yang, W.H.; Wang, M.Y.; Lee, H.Z. Kaempferol induces apoptosis in human lung non-small carcinoma cells accompanied by an induction of antioxidant enzymes. *Food Chem. Toxicol.*, **2007**, *45*, 2005-13.
- [167] Conforti, F.; Menichini, F.; Rigano, D.; Senatore, F. Antiproliferative activity on human cancer cell lines after treatment with polyphenolic compounds isolated from *Iris pseudopumila* flowers and rhizomes. *Z. Naturforsch. C.*, **2009**, *64*, 490-4.
- [168] Tomczyk, M.; Drozdowska, D.; Bielawska, A.; Bielawski, K.; Gudej, J. Human DNA topoisomerase inhibitors from *Potentilla argentea* and their cytotoxic effect against MCF-7. *Pharmazie*, **2008**, *63*, 389-93.
- [169] Ackland, M.L.; van de, W.S.; Jones, R. Synergistic antiproliferative action of the flavonols quercetin and kaempferol in cultured human cancer cell lines. *In Vivo*, **2005**, *19*, 69-76.
- [170] Li, W.; Du, B.; Wang, T.; Wang, S.; Zhang, J. Kaempferol induces apoptosis in human HCT116 colon cancer cells via the Ataxia-Telangiectasia Mutated-p53 pathway with the involvement of p53 Upregulated Modulator of Apoptosis. *Chem. Biol. Interact.*, **2009**, *177*, 121-7.
- [171] Chen, Y.C.; Shen, S.C.; Chow, J.M.; Ko, C.H.; Tseng, S.W. Flavone inhibition of tumor growth via apoptosis *in vitro* and *in vivo*. *Int. J. Oncol.*, **2004**, *25*, 661-70.
- [172] Knowles, L.M.; Zigrassi, D.A.; Tauber, R.A.; Hightower, C.; Milner, J.A. Flavonoids suppress androgen-independent human prostate tumor proliferation. *Nutr. Cancer*, **2000**, *38*, 116-22.
- [173] Lee, S.C.; Kuan, C.Y.; Yang, C.C.; Yang, S.D. Bioflavonoids commonly and potently induce tyrosine dephosphorylation/inactivation of oncogenic proline-directed protein kinase FA in human prostate carcinoma cells. *Anticancer Res.*, **1998**, *18*, 1117-21.
- [174] Li, N.; Liu, J.H.; Zhang, J.; Yu, B.Y. Comparative Evaluation of Cytotoxicity and Antioxidative Activity of 20 Flavonoids. *J. Agric. Food Chem.*, **2008**, *56*, 3876-83.
- [175] Wang, Y.F.; Cao, J.X.; Efferth, T.; Lai, G.F.; Luo, S.D. Cytotoxic and new tetralone derivatives from *Berberis floribunda* (Wall.) Brongn. *Chem. Biodivers.*, **2006**, *3*, 646-53.
- [176] Diaz, J.G.; Carmona, A.J.; Torres, F.; Quintana, J.; Estevez, F.; Herz, W. Cytotoxic activities of flavonoid glycoside acetates from *Consolida oliveriana*. *Planta Med.*, **2008**, *74*, 171-4.
- [177] Benyahia, S.; Benayache, S.; Benayache, F.; Quintana, J.; Lopez, M.; Leon, F.; Hernandez, J.C.; Estevez, F.; Bermejo, J. Isolation from *Eucalyptus occidentalis* and identification of a new kaempferol derivative that induces apoptosis in human myeloid leukemia cells. *J. Nat. Prod.*, **2004**, *67*, 527-31.
- [178] Dimas, K.; Demetzos, C.; Vaos, B.; Marselos, M.; Kokkinopoulos, D. Cytotoxic and antiproliferative effects of heptaacetyltiliroside on human leukemic cell lines. *Leuk. Res.*, **1999**, *23*, 1021-33.
- [179] Wang, I.K.; Lin-Shiau, S.Y.; Lin, J.K. Induction of apoptosis by apigenin and related flavonoids through cytochrome c release and activation of caspase-9 and caspase-3 in leukaemia HL-60 cells. *Eur. J. Cancer*, **1999**, *35*, 1517-25.
- [180] Alexandrakis, M.; Letourneau, R.; Kempuraj, D.; Kandere-Grzybowska, K.; Huang, M.; Christodoulou, S.; Boucher, W.; Seretakis, D.; Theoharides, T.C. Flavones inhibit proliferation and increase mediator content in human leukemic mast cells (HMC-1). *Eur. J. Haematol.*, **2003**, *71*, 448-54.
- [181] Dimas, K.; Demetzos, C.; Mitaku, S.; Marselos, M.; Tzavaras, T.; Kokkinopoulos, D. Cytotoxic activity of kaempferol glycosides against human leukaemic cell lines *in vitro*. *Pharmacol. Res.*, **2000**, *41*, 85-8.
- [182] Li, Y.L.; Gan, G.P.; Zhang, H.Z.; Wu, H.Z.; Li, C.L.; Huang, Y.P.; Liu, Y.W.; Liu, J.W. A flavonoid glycoside isolated from *Smilax china* L. rhizome *in vitro* anticancer effects on human cancer cell lines. *J. Ethnopharmacol.*, **2007**, *113*, 115-24.
- [183] Casagrande, F. and Darbon, J.M. Effects of structurally related flavonoids on cell cycle progression of human melanoma cells: regulation of cyclin-dependent kinases CDK2 and CDK1. *Biochem. Pharmacol.*, **2001**, *61*, 1205-15.
- [184] Zhang, Q.; Zhao, X.H.; Wang, Z.J. Flavones and flavonols exert cytotoxic effects on a human oesophageal adenocarcinoma cell line (OE33) by causing G2/M arrest and inducing apoptosis. *Food Chem. Toxicol.*, **2008**, *46*, 2042-53.
- [185] Zhang, Q.; Zhao, X.H.; Wang, Z.J. Cytotoxicity of flavones and flavonols to a human esophageal squamous cell carcinoma cell line (KYSE-510) by induction of G(2)/M arrest and apoptosis. *Toxicol. In vitro*, **2009**, *23*, 797-807.
- [186] Jeong, J.C.; Kim, M.S.; Kim, T.H.; Kim, Y.K. Kaempferol induces cell death through ERK and Akt-dependent down-regulation of XIAP and survivin in human glioma cells. *Neurochem. Res.*, **2009**, *34*, 991-1001.
- [187] Xu, W.; Liu, J.; Li, C.; Wu, H.Z.; Liu, Y.W. Kaempferol-7-O-beta-D-glucoside (KG) isolated from *Smilax china* L. rhizome induces G(2)/M phase arrest and apoptosis on HeLa cells in a p53-independent manner. *Cancer Lett.*, **2008**.
- [188] Luo, H.; Jiang, B.H.; King, S.M.; Chen, Y.C. Inhibition of cell growth and VEGF expression in ovarian cancer cells by flavonoids. *Nutr. Cancer*, **2008**, *60*, 800-9.
- [189] Luo, H.; Daddysman, M.K.; Rankin, G.O.; Jiang, B.H.; Chen, Y.C. Kaempferol enhances cisplatin's effect on ovarian cancer cells through promoting apoptosis caused by down regulation of cMyc. *Cancer Cell Int.*, **2010**, *10*, 16.
- [190] Yin, F.; Giuliano, A.E.; Van Herle, A.J. Growth inhibitory effects of flavonoids in human thyroid cancer cell lines. *Thyroid*, **1999**, *9*, 369-76.
- [191] Bestwick, C.S.; Milne, L.; Duthie, S.J. Kaempferol induced inhibition of HL-60 cell growth results from a heterogeneous response, dominated by cell cycle alterations. *Chem. Biol. Interact.*, **2007**, *20/170*, 76-85.
- [192] Filomeni, G.; Desideri, E.; Cardaci, S.; Graziani, I.; Piccirillo, S.; Rotilio, G.; Ciriolo, M.R. Carcinoma cells activate AMP-activated protein kinase-dependent autophagy as survival response to kaempferol-mediated energetic impairment. *Autophagy*, **2010**, *6*, 202-16.
- [193] Lupu, R. and Menendez, J.A. Pharmacological inhibitors of Fatty Acid Synthase (FASN)-catalyzed endogenous fatty acid biogenesis: a new family of anti-cancer agents? *Curr. Pharm. Biotechnol.*, **2006**, *7*, 483-93.
- [194] Boege, F.; Straub, T.; Kehr, A.; Boesenberg, C.; Christiansen, K.; Andersen, A.; Jakob, F.; Kohrle, J. Selected novel flavones inhibit the DNA binding or the DNA religation step of eukaryotic topoisomerase I. *J. Biol. Chem.*, **1996**, *271*, 2262-70.
- [195] Bandle, O.J.; Clawson, S.J.; Osheroff, N. Dietary polyphenols as topoisomerase II poisons: B ring and C ring substituents determine the mechanism of enzyme-mediated DNA cleavage enhancement. *Chem. Res. Toxicol.*, **2008**, *21*, 1253-60.
- [196] Constantinou, A.; Mehta, R.; Runyan, C.; Rao, K.; Vaughan, A.; Moon, R. Flavonoids as DNA topoisomerase antagonists and poisons: structure-activity relationships. *J. Nat. Prod.*, **1995**, *58*, 217-25.
- [197] Wang, S.; DeGroff, V.L.; Clinton, S.K. Tomato and soy polyphenols reduce insulin-like growth factor-I-stimulated rat prostate cancer cell proliferation and apoptotic resistance *in vitro*

- via inhibition of intracellular signaling pathways involving tyrosine kinase. *J. Nutr.*, **2003**, *133*, 2367-76.
- [198] Chen, D.; Daniel, K.G.; Chen, M.S.; Kuhn, D.J.; Landis-Piwowar, K.R.; Dou, Q.P. Dietary flavonoids as proteasome inhibitors and apoptosis inducers in human leukemia cells. *Biochem. Pharmacol.*, **2005**, *69*, 1421-32.
- [199] Renschler, M.F. The emerging role of reactive oxygen species in cancer therapy. *Eur. J. Cancer*, **2004**, *40*, 1934-40.
- [200] Wondrak, G.T. Redox-directed cancer therapeutics: molecular mechanisms and opportunities. *Antioxid. Redox. Signal.* **2009**, *11*, 3013-69
- [201] Lopez-Lazaro, M. Dual role of hydrogen peroxide in cancer: possible relevance to cancer chemoprevention and therapy. *Cancer Lett.*, **2007**, *252*, 1-8.
- [202] Nadova, S.; Miadokova, E.; Cipak, L. Flavonoids potentiate the efficacy of cytarabine through modulation of drug-induced apoptosis. *Neoplasma*, **2007**, *54*, 202-6.
- [203] Imai, Y.; Tsukahara, S.; Asada, S.; Sugimoto, Y. Phytoestrogens/flavonoids reverse breast cancer resistance protein/ABCG2-mediated multidrug resistance. *Cancer Res.*, **2004**, *64*, 4346-52.
- [204] Yoshida, T.; Konishi, M.; Horinaka, M.; Yasuda, T.; Goda, A.E.; Taniguchi, H.; Yano, K.; Wakada, M.; Sakai, T. Kaempferol sensitizes colon cancer cells to TRAIL-induced apoptosis. *Biochem. Biophys. Res. Commun.*, **2008**, *375*, 129-33.
- [205] Siegelin, M.D.; Reuss, D.E.; Habel, A.; Herold-Mende, C.; von Deimling, A. The flavonoid kaempferol sensitizes human glioma cells to TRAIL-mediated apoptosis by proteasomal degradation of survivin. *Mol. Cancer Ther.*, **2008**, *7*, 3566-74.
- [206] Harborne, J.B. and Williams, C.A. Advances in flavonoid research since 1992. *Phytochemistry*, **2000**, *55*, 481-504.
- [207] Cushnie, T.P. and Lamb, A.J. Antimicrobial activity of flavonoids. *Int. J. Antimicrob. Agents.*, **2005**, *26*, 343-56.
- [208] Kataoka, M.; Hirata, K.; Kunikata, T.; Ushio, S.; Iwaki, K.; Ohashi, K.; Ikeda, M.; Kurimoto, M. Antibacterial action of tryptanthrin and kaempferol, isolated from the indigo plant (*Polygonum tinctorium* Lour.), against *Helicobacter pylori*-infected Mongolian gerbils. *J. Gastroenterol.*, **2001**, *36*, 5-9.
- [209] Habbu, P.V.; Mahadevan, K.M.; Shastry, R.A.; Manjunatha, H. Antimicrobial activity of flavanoid sulphates and other fractions of *Argyrea speciosa* (Burm.f) Boj. *Indian J. Exp. Biol.*, **2009**, *47*, 121-8.
- [210] Lim, Y.H.; Kim, I.H.; Seo, J.J. *In vitro* activity of kaempferol isolated from the *Impatiens balsamina* alone and in combination with erythromycin or clindamycin against *Propionibacterium acnes*. *J. Microbiol.*, **2007**, *45*, 473-7.
- [211] Otsuka, N.; Liu, M.H.; Shiota, S.; Ogawa, W.; Kuroda, T.; Hatano, T.; Tsuchiya, T. Anti-methicillin resistant *Staphylococcus aureus* (MRSA) compounds isolated from *Laurus nobilis*. *Biol. Pharm. Bull.*, **2008**, *31*, 1794-7.
- [212] Xu, H.X. and Lee, S.F. Activity of plant flavonoids against antibiotic-resistant bacteria. *Phytother. Res.*, **2001**, *15*, 39-43.
- [213] Amoros, M.; Simoes, C.M.; Girre, L.; Sauvager, F.; Cormier, M. Synergistic effect of flavones and flavonols against herpes simplex virus type 1 in cell culture. Comparison with the antiviral activity of propolis. *J. Nat. Prod.*, **1992**, *55*, 1732-40.
- [214] Debiaggi, M.; Tateo, F.; Pagani, L.; Luini, M.; Romero, E. Effects of propolis flavonoids on virus infectivity and replication. *Microbiologica*, **1990**, *13*, 207-13.
- [215] Lyu, S.Y.; Rhim, J.Y.; Park, W.B. Antiherpetic activities of flavonoids against herpes simplex virus type 1 (HSV-1) and type 2 (HSV-2) *in vitro*. *Arch. Pharm. Res.*, **2005**, *28*, 1293-301.
- [216] Mitrocotsa, D.; Mitaku, S.; Axarlis, S.; Harvala, C.; Malamas, M. Evaluation of the antiviral activity of kaempferol and its glycosides against human cytomegalovirus. *Planta Med.*, **2000**, *66*, 377-9.
- [217] Jeong, H.J.; Ryu, Y.B.; Park, S.J.; Kim, J.H.; Kwon, H.J.; Kim, J.H.; Park, K.H.; Rho, M.C.; Lee, W.S. Neuraminidase inhibitory activities of flavonols isolated from *Rhodiola rosea* roots and their *in vitro* anti-influenza viral activities. *Bioorg. Med. Chem.*, **2009**, *17*, 6816-23.
- [218] Min, B.S.; Tomiyama, M.; Ma, C.M.; Nakamura, N.; Hattori, M. Kaempferol acetylramnosides from the rhizome of *Dryopteris crassirhizoma* and their inhibitory effects on three different activities of human immunodeficiency virus-1 reverse transcriptase. *Chem. Pharm. Bull. (Tokyo)*, **2001**, *49*, 546-50.
- [219] Mahmood, N.; Piacente, S.; Pizza, C.; Burke, A.; Khan, A.I.; Hay, A.J. The anti-HIV activity and mechanisms of action of pure compounds isolated from *Rosa damascena*. *Biochem. Biophys. Res. Commun.*, **1996**, *229*, 73-9.
- [220] Chu, S.C.; Hsieh, Y.S.; Lin, J.Y. Inhibitory effects of flavonoids on Moloney murine leukemia virus reverse transcriptase activity. *J. Nat. Prod.*, **1992**, *55*, 179-83.
- [221] Hwang, E.I.; Ahn, B.T.; Lee, H.B.; Kim, Y.K.; Lee, K.S.; Bok, S.H.; Kim, Y.T.; Kim, S.U. Inhibitory activity for chitin synthase II from *Saccharomyces cerevisiae* by tannins and related compounds. *Planta Med.*, **2001**, *67*, 501-4.
- [222] Yordanov, M.; Dimitrova, P.; Patkar, S.; Saso, L.; Ivanovska, N. Inhibition of *Candida albicans* extracellular enzyme activity by selected natural substances and their application in *Candida* infection. *Can. J. Microbiol.*, **2008**, *54*, 435-40.
- [223] Brunskole, M.; Zorko, K.; Kerbler, V.; Martens, S.; Stojan, J.; Gobec, S.; Lanisnik, R.T. Trihydroxynaphthalene reductase of *Curvularia lunata*-A target for flavonoid action? *Chem. Biol. Interact.*, **2009**, *178*, 259-67.
- [224] Ozelik, B.; Orhan, I.; Toker, G. Antiviral and antimicrobial assessment of some selected flavonoids. *Z. Naturforsch. [C.]*, **2006**, *61*, 632-8.
- [225] Barbosa, E.; Calzada, F.; Campos, R. *In vivo* anti-giardial activity of three flavonoids isolated of some medicinal plants used in Mexican traditional medicine for the treatment of diarrhea. *J. Ethnopharmacol.*, **2007**, *109*, 552-4.
- [226] Calzada, F. Additional antiprotozoal constituents from *Cuphea pinetorum*, a plant used in Mayan traditional medicine to treat diarrhoea. *Phytother. Res.*, **2005**, *19*, 725-7.
- [227] Calzada, F.; Meckes, M.; Cedillo-Rivera, R. Antiamoebic and anti-giardial activity of plant flavonoids. *Planta Med.*, **1999**, *65*, 78-80.
- [228] Meckes, M.; Calzada, F.; Tapia-Contreras, A.; Cedillo-Rivera, R. Antiprotozoal properties of *Helianthemum glomeratum*. *Phytother. Res.*, **1999**, *13*, 102-5.
- [229] Marin, C.; Boutaleb-Charki, S.; Diaz, J.G.; Huertas, O.; Rosales, M.J.; Perez-Cordon, G.; Guitierrez-Sanchez, R.; Sanchez-Moreno, M. Antileishmaniasis activity of flavonoids from *Consolida oliveriana*. *J. Nat. Prod.*, **2009**, *72*, 1069-74.
- [230] Muzitano, M.F.; Tinoco, L.W.; Guette, C.; Kaiser, C.R.; Rossi-Bergmann, B.; Costa, S.S. The antileishmanial activity assessment of unusual flavonoids from *Kalanchoe pinnata*. *Phytochemistry*, **2006**, *67*, 2071-7.
- [231] Dhalla, N.S.; Temsah, R.M.; Neticadan, T. Role of oxidative stress in cardiovascular diseases. *J. Hypertens.*, **2000**, *18*, 655-73.
- [232] Willerson, J.T. and Ridker, P.M. Inflammation as a cardiovascular risk factor. *Circulation*, **2004**, *109*, II2-10.
- [233] Oh, H.; Kang, D.G.; Kwon, J.W.; Kwon, T.O.; Lee, S.Y.; Lee, D.B.; Lee, H.S. Isolation of angiotensin converting enzyme (ACE) inhibitory flavonoids from *Sedum sarmentosum*. *Biol. Pharm. Bull.*, **2004**, *27*, 2035-7.
- [234] Loizzo, M.R.; Said, A.; Tundis, R.; Rashed, K.; Statti, G.A.; Hufner, A.; Menichini, F. Inhibition of angiotensin converting enzyme (ACE) by flavonoids isolated from *Ailanthus excelsa* (Roxb) (Simaroubaceae). *Phytother. Res.*, **2007**, *21*, 32-6.
- [235] Kameda, K.; Takaku, T.; Okuda, H.; Kimura, Y.; Okuda, T.; Hatano, T.; Agata, I.; Arichi, S. Inhibitory effects of various flavonoids isolated from leaves of persimmon on angiotensin-converting enzyme activity. *J. Nat. Prod.*, **1987**, *50*, 680-3.
- [236] Olszanecki, R.; Bujak-Gizycka, B.; Madej, J.; Suski, M.; Wolkow, P.P.; Jawien, J.; Korbut, R. Kaempferol, but not resveratrol inhibits angiotensin converting enzyme. *J. Physiol. Pharmacol.*, **2008**, *59*, 387-92.
- [237] Xu, Y.C.; Yeung, D.K.; Man, R.Y.; Leung, S.W. Kaempferol enhances endothelium-independent and dependent relaxation in the porcine coronary artery. *Mol. Cell Biochem.*, **2006**, *287*, 61-7.
- [238] Padilla, E.; Ruiz, E.; Redondo, S.; Gordillo-Moscoco, A.; Slowing, K.; Tejerina, T. Relationship between vasodilation capacity and phenolic content of Spanish wines. *Eur. J. Pharmacol.*, **2005**, *517*, 84-91.
- [239] Xu, Y.C.; Leung, G.P.; Wong, P.Y.; Vanhoutte, P.M.; Man, R.Y. Kaempferol stimulates large conductance Ca²⁺-activated K⁺ (BKCa) channels in human umbilical vein endothelial cells via a cAMP/PKA-dependent pathway. *Br. J. Pharmacol.*, **2008**, *154*, 1247-53.

- [240] Chung, M.I.; Gan, K.H.; Lin, C.N.; Ko, F.N.; Teng, C.M. Antiplatelet effects and vasorelaxing action of some constituents of Formosan plants. *J. Nat. Prod.*, **1993**, *56*, 929-34.
- [241] Hannum, S.M. Potential impact of strawberries on human health: a review of the science. *Crit Rev. Food Sci. Nutr.*, **2004**, *44*, 1-17.
- [242] Belguith-Hadriche, O.; Bouaziz, M.; Jamoussi, K.; El Feki, A.; Sayadi, S.; Makni-Ayedi, F. Lipid-lowering and antioxidant effects of an ethyl acetate extract of fenugreek seeds in high-cholesterol-fed rats. *J. Agric. Food Chem.*, **2010**, *58*, 2116-22.
- [243] Zern, T.L.; West, K.L.; Fernandez, M.L. Grape polyphenols decrease plasma triglycerides and cholesterol accumulation in the aorta of ovariectomized guinea pigs. *J. Nutr.*, **2003**, *133*, 2268-72.
- [244] Yu, S.F.; Shun, C.T.; Chen, T.M.; Chen, Y.H. 3-O-beta-D-glucosyl-(1->6)-beta-D-glucosyl-kaempferol isolated from *Sauropus androgenus* reduces body weight gain in Wistar rats. *Biol. Pharm. Bull.*, **2006**, *29*, 2510-3.
- [245] Chen, Q.C.; Zhang, W.Y.; Jin, W.; Lee, I.S.; Min, B.S.; Jung, H.J.; Na, M.; Lee, S.; Bae, K. Flavonoids and isoflavonoids from *Sophorae Flos* improve glucose uptake *in vitro*. *Planta Med.*, **2010**, *76*, 79-81.
- [246] de Sousa, E.; Zanatta, L.; Seifriz, I.; Creczynski-Pasa, T.B.; Pizzolatti, M.G.; Szpoganicz, B.; Silva, F.R. Hypoglycemic effect and antioxidant potential of kaempferol-3,7-O-(alpha)-dirhamnoside from *Bauhinia forficata* leaves. *J. Nat. Prod.*, **2004**, *67*, 829-32.
- [247] Jorge, A.P.; Horst, H.; de Sousa, E.; Pizzolatti, M.G.; Silva, F.R. Insulinomimetic effects of kaempferitrin on glycaemia and on 14C-glucose uptake in rat soleus muscle. *Chem. Biol. Interact.*, **2004**, *149*, 89-96.
- [248] Andrade, C.A.; Wiedenfeld, H.; Revilla, M.C.; Sergio, I.A. Hypoglycemic effect of *Equisetum myriochaetum* aerial parts on streptozotocin diabetic rats. *J. Ethnopharmacol.*, **2000**, *72*, 129-33.
- [249] Revilla, M.C.; Andrade-Cetto, A.; Islas, S.; Wiedenfeld, H. Hypoglycemic effect of *Equisetum myriochaetum* aerial parts on type 2 diabetic patients. *J. Ethnopharmacol.*, **2002**, *81*, 117-20.
- [250] Cunha, W.R.; Arantes, G.M.; Ferreira, D.S.; Lucarini, R.; Silva, M.L.; Furtado, N.A.; Silva Filho, A.A.; Crotti, A.E.; Araujo, A.R. Hypoglycemic effect of *Leandra lacunosa* in normal and alloxan-induced diabetic rats. *Fitoterapia*, **2008**, *79*, 356-60.
- [251] De Souza Schmidt Goncalves AE; Lajolo, F.M.; Genovese, M.I. Chemical composition and antioxidant/antidiabetic potential of Brazilian native fruits and commercial frozen pulps. *J. Agric. Food Chem.*, **2010**, *58*, 4666-74.
- [252] Anwar, F.; Latif, S.; Ashraf, M.; Gilani, A.H. *Moringa oleifera*: a food plant with multiple medicinal uses. *Phytother. Res.*, **2007**, *21*, 17-25.
- [253] Basnet, P.; Kadota, S.; Terashima, S.; Shimizu, M.; Namba, T. Two new 2-arylbenzofuran derivatives from hypoglycemic activity-bearing fractions of *Morus insignis*. *Chem. Pharm. Bull. (Tokyo)*, **1993**, *41*, 1238-43.
- [254] Lee, M.J.; Rao, Y.K.; Chen, K.; Lee, Y.C.; Tzeng, Y.M. Effect of flavonol glycosides from *Cinnamomum osmophloeum* leaves on adiponectin secretion and phosphorylation of insulin receptor-beta in 3T3-L1 adipocytes. *J. Ethnopharmacol.*, **2009**, *126*, 79-85.
- [255] Asgary, S.; Naderi, G.A.; Zadegan, N.S.; Vakili, R. The inhibitory effects of pure flavonoids on *in vitro* protein glycosylation. *J. Herb. Pharmacother.*, **2002**, *2*, 47-55.
- [256] Ghaffari, M.A. and Mojab, S. Influence of flavonols as *in vitro* on low density lipoprotein glycation. *Iran Biomed. J.*, **2007**, *11*, 185-91.
- [257] Cazarolli, L.H.; Zanatta, L.; Jorge, A.P.; de Sousa, E.; Horst, H.; Woehl, V.M.; Pizzolatti, M.G.; Szpoganicz, B.; Silva, F.R. Follow-up studies on glycosylated flavonoids and their complexes with vanadium: their anti-hyperglycemic potential role in diabetes. *Chem. Biol. Interact.*, **2006**, *163*, 177-91.
- [258] Cazarolli, L.H.; Folador, P.; Pizzolatti, M.G.; Mena Barreto Silva, F.R. Signaling pathways of kaempferol-3-neohesperidoside in glycogen synthesis in rat soleus muscle. *Biochimie*, **2009**, *91*, 843-9.
- [259] Zanatta, L.; Rosso, A.; Folador, P.; Figueiredo, M.S.; Pizzolatti, M.G.; Leite, L.D.; Silva, F.R. Insulinomimetic effect of kaempferol 3-neohesperidoside on the rat soleus muscle. *J. Nat. Prod.*, **2008**, *71*, 532-5.
- [260] Fang, X.K.; Gao, J.; Zhu, D.N. Kaempferol and quercetin isolated from *Euonymus alatus* improve glucose uptake of 3T3-L1 cells without adipogenesis activity. *Life Sci.*, **2008**, *82*, 615-22.
- [261] Lee, Y.J.; Suh, K.S.; Choi, M.C.; Chon, S.; Oh, S.; Woo, J.T.; Kim, S.W.; Kim, J.W.; Kim, Y.S. Kaempferol protects HIT-T15 pancreatic beta cells from 2-deoxy-D-ribose-induced oxidative damage. *Phytother. Res.*, **2010**, *24*, 419-23.
- [262] Harris, D.M.; Besselink, E.; Henning, S.M.; Go, V.L.; Heber, D. Phytoestrogens induce differential estrogen receptor alpha- or Beta-mediated responses in transfected breast cancer cells. *Exp. Biol. Med.*, **2005**, *230*, 558-68.
- [263] Oh, S.M. and Chung, K.H. Estrogenic activities of Ginkgo biloba extracts. *Life Sci.*, **2004**, *74*, 1325-35.
- [264] Sathyamoorthy, N.; Wang, T.T.; Phang, J.M. Stimulation of pS2 expression by diet-derived compounds. *Cancer Res.*, **1994**, *54*, 957-61.
- [265] Kuiper, G.G.; Lemmen, J.G.; Carlsson, B.; Corton, J.C.; Safe, S.H.; van der Saag, P.T.; van der, B.B.; Gustafsson, J.A. Interaction of estrogenic chemicals and phytoestrogens with estrogen receptor beta. *Endocrinology*, **1998**, *139*, 4252-63.
- [266] Breinholt, V. and Larsen, J.C. Detection of weak estrogenic flavonoids using a recombinant yeast strain and a modified MCF7 cell proliferation assay. *Chem. Res. Toxicol.*, **1998**, *11*, 622-9.
- [267] Stroheker, T.; Chagnon, M.C.; Pinnert, M.F.; Berges, R.; Canivenc-Lavie, M.C. Estrogenic effects of food wrap packaging xenostrogens and flavonoids in female Wistar rats: a comparative study. *Reprod. Toxicol.*, **2003**, *17*, 421-32.
- [268] Breinholt, V.; Hossaini, A.; Svendsen, G.W.; Brouwer, C.; Nielsen, E. Estrogenic activity of flavonoids in mice. The importance of estrogen receptor distribution, metabolism and bioavailability. *Food Chem. Toxicol.*, **2000**, *38*, 555-64.
- [269] Wattel, A.; Kamel, S.; Mentaverri, R.; Lorget, F.; Prouillet, C.; Petit, J.P.; Fardelonne, P.; Brazier, M. Potent inhibitory effect of naturally occurring flavonoids quercetin and kaempferol on *in vitro* osteoclastic bone resorption. *Biochem. Pharmacol.*, **2003**, *65*, 35-42.
- [270] Trivedi, R.; Kumar, S.; Kumar, A.; Siddiqui, J.A.; Swarnkar, G.; Gupta, V.; Kendurker, A.; Dwivedi, A.K.; Romero, J.R.; Chattopadhyay, N. Kaempferol has osteogenic effect in ovariectomized adult Sprague-Dawley rats. *Mol. Cell Endocrinol.*, **2008**, *289*, 85-93.
- [271] Trivedi, R.; Kumar, A.; Gupta, V.; Kumar, S.; Nagar, G.K.; Romero, J.R.; Dwivedi, A.K.; Chattopadhyay, N. Effects of Egb 761 on bone mineral density, bone microstructure, and osteoblast function: Possible roles of quercetin and kaempferol. *Mol. Cell Endocrinol.*, **2009**, *302*, 86-91.
- [272] Kumar, A.; Singh, A.K.; Gautam, A.K.; Chandra, D.; Singh, D.; Changkija, B.; Singh, M.P.; Trivedi, R. Identification of kaempferol-regulated proteins in rat calvarial osteoblasts during mineralization by proteomics. *Proteomics*, **2010**, *10*, 1730-9.
- [273] Prouillet, C.; Maziere, J.C.; Maziere, C.; Wattel, A.; Brazier, M.; Kamel, S. Stimulatory effect of naturally occurring flavonols quercetin and kaempferol on alkaline phosphatase activity in MG-63 human osteoblasts through ERK and estrogen receptor pathway. *Biochem. Pharmacol.*, **2004**, *67*, 1307-13.
- [274] Pang, J.L.; Ricupero, D.A.; Huang, S.; Fatma, N.; Singh, D.P.; Romero, J.R.; Chattopadhyay, N. Differential activity of kaempferol and quercetin in attenuating tumor necrosis factor receptor family signaling in bone cells. *Biochem. Pharmacol.*, **2006**, *71*, 818-26.
- [275] Lopez-Sanchez, C.; Martin-Romero, F.J.; Sun, F.; Luis, L.; Samhan-Arias, A.K.; Garcia-Martinez, V.; Gutierrez-Merino, C. Blood micromolar concentrations of kaempferol afford protection against ischemia/reperfusion-induced damage in rat brain. *Brain Res.*, **2007**, *1182*, 123-37.
- [276] Filomeni, G.; Graziani, I.; De Zio, D.; Dini, L.; Centonze, D.; Rotilio, G.; Ciriolo, M.R. Neuroprotection of kaempferol by autophagy in models of rotenone-mediated acute toxicity: possible implications for Parkinson's disease. *Neurobiol. Aging.*, [Epub ahead of print]
- [277] Lagoa, R.; Lopez-Sanchez, C.; Samhan-Arias, A.K.; Ganan, C.M.; Garcia-Martinez, V.; Gutierrez-Merino, C. Kaempferol protects against rat striatal degeneration induced by 3-nitropropionic acid. *J. Neurochem.*, **2009**, *111*, 473-87.

- [278] Smith, J.V. and Luo, Y. Elevation of oxidative free radicals in Alzheimer's disease models can be attenuated by Ginkgo biloba extract EGb 761. *J. Alzheimers. Dis.*, **2003**, *5*, 287-300.
- [279] Wang, C.N.; Chi, C.W.; Lin, Y.L.; Chen, C.F.; Shiao, Y.J. The neuroprotective effects of phytoestrogens on amyloid beta protein-induced toxicity are mediated by abrogating the activation of caspase cascade in rat cortical neurons. *J. Biol. Chem.*, **2001**, *276*, 5287-95.
- [280] Roth, A.; Schaffner, W.; Hertel, C. Phytoestrogen kaempferol (3,4',5,7-tetrahydroxyflavone) protects PC12 and T47D cells from beta-amyloid-induced toxicity. *J. Neurosci. Res.*, **1999**, *57*, 399-404.
- [281] Kim, Y.; Park, E.J.; Kim, J.; Kim, Y.; Kim, S.R.; Kim, Y.Y. Neuroprotective constituents from *Hedyotis diffusa*. *J. Nat. Prod.*, **2001**, *64*, 75-8.
- [282] Liu, R.N.; Wang, W.; Ding, Y.; Xie, W.D.; Ma, C.; Du, L.J. A new flavonol glycoside and activity of compounds from the flower of *Nymphaea candida*. *J. Asian Nat. Prod. Res.*, **2007**, *9*, 333-8.
- [283] Silva, B.; Oliveira, P.J.; Dias, A.; Malva, J.O. Quercetin, kaempferol and biapigenin from *Hypericum perforatum* are neuroprotective against excitotoxic insults. *Neurotox. Res.*, **2008**, *13*, 265-79.
- [284] Samhan-Arias, A.K.; Martin-Romero, F.J.; Gutierrez-Merino, C. Kaempferol blocks oxidative stress in cerebellar granule cells and reveals a key role for reactive oxygen species production at the plasma membrane in the commitment to apoptosis. *Free Radic. Biol. Med.*, **2004**, *37*, 48-61.
- [285] Khaengkhan, P.; Nishikaze, Y.; Niidome, T.; Kanaori, K.; Tajima, K.; Ichida, M.; Harada, S.; Sugimoto, H.; Kamei, K. Identification of an anti-amyloidogenic substance from mulberry leaves. *Neuroreport*, **2009**, *20*, 1214-8.
- [286] Ono, K.; Yoshiike, Y.; Takashima, A.; Hasegawa, K.; Naiki, H.; Yamada, M. Potent anti-amyloidogenic and fibril-destabilizing effects of polyphenols *in vitro*: implications for the prevention and therapeutics of Alzheimer's disease. *J. Neurochem.*, **2003**, *87*, 172-81.
- [287] Hou, Y.; Aboukhatwa, M.A.; Lei, D.L.; Manaye, K.; Khan, I.; Luo, Y. Anti-depressant natural flavonols modulate BDNF and beta amyloid in neurons and hippocampus of double TgAD mice. *Neuropharmacology*, **2010**, *58*, 911-20.
- [288] Herrera-Ruiz, M.; Roman-Ramos, R.; Zamilpa, A.; Tortoriello, J.; Jimenez-Ferrer, J.E. Flavonoids from *Tilia americana* with anxiolytic activity in plus-maze test. *J. Ethnopharmacol.*, **2008**, *118*, 312-7.
- [289] Aguirre-Hernandez, E.; Gonzalez-Trujano, M.E.; Martinez, A.L.; Moreno, J.; Kite, G.; Terrazas, T.; Soto-Hernandez, M. HPLC/MS analysis and anxiolytic-like effect of quercetin and kaempferol flavonoids from *Tilia americana* var. *mexicana*. *J. Ethnopharmacol.*, **2010**, *127*, 91-7.
- [290] Grundmann, O.; Nakajima, J.; Kamata, K.; Seo, S.; Butterweck, V. Kaempferol from the leaves of *Apocynum venetum* possesses anxiolytic activities in the elevated plus maze test in mice. *Phytomedicine*, **2009**, *16*, 295-302.
- [291] Perez-Ortega, G.; Guevara-Fefer, P.; Chavez, M.; Herrera, J.; Martinez, A.; Martinez, A.L.; Gonzalez-Trujano, M.E. Sedative and anxiolytic efficacy of *Tilia americana* var. *mexicana* inflorescences used traditionally by communities of State of Michoacan, Mexico. *J. Ethnopharmacol.*, **2008**, *116*, 461-8.
- [292] Loscalzo, L.M.; Wasowski, C.; Marder, M. Neuroactive flavonoid glycosides from *Tilia petiolaris* DC. extracts. *Phytother. Res.*, **2009**, *23*, 1453-7.
- [293] Amellal, M.; Bronner, C.; Briancon, F.; Haag, M.; Anton, R.; Landry, Y. Inhibition of mast cell histamine release by flavonoids and biflavonoids. *Planta Med.*, **1985**, *51*, 16-20.
- [294] Park, H.H.; Lee, S.; Son, H.Y.; Park, S.B.; Kim, M.S.; Choi, E.J.; Singh, T.S.; Ha, J.H.; Lee, M.G.; Kim, J.E.; Hyun, M.C.; Kwon, T.K.; Kim, Y.H.; Kim, S.H. Flavonoids inhibit histamine release and expression of proinflammatory cytokines in mast cells. *Arch. Pharm. Res.*, **2008**, *31*, 1303-11.
- [295] Itoh, T.; Ninomiya, M.; Yasuda, M.; Koshikawa, K.; Deyashiki, Y.; Nozawa, Y.; Akao, Y.; Koketsu, M. Inhibitory effects of flavonoids isolated from *Fragaria ananassa* Duch on IgE-mediated degranulation in rat basophilic leukemia RBL-2H3. *Bioorg. Med. Chem.*, **2009**, *17*, 5374-9.
- [296] Lee, E.J.; Ji, G.E.; Sung, M.K. Quercetin and kaempferol suppress immunoglobulin E-mediated allergic inflammation in RBL-2H3 and Caco-2 cells. *Inflamm. Res.*, **2010**, *59*, 847-54.
- [297] Hirose, E.; Matsushima, M.; Takagi, K.; Ota, Y.; Ishigami, K.; Hirayama, T.; Hayashi, Y.; Nakamura, T.; Hashimoto, N.; Imaizumi, K.; Baba, K.; Hasegawa, Y.; Kawabe, T. Involvement of heme oxygenase-1 in kaempferol-induced anti-allergic actions in RBL-2H3 cells. *Inflammation*, **2009**, *32*, 99-108.
- [298] Jung, C.H.; Lee, J.Y.; Park, J.H.; Cho, B.J.; Sim, S.S.; Kim, C.J. Flavonols attenuate the immediate and late-phase asthmatic responses to aerosolized-ovalbumin exposure in the conscious guinea pig. *Fitoterapia*, **2010**, *81*, 803-12.
- [299] Palanichamy, S. and Nagarajan, S. Analgesic activity of *Cassia alata* leaf extract and kaempferol 3-o-sophoroside. *J. Ethnopharmacol.*, **1990**, *29*, 73-8.
- [300] Cardenas, L.C.; Rodriguez, J.; Villaverde, M.C.; Riguera, R.; Cadena, R.; Otero, J.A. The analgesic activity of *Hedyosmum bonplandianum*: flavonoid glycosides. *Planta Med.*, **1993**, *59*, 26-7.
- [301] Maleki-Dizaji, N.; Fathiazad, F.; Garjani, A. Antinociceptive properties of extracts and two flavonoids isolated from leaves of *Danae racemosa*. *Arch. Pharm. Res.*, **2007**, *30*, 1536-42.
- [302] De Melo, G.O.; Malvar, D.C.; Vanderlinde, F.A.; Pires, P.A.; Cortes, W.S.; Filho, P.G.; Muzitano, M.F.; Kaiser, C.R.; Costa, S.S. Phytochemical and pharmacological study of *Sedum dendroideum* leaf juice. *J. Ethnopharmacol.*, **2005**, *102*, 217-20.
- [303] Okamoto, I.; Iwaki, K.; Koya-Miyata, S.; Tanimoto, T.; Kohno, K.; Ikeda, M.; Kurimoto, M. The flavonoid Kaempferol suppresses the graft-versus-host reaction by inhibiting type 1 cytokine production and CD8+ T cell engraftment. *Clin. Immunol.*, **2002**, *103*, 132-44.
- [304] Cortes, J.R.; Perez, G.; Rivas, M.D.; Zamorano, J. Kaempferol inhibits IL-4-induced STAT6 activation by specifically targeting JAK3. *J. Immunol.*, **2007**, *179*, 3881-7.
- [305] Gee, J.M. and Johnson, I.T. Polyphenolic compounds: interactions with the gut and implications for human health. *Curr. Med. Chem.*, **2001**, *8*, 1245-55.
- [306] Oliveira, E.J.; Watson, D.G.; Grant, M.H. Metabolism of quercetin and kaempferol by rat hepatocytes and the identification of flavonoid glycosides in human plasma. *Xenobiotica*, **2002**, *32*, 279-87.
- [307] Lehtonen, H.M.; Lehtinen, O.; Suomela, J.P.; Viitanen, M.; Kallio, H. Flavonol glycosides of sea buckthorn (*Hippophae rhamnoides* ssp. *sinensis*) and lingonberry (*Vaccinium vitis-idaea*) are bioavailable in humans and monoglucuronidated for excretion. *J. Agric. Food Chem.*, **2010**, *58*, 620-7.
- [308] Crespy, V.; Morand, C.; Besson, C.; Cotelle, N.; Vezin, H.; Dmignie, C.; Remesy, C. The splanchnic metabolism of flavonoids highly differed according to the nature of the compound. *Am. J. Physiol. Gastrointest. Liver Physiol.*, **2003**, *284*, G980-G988.
- [309] Bokkenheuser, V.D.; Shackleton, C.H.; Winter, J. Hydrolysis of dietary flavonoid glycosides by strains of intestinal Bacteroides from humans. *Biochem. J.*, **1987**, *248*, 953-6.
- [310] Winter, J.; Moore, L.H.; Dowell, V.R., Jr.; Bokkenheuser, V.D. C-ring cleavage of flavonoids by human intestinal bacteria. *Appl. Environ. Microbiol.*, **1989**, *55*, 1203-8.
- [311] Labib, S.; Hummel, S.; Richling, E.; Humpf, H.U.; Schreier, P. Use of the pig caecum model to mimic the human intestinal metabolism of hispidulin and related compounds. *Mol. Nutr. Food Res.*, **2006**, *50*, 78-86.
- [312] Simons, A.L.; Renouf, M.; Hendrich, S.; Murphy, P.A. Human gut microbial degradation of flavonoids: structure-function relationships. *J. Agric. Food Chem.*, **2005**, *53*, 4258-63.
- [313] Hein, E.M.; Rose, K.; Van't Slot, G.; Friedrich, A.W.; Humpf, H.U. Deconjugation and Degradation of Flavonol Glycosides by Pig Cecal Microbiota Characterized by Fluorescence in Situ Hybridization (FISH). *J. Agric. Food Chem.*, **2008**, *56*, 2281-90.
- [314] Schneider, H. and Blaut, M. Anaerobic degradation of flavonoids by *Eubacterium ramulus*. *Arch. Microbiol.*, **2000**, *173*, 71-5.
- [315] Oliveira, E.J. and Watson, D.G. *In vitro* glucuronidation of kaempferol and quercetin by human UGT-1A9 microsomes. *FEBS Lett.*, **2000**, *471*, 1-6.
- [316] Yodogawa, S.; Arakawa, T.; Sugihara, N.; Furuno, K. Glucuronidation and sulfo-conjugation of kaempferol in rat liver subcellular preparations and cultured hepatocytes. *Biol. Pharm. Bull.*, **2003**, *26*, 1120-4.

- [317] Barve, A.; Chen, C.; Hebbar, V.; Desiderio, J.; Saw, C.L.; Kong, A.N. Metabolism, oral bioavailability and pharmacokinetics of chemopreventive kaempferol in rats. *Biopharm. Drug Dispos.*, **2009**, *30*, 356-65.
- [318] DuPont, M.S.; Day, A.J.; Bennett, R.N.; Mellon, F.A.; Kroon, P.A. Absorption of kaempferol from endive, a source of kaempferol-3-glucuronide, in humans. *Eur. J. Clin. Nutr.*, **2004**, *58*, 947-54.
- [319] Wang, F.M.; Yao, T.W.; Zeng, S. Disposition of quercetin and kaempferol in human following an oral administration of Ginkgo biloba extract tablets. *Eur. J. Drug Metab Pharmacokinet.*, **2003**, *28*, 173-7.
- [320] Bonetti, A.; Marotti, I.; Dinelli, G. Urinary excretion of kaempferol from common beans (*Phaseolus vulgaris* L.) in humans. *Int. J. Food Sci. Nutr.*, **2007**, *58*, 261-9.
- [321] de Vries, J.H.; Hollman, P.C.; Meyboom, S.; Buysman, M.N.; Zock, P.L.; van Staveren, W.A.; Katan, M.B. Plasma concentrations and urinary excretion of the antioxidant flavonols quercetin and kaempferol as biomarkers for dietary intake. *Am. J. Clin. Nutr.*, **1998**, *68*, 60-5.
- [322] Radtke, J.; Linseisen, J.; Wolfram, G. Fasting plasma concentrations of selected flavonoids as markers of their ordinary dietary intake. *Eur. J. Nutr.*, **2002**, *41*, 203-9.
- [323] Wang, F.M.; Yao, T.W.; Zeng, S. Determination of quercetin and kaempferol in human urine after orally administrated tablet of ginkgo biloba extract by HPLC. *J. Pharm. Biomed. Anal.*, **2003**, *19*, 317-21.
- [324] Cao, J.; Zhang, Y.; Chen, W.; Zhao, X. The relationship between fasting plasma concentrations of selected flavonoids and their ordinary dietary intake. *Br. J. Nutr.*, **2010**, *103*, 249-55.
- [325] Silva, I.D.; Rodrigues, A.S.; Gaspar, J.; Laires, A.; Rueff, J. Metabolism of galangin by rat cytochromes P450: relevance to the genotoxicity of galangin. *Mutat. Res.*, **1997**, *393*, 247-57.
- [326] Nielsen, S.E.; Breinholt, V.; Justesen, U.; Cornett, C.; Dragsted, L.O. *In vitro* biotransformation of flavonoids by rat liver microsomes. *Xenobiotica*, **1998**, *28*, 389-401.
- [327] Breinholt, V.M.; Offord, E.A.; Brouwer, C.; Nielsen, S.E.; Brosen, K.; Friedberg, T. *In vitro* investigation of cytochrome P450-mediated metabolism of dietary flavonoids. *Food Chem. Toxicol.*, **2002**, *40*, 609-16.
- [328] Boots, A.W.; Haenen, G.R.; Bast, A. Health effects of quercetin: from antioxidant to nutraceutical. *Eur. J. Pharmacol.*, **2008**, *585*, 325-37.
- [329] Murakami, A.; Ashida, H.; Terao, J. Multitargeted cancer prevention by quercetin. *Cancer Lett.*, **2008**, *269*, 315-25.
- [330] Edenharder, R.; von, P., I.; Rauscher, R. Antimutagenic effects of flavonoids, chalcones and structurally related compounds on the activity of 2-amino-3-methylimidazo[4,5-f]quinoline (IQ) and other heterocyclic amine mutagens from cooked food. *Mutat. Res.*, **1993**, *287*, 261-74.
- [331] Huang, M.T.; Wood, A.W.; Newmark, H.L.; Sayer, J.M.; Yagi, H.; Jerina, D.M.; Conney, A.H. Inhibition of the mutagenicity of bay-region diol-epoxides of polycyclic aromatic hydrocarbons by phenolic plant flavonoids. *Carcinogenesis*, **1983**, *4*, 1631-7.
- [332] Edenharder, R. and Tang, X. Inhibition of the mutagenicity of 2-nitrofluorene, 3-nitrofluoranthene and 1-nitropyrene by flavonoids, coumarins, quinones and other phenolic compounds. *Food Chem. Toxicol.*, **1997**, *35*, 357-72.
- [333] MacGregor, J.T. and Jurd, L. Mutagenicity of plant flavonoids: structural requirements for mutagenic activity in *Salmonella typhimurium*. *Mutat. Res.*, **1978**, *54*, 297-309.
- [334] Maruta, A.; Enaka, K.; Umeda, M. Mutagenicity of quercetin and kaempferol on cultured mammalian cells. *Gann*, **1979**, *70*, 273-6.
- [335] Brown, J.P. and Dietrich, P.S. Mutagenicity of plant flavonols in the *Salmonella/mammalian* microsome test: activation of flavonol glycosides by mixed glycosidases from rat cecal bacteria and other sources. *Mutat. Res.*, **1979**, *66*, 223-40.
- [336] Sahu, R.K.; Basu, R.; Sharma, A. Genetic toxicological of some plant flavonoids by the micronucleus test. *Mutat. Res.*, **1981**, *89*, 69-74.
- [337] Silva, I.D.; Rodrigues, A.; Gaspar, J.; Maia, R.; Laires, A.; Rueff, J. Mutagenicity of kaempferol in V79 cells: the role of cytochromes P450. *Teratog. Carcinog. Mutagen.*, **1996**, *16*, 229-41.
- [338] Silva, I.D.; Rodrigues, A.S.; Gaspar, J.; Maia, R.; Laires, A.; Rueff, J. Involvement of rat cytochrome 1A1 in the biotransformation of kaempferol to quercetin: relevance to the genotoxicity of kaempferol. *Mutagenesis*, **1997**, *12*, 383-90.
- [339] Sahu, S.C. and Gray, G.C. Kaempferol-induced nuclear DNA damage and lipid peroxidation. *Cancer Lett.*, **1994**, *85*, 159-64.
- [340] Sahu, S.C. and Gray, G.C. Pro-oxidant activity of flavonoids: effects on glutathione and glutathione S-transferase in isolated rat liver nuclei. *Cancer Lett.*, **1996**, *104*, 193-6.
- [341] Niering, P.; Michels, G.; Watjen, W.; Ohler, S.; Steffan, B.; Chovolou, Y.; Kampkotter, A.; Proksch, P.; Kahl, R. Protective and detrimental effects of kaempferol in rat H4IIE cells: implication of oxidative stress and apoptosis. *Toxicol. Appl. Pharmacol.*, **2005**, *209*, 114-22.
- [342] Canada, A.T.; Giannella, E.; Nguyen, T.D.; Mason, R.P. The production of reactive oxygen species by dietary flavonols. *Free Radic. Biol. Med.*, **1990**, *9*, 441-9.
- [343] Galati, G.; Sabzevari, O.; Wilson, J.X.; O'Brien, P.J. Prooxidant activity and cellular effects of the phenoxy radicals of dietary flavonoids and other polyphenolics. *Toxicology*, **2002**, *177*, 91-104.
- [344] Harwood, M.; Danielewska-Nikiel, B.; Borzelleca, J.F.; Flamm, G.W.; Williams, G.M.; Lines, T.C. A critical review of the data related to the safety of quercetin and lack of evidence of *in vivo* toxicity, including lack of genotoxic/carcinogenic properties. *Food Chem. Toxicol.*, **2007**, *45*, 2179-205.
- [345] Utesch, D.; Feige, K.; Dasenbrock, J.; Broschard, T.H.; Harwood, M.; Danielewska-Nikiel, B.; Lines, T.C. Evaluation of the potential *in vivo* genotoxicity of quercetin. *Mutat. Res.*, **2008**, *654*, 38-44.
- [346] Takashi, H.; Aiso, S.; Hirono, I.; Matsushima, T.; Sugimura, T. Carcinogenicity test of quercetin and kaempferol in rats by oral administration. *J. Food Safety*, **1983**, *5*, 55-60.
- [347] Hu, Y.; Cheng, Z.; Heller, L.I.; Krasnoff, S.B.; Glahn, R.P.; Welch, R.M. Kaempferol in red and pinto bean seed (*Phaseolus vulgaris* L.) coats inhibits iron bioavailability using an *in vitro* digestion/human Caco-2 cell model. *J. Agric. Food Chem.*, **2006**, *54*, 9254-61.
- [348] Lemos, C.; Peters, G.J.; Jansen, G.; Martel, F.; Calhau, C. Modulation of folate uptake in cultured human colon adenocarcinoma Caco-2 cells by dietary compounds. *Eur. J. Nutr.*, **2007**, *46*, 329-36.
- [349] Li, C.; Li, X.; Choi, J.S. Enhanced bioavailability of etoposide after oral or intravenous administration of etoposide with kaempferol in rats. *Arch. Pharm. Res.*, **2009**, *32*, 133-8.
- [350] Gao, S.S.; Choi, B.M.; Chen, X.Y.; Zhu, R.Z.; Kim, Y.; So, H.; Park, R.; Sung, M.; Kim, B.R. Kaempferol suppresses cisplatin-induced apoptosis via inductions of heme oxygenase-1 and glutamate-cysteine ligase catalytic subunit in HEI-OC1 cell. *Pharm. Res.*, **2010**, *27*, 235-45.
- [351] Lopez-Lazaro, M. Anticancer and carcinogenic properties of curcumin: considerations for its clinical development as a cancer chemopreventive and chemotherapeutic agent. *Mol. Nutr. Food Res.*, **2008**, *52*, S103-S127.
- [352] de Vries, J.H.; Janssen, P.L.; Hollman, P.C.; van Staveren, W.A.; Katan, M.B. Consumption of quercetin and kaempferol in free-living subjects eating a variety of diets. *Cancer Lett.*, **1997**, *19*, 141-4.
- [353] Sikorska, M. and Matlawska, I. Polyphenolic compounds from *Abutilon grandiflorum* leaves. *Acta Pol. Pharm.*, **2008**, *65*, 467-71.
- [354] Matlawska, I. and Sikorska, M. Flavonoids from *Abutilon theophrasti* flowers. *Acta Pol. Pharm.*, **2005**, *62*, 135-9.
- [355] Singh, R.; Singh, B.; Singh, S.; Kumar, N.; Kumar, S.; Arora, S. Anti-free radical activities of kaempferol isolated from *Acacia nilotica* (L.) Willd. Ex. Del. *Toxicol. In vitro*, **2008**, *22*, 1965-70.
- [356] Backhouse, N.; Delporte, C.; Negrete, R.; Feliciano, S.A.; Lopez-Perez, J.L. Bioactive phenolic derivatives from *Acacia splendens* methanol extract. *Phytother. Res.*, **2002**, *16*, 562-6.
- [357] Adesina, S.K.; Idowu, O.; Ogundaini, A.O.; Oladimeji, H.; Olugbade, T.A.; Onawunmi, G.O.; Pais, M. Antimicrobial constituents of the leaves of *Acalypha wilkesiana* and *Acalypha hispida*. *Phytother. Res.*, **2000**, *14*, 371-4.
- [358] Sawada, H.; Miyakoshi, M.; Isoda, S.; Ida, Y.; Shoji, J. Saponins from leaves of *Acanthopanax sieboldianus*. *Phytochemistry*, **1993**, *34*, 1117-21.
- [359] Han, J.T.; Bang, M.H.; Chun, O.K.; Kim, D.O.; Lee, C.Y.; Baek, N.I. Flavonol glycosides from the aerial parts of *Aceriphyllum rossii* and their antioxidant activities. *Arch. Pharm. Res.*, **2004**, *27*, 390-5.

- [360] Braca, A.; Fico, G.; Morelli, I.; De Simone, F.; Tome, F.; De Tommasi, N. Antioxidant and free radical scavenging activity of flavonol glycosides from different *Aconitum* species. *J. Ethnopharmacol.*, **2003**, *86*, 63-7.
- [361] Fico, G.; Braca, A.; De Tommasi, N.; Tome, F.; Morelli, I. Flavonoids from *Aconitum napellus* subsp. *neomontanum*. *Phytochemistry*, **2001**, *57*, 543-6.
- [362] Shrestha, B.B.; Dall'Acqua, S.; Gewali, M.B.; Jha, P.K.; Innocenti, G. New flavonoid glycosides from *Aconitum naviculare* (Bruhl) Stapf, a medicinal herb from the trans-Himalayan region of Nepal. *Carbohydr. Res.*, **2006**, *341*, 2161-5.
- [363] Fico, G.; Braca, A.; Bilia, A.R.; Tome, F.; Morelli, I. Flavonol glycosides from the flowers of *Aconitum paniculatum*. *J. Nat. Prod.*, **2000**, *63*, 1563-5.
- [364] Vitalini, S.; Braca, A.; Passerella, D.; Fico, G. New flavonol glycosides from *Aconitum burnatii* Gay and *Aconitum variegatum* L. *Fitoterapia*, **2010**, *81*, 940-7.
- [365] Xin, H.L.; Wu, Y.C.; Su, Y.H.; Sheng, J.Y.; Ling, C.Q. Novel Flavonoids from the Leaves of *Actinidia valvata* Dunn: Structural Elucidation and Antioxidant Activity. *Planta Med.*, **2011**, *77*, 70-3.
- [366] Itoh, A.; Tanahashi, T.; Nagakura, N.; Takenaka, Y.; Chen, C.C.; Pelletier, J. Flavonoid glycosides from *Adina racemosa* and their inhibitory activities on eukaryotic protein synthesis. *J. Nat. Prod.*, **2004**, *67*, 427-31.
- [367] Tran, M.H.; Nguyen, H.D.; Kim, J.C.; Choi, J.S.; Lee, H.K.; Min, B.S. Phenolic glycosides from *Alangium salviifolium* leaves with inhibitory activity on LPS-induced NO, PGE(2), and TNF-alpha production. *Bioorg. Med. Chem. Lett.*, **2009**, *19*, 4389-93.
- [368] el Mousallamy, A.M. Leaf flavonoids of *Albizia lebbek*. *Phytochemistry*, **1998**, *48*, 759-61.
- [369] Rodriguez, G.B.; Rodriguez Rodriguez, E.M.; Diaz, R.C. Flavonoids in onion cultivars (*Allium cepa* L.). *J. Food Sci.*, **2008**, *73*, C599-C605.
- [370] Barile, E.; Capasso, R.; Izzo, A.A.; Lanzotti, V.; Sajjadi, S.E.; Zolfaghari, B. Structure-activity relationships for saponins from *Allium hirtifolium* and *Allium elburzense* and their antispasmodic activity. *Planta Med.*, **2005**, *71*, 1010-8.
- [371] Carotenuto, A.; Fattorusso, E.; Lanzotti, V.; Magno, S.; De, F., V.; Cicala, C. The flavonoids of *Allium neapolitanum*. *Phytochemistry*, **1997**, *44*, 949-57.
- [372] Fattorusso, E.; Lanzotti, V.; Tagliatalata-Scafati, O.; Cicala, C. The flavonoids of leek, *Allium porrum*. *Phytochemistry*, **2001**, *57*, 565-9.
- [373] Corea, G.; Fattorusso, E.; Lanzotti, V. Saponins and flavonoids of *Allium triquetrum*. *J. Nat. Prod.*, **2003**, *66*, 1405-11.
- [374] Carotenuto, A.; De, F., V.; Fattorusso, E.; Lanzotti, V.; Magno, S.; Cicala, C. The flavonoids of *Allium ursinum*. *Phytochemistry*, **1996**, *41*, 531-6.
- [375] Lee, K.T.; Choi, J.H.; Kim, D.H.; Son, K.H.; Kim, W.B.; Kwon, S.H.; Park, H.J. Constituents and the antitumor principle of *Allium victorialis* var. *platyphyllum*. *Arch. Pharm. Res.*, **2001**, *24*, 44-50.
- [376] Keyhanian, S. and Stahl-Biskup, E. Phenolic constituents in dried flowers of aloe vera (*Aloe barbadensis*) and their *in vitro* antioxidative capacity. *Planta Med.*, **2007**, *73*, 599-602.
- [377] Scio, E.; Ribeiro, A.; Alves, T.M.; Romanha, A.J.; Dias de Souza, F.J.; Cordell, G.A.; Zani, C.L. Diterpenes from *Alomia myriadenia* (Asteraceae) with cytotoxic and trypanocidal activity. *Phytochemistry*, **2003**, *64*, 1125-31.
- [378] Salvador, M.J.; Ferreira, E.O.; Mertens-Talcott, S.U.; De Castro, W.V.; Butterweck, V.; Derendorf, H.; Dias, D.A. Isolation and HPLC quantitative analysis of antioxidant flavonoids from *Alternanthera tenella* Colla. *Z. Naturforsch. [C.]*, **2006**, *61*, 19-25.
- [379] Papiez, M.; Gancarczyk, M.; Bilinska, B. The compounds from the hollyhock extract (*Althaea rosea* Cav. var. *nigra*) affect the aromatization in rat testicular cells *in vivo* and *in vitro*. *Folia Histochem. Cytobiol.*, **2002**, *40*, 353-9.
- [380] Stintzing, F.C.; Kammerer, D.; Schieber, A.; Adama, H.; Nacoulma, O.G.; Carle, R. Betacyanins and phenolic compounds from *Amaranthus spinosus* L. and *Boerhavia erecta* L. *Z. Naturforsch. [C.]*, **2004**, *59*, 1-8.
- [381] Costa-Lotufo, L.V.; Jimenez, P.C.; Wilke, D.V.; Leal, L.K.; Cunha, G.M.; Silveira, E.R.; Canuto, K.M.; Viana, G.S.; Moraes, M.E.; de Moraes, M.O.; Pessoa, C. Antiproliferative effects of several compounds isolated from *Amburana cearensis* A. C. Smith. *Z. Naturforsch. [C.]*, **2003**, *58*, 675-80.
- [382] Singab, A.N. Acetylated flavonol triglycosides from *Ammi majus* L. *Phytochemistry*, **1998**, *49*, 2177-80.
- [383] Abdelfattah, M.S.; Toume, K.; Ahmed, F.; Sadhu, S.K.; Ishibashi, M. Cucullamide, a new putrescine bisamide from *Amoora cucullata*. *Chem. Pharm. Bull. (Tokyo)*, **2010**, *58*, 1116-8.
- [384] Wu, Y.Q.; Li, Y.; Lin, X.F.; Liu, Z.L.; Li, Y. Phenolic compounds from *Anaphalis aureo-punctata*. *Pharmazie*, **2003**, *58*, 833-5.
- [385] Chang, F.R.; Wei, J.L.; Teng, C.M.; Wu, Y.C. Antiplatelet aggregation constituents from *Annona purpurea*. *J. Nat. Prod.*, **1998**, *61*, 1457-61.
- [386] Xiong, Q.; Fan, W.; Tezuka, Y.; Adnyana, I.K.; Stampoulis, P.; Hattori, M.; Namba, T.; Kadota, S. Hepatoprotective effect of *Apocynum venetum* and its active constituents. *Planta Med.*, **2000**, *66*, 127-33.
- [387] Ryan, K.G.; Swinny, E.E.; Winefield, C.; Markham, K.R. Flavonoids and UV photoprotection in *Arabidopsis* mutants. *Z. Naturforsch. [C.]*, **2001**, *56*, 745-54.
- [388] Veit, M. and Pauli, G.F. Major flavonoids from *Arabidopsis thaliana* leaves. *J. Nat. Prod.*, **1999**, *62*, 1301-3.
- [389] Sumino, M.; Sekine, T.; Ruangrunsi, N.; Igarashi, K.; Ikegami, F. Ardisiphenols and other antioxidant principles from the fruits of *Ardisia colorata*. *Chem. Pharm. Bull. (Tokyo)*, **2002**, *50*, 1484-7.
- [390] Li, Y.F.; Hu, L.H.; Lou, F.C.; Li, J.; Shen, Q. PTP1B inhibitors from *Ardisia japonica*. *J. Asian Nat. Prod. Res.*, **2005**, *7*, 13-8.
- [391] Sikorska, M. Flavonoids in the leaves of *Asclepias incarnata* L. *Acta Pol. Pharm.*, **2003**, *60*, 471-5.
- [392] Sikorska, M.; Matlawska, I.; Franski, R. Kaempferol and its glycosides in the seeds hair of *Asclepias syriaca* L. *Acta Pol. Pharm.*, **2001**, *58*, 211-5.
- [393] Mizuno, M.; Kyotani, Y.; Inuma, M.; Tanaka, T.; Iwatsuki, K. Kaempferol 3-rhamnoside-7-[6-feruloyl]glucosyl (1----3)rhamnoside] from *Asplenium prolougatum*. *Phytochemistry*, **1990**, *29*, 2742-3.
- [394] Dall'Acqua, S.; Tome, F.; Vitalini, S.; Agradi, E.; Innocenti, G. *In vitro* estrogenic activity of *Asplenium trichomanes* L. extracts and isolated compounds. *J. Ethnopharmacol.*, **2009**, *122*, 424-9.
- [395] Semmar, N.; Fenet, B.; Lacaille-Dubois, M.A.; Gluchoff-Fiasson, K.; Chemli, R.; Jay, M. Two new glycosides from *Astragalus caprinus*. *J. Nat. Prod.*, **2001**, *64*, 656-8.
- [396] Semmar, N.; Fenet, B.; Gluchoff-Fiasson, K.; Hasan, A.; Jay, M. Four new flavonol glycosides from the leaves of *Astragalus caprinus*. *J. Nat. Prod.*, **2002**, *65*, 576-9.
- [397] Yahara, S.; Kohjyouma, M.; Kohoda, H. Flavonoid glycosides and saponins from *Astragalus shikokianus*. *Phytochemistry*, **2000**, *53*, 469-71.
- [398] De Leo, M.; Braca, A.; De Tommasi, N.; Norscia, I.; Morelli, I.; Battinelli, L.; Mazzanti, G. Phenolic compounds from *Baseonema acuminatum* leaves: isolation and antimicrobial activity. *Planta Med.*, **2004**, *70*, 841-6.
- [399] Kaewamatawong, R.; Kitajima, M.; Kogure, N.; Takayama, H. Flavonols from *Bauhinia malabarica*. *Nat. Med. (Tokyo)*, **2008**, *62*, 364-5.
- [400] Rodriguez, P.; Gonzalez-Mujica, F.; Bermudez, J.; Hasegawa, M. Inhibition of Glucose Intestinal Absorption by Kaempferol 3-O-alpha-rhamnoside Purified from *Bauhinia megalandra* leaves. *Fitoterapia*, **2010**, *81*, 1220-3.
- [401] Meyre-Silva, C.; Yunes, R.A.; Delle, M.F.; Santos, A.R.; Schmeling, L.O.; Gadotti, V.M.; Liz, F.; Cechinel-Filho, V. Phytochemical and pharmacological analysis of *Bauhinia microstachya* (Raddi) Macbr. (Leguminosae). *Z. Naturforsch. [C.]*, **2001**, *56*, 939-42.
- [402] Kaouadji, M. Flavonol diglycosides from *Blackstonia perfoliata*. *Phytochemistry*, **1990**, *29*, 1345-7.
- [403] Kaouadji, M.; Doucoure, A.; Mariotte, A.M.; Chulia, A.J.; Thomasson, F. Flavonol triglycosides from *Blackstonia perfoliata*. *Phytochemistry*, **1990**, *29*, 1283-6.
- [404] Swinny, E.E. A novel acetylated 3-deoxyanthocyanidin laminaribioside from the fern *Blechnum novae-zelandiae*. *Z. Naturforsch. [C.]*, **2001**, *56*, 177-80.
- [405] Nielsen, J.K.; Olsen, C.E.; Petersen, M.K. Acylated flavonol glycosides from cabbage leaves. *Phytochemistry*, **1993**, *34*, 539-44.
- [406] Harbaum, B.; Hubbermann, E.M.; Wolff, C.; Herges, R.; Zhu, Z.; Schwarz, K. Identification of flavonoids and hydroxycinnamic acids in pak choi varieties (*Brassica campestris* L. ssp. *chinensis* var. *communis*) by HPLC-ESI-MSn and NMR and their

- quantification by HPLC-DAD. *J. Agric. Food Chem.*, **2007**, *55*, 8251-60.
- [407] Kim, J.E.; Jung, M.J.; Jung, H.A.; Woo, J.J.; Cheigh, H.S.; Chung, H.Y.; Choi, J.S. A new kaempferol 7-O-triglucoside from the leaves of *Brassica juncea* L. *Arch. Pharm. Res.*, **2002**, *25*, 621-4.
- [408] Olsen, H.; Aaby, K.; Borge, G.I. Characterization and quantification of flavonoids and hydroxycinnamic acids in curly kale (*Brassica oleracea* L. Convar. *acephala* Var. *sabellica*) by HPLC-DAD-ESI-MSn. *J. Agric. Food Chem.*, **2009**, *57*, 2816-25.
- [409] Rochfort, S.J.; Imsic, M.; Jones, R.; Trenerry, V.C.; Tomkins, B. Characterization of flavonol conjugates in immature leaves of pak choi [*Brassica rapa* L. Ssp. *chinensis* L. (Hanelt.)] by HPLC-DAD and LC-MS/MS. *J. Agric. Food Chem.*, **2006**, *54*, 4855-60.
- [410] Bennett, R.N.; Rosa, E.A.; Mellon, F.A.; Kroon, P.A. Ontogenic profiling of glucosinolates, flavonoids, and other secondary metabolites in *Eruca sativa* (salad rocket), *Diplotaxis erucoides* (wall rocket), *Diplotaxis tenuifolia* (wild rocket), and *Bunias orientalis* (Turkish rocket). *J. Agric. Food Chem.*, **2006**, *54*, 4005-15.
- [411] Sharififar, F.; Yassa, N.; Mozaffarian, V. Bioactivity of major components from the seeds of *Bunium persicum* (Boiss.) Fedtch. *Pak. J. Pharm. Sci.*, **2010**, *23*, 300-4.
- [412] Pistelli, L.; Nocioli, C.; Giachi, I.; Dimitrova, B.; Gevrenova, R.; Morelli, I.; Potenza, D. Lupane-triterpenes from *Bupleurum flavum*. *Nat. Prod. Res.*, **2005**, *19*, 783-8.
- [413] Badria, F.A.; Ameen, M.; Akl, M.R. Evaluation of cytotoxic compounds from calligonum comosum L. growing in Egypt. *Z. Naturforsch. [C.]*, **2007**, *62*, 656-60.
- [414] Mahmoud, I.I.; Moharram, F.A.; Marzouk, M.S.; Linscheid, M.W.; Saleh, M.I. Polyphenolic constituents of *Callistemon lanceolatus* leaves. *Pharmazie*, **2002**, *57*, 494-6.
- [415] Sekine, T.; Arita, J.; Yamaguchi, A.; Saito, K.; Okonogi, S.; Morisaki, N.; Iwasaki, S.; Murakoshi, I. Two flavonol glycosides from seeds of *Camellia sinensis*. *Phytochemistry*, **1991**, *30*, 991-5.
- [416] Finger, A.; Engelhardt, U.H.; Wray, V. Flavonol triglycosides containing galactose in tea. *Phytochemistry*, **1991**, *30*, 2057-60.
- [417] Sekine, T.; Arai, Y.; Ikegami, F.; Fujii, Y.; Shindo, S.; Yamagisawa, T.; Ishida, Y.; Okonogi, S.; Murakoshi, I. Isolation of camelliaside C from "tea seed cake" and inhibitory effects of its derivatives on arachidonate 5-lipoxygenase. *Chem. Pharm. Bull. (Tokyo)*, **1993**, *41*, 1185-7.
- [418] Kinjo, J.; Nagao, T.; Tanaka, T.; Nonaka, G.; Okawa, M.; Nohara, T.; Okabe, H. Activity-guided fractionation of green tea extract with antiproliferative activity against human stomach cancer cells. *Biol. Pharm. Bull.*, **2002**, *25*, 1238-40.
- [419] Park, J.S.; Rho, H.S.; Kim, D.H.; Chang, I.S. Enzymatic preparation of kaempferol from green tea seed and its antioxidant activity. *J. Agric. Food Chem.*, **2006**, *19*:54, 2951-6.
- [420] Lee, V.S.; Chen, C.R.; Liao, Y.W.; Tzen, J.T.; Chang, C.I. Structural determination and DPPH radical-scavenging activity of two acylated flavonoid tetraglycosides in oolong tea (*Camellia sinensis*). *Chem. Pharm. Bull. (Tokyo)*, **2008**, *56*, 851-3.
- [421] Dumlu, M.U.; Gurkan, E.; Tuzlaci, E. Chemical composition and antioxidant activity of *Campanula alliariifolia*. *Nat. Prod. Res.*, **2008**, *22*, 477-82.
- [422] Cuendet, M.; Poterat, O.; Hostettmann, K. Flavonoids and phenylpropanoid derivatives from *Campanula barbata*. *Phytochemistry*, **2001**, *56*, 631-6.
- [423] Li, N.; Li, X.; Meng, D.L.; Guo, Y.Q.; Wang, J.H. Flavonoids from *Camptosorus sibiricus* Rupr. *J. Asian Nat. Prod. Res.*, **2006**, *8*, 167-71.
- [424] Murakami, T.; Kohno, K.; Kishi, A.; Matsuda, H.; Yoshikawa, M. Medicinal foodstuffs. XIX. Absolute stereostructures of canavalioides, a new Ent-kaurene-type diterpene glycoside, and gladiatosides A1, A2, A3, B1, B2, B3, C1, and C2, new acylated flavonol glycosides, from sword bean, the seeds of *Canavalia gladiata*. *Chem. Pharm. Bull. (Tokyo)*, **2000**, *48*, 1673-80.
- [425] Ross, S.A.; ElSohly, M.A.; Sultana, G.N.; Mehmedic, Z.; Hossain, C.F.; Chandra, S. Flavonoid glycosides and cannabinoids from the pollen of *Cannabis sativa* L. *Phytochem. Anal.*, **2005**, *16*, 45-8.
- [426] Taskova, R.; Mitova, M.; Mikhova, B.; Duddeck, H. Bioactive phenolics from *Carthamus lanatus* L. *Z. Naturforsch. [C.]*, **2003**, *58*, 704-7.
- [427] Ahmed, K.M.; Marzouk, M.S.; el Khriy, E.A.; Wahab, S.A.; el Din, S.S. A new flavone diglycoside from *Carthamus tinctorius* seeds. *Pharmazie*, **2000**, *55*, 621-2.
- [428] Moriyama, H.; Iizuka, T.; Nagai, M. A stabilized flavonoid glycoside in heat-treated *Cassia alata* leaves and its structural elucidation. *Yakugaku Zasshi.*, **2001**, *121*, 817-20.
- [429] Moriyama, H.; Iizuka, T.; Nagai, M.; Miyataka, H.; Satoh, T. Antiinflammatory activity of heat-treated *Cassia alata* leaf extract and its flavonoid glycoside. *Yakugaku Zasshi.*, **2003**, *123*, 607-11.
- [430] Terreaux, C.; Wang, Q.; Ioset, J.R.; Ndjoko, K.; Grimminger, W.; Hostettmann, K. Complete LC/MS analysis of a Tinneveli senna pod extract and subsequent isolation and identification of two new benzophenone glucosides. *Planta Med.*, **2002**, *68*, 349-54.
- [431] Kumar, R.; Ilyas, M.; Parveen, M.; Shafiqullah A new chromone from *Cassia nodosa*. *J. Asian Nat. Prod. Res.*, **2006**, *8*, 595-8.
- [432] Nsonde Ntandou, G.F.; Banzouzi, J.T.; Mbatchi, B.; Elion-Itou, R.D.; Etou-Ossibi, A.W.; Ramos, S.; Benoit-Vical, F.; Abena, A.A.; Ouamba, J.M. Analgesic and anti-inflammatory effects of *Cassia siamea* Lam. stem bark extracts. *J. Ethnopharmacol.*, **2010**, *127*, 108-11.
- [433] Drewes, S.E. and Taylor, C.W. Methylated A-type proanthocyanidins and related metabolites from *Cassipourea gummiflua*. *Phytochemistry*, **1994**, *37*, 551-5.
- [434] Ly, T.N.; Shimoyamada, M.; Yamauchi, R. Isolation and characterization of rosmarinic acid oligomers in *Celastrus hindsii* Benth leaves and their antioxidative activity. *J. Agric. Food Chem.*, **2006**, *54*, 3786-93.
- [435] Rzakowska-Bodalska, H.; Olechnowicz-Stepien, W.; Rzepka, A. The components of ethereal and methanolic extracts from the fruits of *Celastrus tatarinowii* (Rupr.). *Pol. J. Pharmacol. Pharm.*, **1975**, *27*, 335-8.
- [436] Karamenderes, C.; Bedir, E.; Pawar, R.; Baykan, S.; Khan, I.A. Elemanolide sesquiterpenes and eudesmane sesquiterpene glycosides from *Centaurea hierapolitana*. *Phytochemistry*, **2007**, *68*, 609-15.
- [437] Satake, T.; Kamiya, K.; An, Y.; Oishi Nee, T.T.; Yamamoto, J. The anti-thrombotic active constituents from *Centella asiatica*. *Biol. Pharm. Bull.*, **2007**, *30*, 935-40.
- [438] Suntornasuk, L. and Anurukvorakun, O. Precision improvement for the analysis of flavonoids in selected Thai plants by capillary zone electrophoresis. *Electrophoresis*, **2005**, *26*, 648-60.
- [439] Liu, Q.; Liu, M.; Mabry, T.J.; Dixon, R.A. Flavonol glycosides from *Cephalocereus senilis*. *Phytochemistry*, **1994**, *36*, 229-31.
- [440] Gohar, A.A.; Maatooq, G.T.; Niwa, M. Two flavonoid glycosides from *Chenopodium murale*. *Phytochemistry*, **2000**, *53*, 299-303.
- [441] De Simone, F.; Dini, A.; Pizzi, C.; Saturnino, P.; Schettino, O. Two flavonol glycosides from *Chenopodium quinoa*. *Phytochemistry*, **1990**, *29*, 3690-2.
- [442] Kwak, J.H.; Kang, M.W.; Roh, J.H.; Choi, S.U.; Zee, O.P. Cytotoxic phenolic compounds from *Chionanthus retusus*. *Arch. Pharm. Res.*, **2009**, *32*, 1681-7.
- [443] Ling, S.K.; Pizar, M.M.; Man, S. Platelet-activating factor (PAF) receptor binding antagonist activity of the methanol extracts and isolated flavonoids from *Chromolaena odorata* (L.) King and Robinson. *Biol. Pharm. Bull.*, **2007**, *30*, 1150-2.
- [444] Landa, A.; Casado, R.; Calvo, M.I. Identification and quantification of flavonoids from *Chiquiraga spinosa* (Asteraceae). *Nat. Prod. Commun.*, **2009**, *4*, 1353-5.
- [445] DuPont, M.S.; Mondin, Z.; Williamson, G.; Price, K.R. Effect of variety, processing, and storage on the flavonoid glycoside content and composition of lettuce and endive. *J. Agric. Food Chem.*, **2000**, *48*, 3957-64.
- [446] Fang, S.H.; Rao, Y.K.; Tzeng, Y.M. Inhibitory effects of flavonol glycosides from *Cinnamomum osmophloeum* on inflammatory mediators in LPS/IFN-gamma-activated murine macrophages. *Bioorg. Med. Chem.*, **2005**, *13*, 2381-8.
- [447] Nazaruk, J. and Jakoniuk, P. Flavonoid composition and antimicrobial activity of *Cirsium rivulare* (Jacq.) All. flowers. *J. Ethnopharmacol.*, **2005**, *102*, 208-12.
- [448] Beltrame, F.L.; Ferreira, A.G.; Cortez, D.A. Coumarin glycoside from *Cissus sicyoides*. *Nat. Prod. Lett.*, **2002**, *16*, 213-6.
- [449] Chaves, N.; Sosa, T.; Escudero, J.C. Plant growth inhibiting flavonoids in exudate of *Cistus ladanifer* and in associated soils. *J. Chem. Ecol.*, **2001**, *27*, 623-31.

- [450] Sosa, T.; Chaves, N.; Alias, J.C.; Escudero, J.C.; Henao, F.; Gutierrez-Merino, C. Inhibition of mouth skeletal muscle relaxation by flavonoids of *Cistus ladanifer* L.: a plant defense mechanism against herbivores. *J. Chem. Ecol.*, **2004**, *30*, 1087-101.
- [451] Sadhu, S.K.; Okuyama, E.; Fujimoto, H.; Ishibashi, M.; Yesilada, E. Prostaglandin inhibitory and antioxidant components of *Cistus laurifolius*, a Turkish medicinal plant. *J. Ethnopharmacol.*, **2006**, *108*, 371-8.
- [452] Kupeli, E.; Orhan, D.D.; Yesilada, E. Effect of *Cistus laurifolius* L. leaf extracts and flavonoids on acetaminophen-induced hepatotoxicity in mice. *J. Ethnopharmacol.*, **2006**, *20*, 103, 455-60.
- [453] Berhow, M.A.; Bennett, R.D.; Poling, S.M.; Vannier, S.; Hidaka, T.; Omura, M. Acylated flavonoids in callus cultures of *Citrus aurantifolia*. *Phytochemistry*, **1994**, *36*, 1225-7.
- [454] Kazuma, K.; Noda, N.; Suzuki, M. Malonylated flavonol glycosides from the petals of *Clitoria ternatea*. *Phytochemistry*, **2003**, *62*, 229-37.
- [455] Kuti, J.O. and Konuru, H.B. Antioxidant capacity and phenolic content in leaf extracts of tree spinach (*Cnidioscolus* spp.). *J. Agric. Food Chem.*, **2004**, *52*, 117-21.
- [456] Lee, S.S.; Chen, W.C.; Chen, C.H. New jujubogenin glycosides from *Colubrina asiatica*. *J. Nat. Prod.*, **2000**, *63*, 1580-3.
- [457] Mahmoud, A.A.; Al Shihry, S.S.; Hegazy, M.E. Biological activity of a phloroglucinol glucoside derivative from *Conyza aegyptiaca*. *Z. Naturforsch. C.*, **2009**, *64*, 513-7.
- [458] Calzada, F.; Cedillo-Rivera, R.; Mata, R. Antiprotozoal activity of the constituents of *Conyza filaginoides*. *J. Nat. Prod.*, **2001**, *64*, 671-3.
- [459] Amaral, J.S.; Ferreres, F.; Andrade, P.B.; Valentao, P.; Pinheiro, C.; Santos, A.; Seabra, R. Phenolic profile of hazelnut (*Corylus avellana* L.) leaves cultivars grown in Portugal. *Nat. Prod. Res.*, **2005**, *19*, 157-63.
- [460] Autore, G.; Rastrelli, L.; Lauro, M.R.; Marzocco, S.; Sorrentino, R.; Sorrentino, U.; Pinto, A.; Aquino, R. Inhibition of nitric oxide synthase expression by a methanolic extract of *Crescentia alata* and its derived flavonols. *Life Sci.*, **2001**, *70*, 523-34.
- [461] Norbaek, R. and Kondo, T. Flavonol glycosides from flowers of *Crocus speciosus* and *C. antalyensis*. *Phytochemistry*, **1999**, *51*, 1113-9.
- [462] Li, C.Y.; Lee, E.J.; Wu, T.S. Antityrosinase principles and constituents of the petals of *Crocus sativus*. *J. Nat. Prod.*, **2004**, *67*, 437-40.
- [463] Kubo, I. and Kinst-Hori, I. Flavonols from saffron flower: tyrosinase inhibitory activity and inhibition mechanism. *J. Agric. Food Chem.*, **1999**, *47*, 4121-5.
- [464] Maciel, M.A.; Pinto, A.C.; Arruda, A.C.; Pamplona, S.G.; Vanderlinde, F.A.; Lapa, A.J.; Echevarria, A.; Grynberg, N.F.; Colus, I.M.; Farias, R.A.; Luna Costa, A.M.; Rao, V.S. Ethnopharmacology, phytochemistry and pharmacology: a successful combination in the study of *Croton cajucara*. *J. Ethnopharmacol.*, **2000**, *70*, 41-55.
- [465] Quintyne-Walcott, S.; Maxwell, A.R.; Reynolds, W.F. Crotagosamide, a cyclic nonapeptide from the latex of *Croton gossypifolius*. *J. Nat. Prod.*, **2007**, *70*, 1374-6.
- [466] Deng, J.Z.; Marshall, R.; Jones, S.H.; Johnson, R.K.; Hecht, S.M. DNA-damaging agents from *Crypteronia paniculata*. *J. Nat. Prod.*, **2002**, *65*, 1930-2.
- [467] Zou, Y.S.; Hou, A.J.; Zhu, G.F.; Chen, Y.F.; Sun, H.D.; Zhao, Q.S. Cytotoxic isoprenylated xanthenes from *Cudrania tricuspidata*. *Bioorg. Med. Chem.*, **2004**, *12*, 1947-53.
- [468] Lee, Y.J.; Kim, S.; Lee, S.J.; Ham, I.; Whang, W.K. Antioxidant activities of new flavonoids from *Cudrania tricuspidata* root bark. *Arch. Pharm. Res.*, **2009**, *32*, 195-200.
- [469] Ye, M.; Yan, Y.; Guo, D.A. Characterization of phenolic compounds in the Chinese herbal drug Tu-Si-Zi by liquid chromatography coupled to electrospray ionization mass spectrometry. *Rapid Commun. Mass Spectrom.*, **2005**, *19*, 1469-84.
- [470] Umehara, K.; Nemoto, K.; Ohkubo, T.; Miyase, T.; Degawa, M.; Noguchi, H. Isolation of a new 15-membered macrocyclic glycolipid lactone, Cuscutic Resinoid A from the seeds of *Cuscuta chinensis*: a stimulator of breast cancer cell proliferation. *Planta Med.*, **2004**, *70*, 299-304.
- [471] Harinantenaina Liva, R.R.; Kasai, R.; Yamasaki, K. Clerodane and labdane diterpene glycosides from a Malagasy endemic plant, *Cussonia racemosa*. *Phytochemistry*, **2002**, *60*, 339-43.
- [472] Liu, H.; Gao, Y.; Wang, K.; Hu, Z. Determination of active components in *Cynanchum chinense* R. Br. by capillary electrophoresis. *Biomed. Chromatogr.*, **2006**, *20*, 451-4.
- [473] Gamez, E.J.; Luyengi, L.; Lee, S.K.; Zhu, L.F.; Zhou, B.N.; Fong, H.H.; Pezzuto, J.M.; Kinghorn, A.D. Antioxidant flavonoid glycosides from *Daphniphyllum calycinum*. *J. Nat. Prod.*, **1998**, *61*, 706-8.
- [474] Sajeli, B.A.; Sahai, M.; Fujimoto, Y.; Asai, K.; Schneider, K.; Nicholson, G.; Suessmuth, R. A new kaempferol diglycoside from *Datura suaveolens* Humb. & Bonpl. ex. Willd. *Nat. Prod. Res.*, **2006**, *20*, 1231-6.
- [475] Diaz, J.G. and Herz, W. Acylated flavonol tetraglycosides from *Delphinium gracile*. *Phytochemistry*, **2010**, *71*, 463-8.
- [476] Mallavadhani, U.V.; Narasimhan, K.; Sudhakar, A.V.; Mahapatra, A.; Li, W.; van Breemen, R.B. Three new pentacyclic triterpenes and some flavonoids from the fruits of an Indian Ayurvedic plant *Dendrophthoe falcata* and their estrogen receptor binding activity. *Chem. Pharm. Bull. (Tokyo)*, **2006**, *54*, 740-4.
- [477] Li, M.M.; Wang, K.; Pan, Z.H.; Chen, X.Q.; Peng, L.Y.; Li, Y.; Cheng, X.; Zhao, Q.S. Two new sesquiterpene glucosides from *Dennstaedtia scabra* (Wall.) Moore. *Chem. Pharm. Bull. (Tokyo)*, **2009**, *57*, 1123-5.
- [478] Xu, L.R.; Wu, J.; Zhang, S. A new acylated flavonol glycoside from *Derris trifoliata*. *J. Asian Nat. Prod. Res.*, **2006**, *8*, 9-13.
- [479] Xu, L.R.; Zhou, P.; Zhi, Y.E.; Wu, J.; Zhang, S. Three new flavonol triglycosides from *Derris trifoliata*. *J. Asian Nat. Prod. Res.*, **2009**, *11*, 79-84.
- [480] Cordell, G.A.; Lyon, R.L.; Fong, H.H.; Benoit, P.S.; Farnsworth, N.R. Biological and phytochemical investigations of *Dianthus barbatus* cv. "China Doll" (Caryophyllaceae). *Lloydia*, **1977**, *40*, 361-3.
- [481] Lee, J.H.; Ku, C.H.; Baek, N.I.; Kim, S.H.; Park, H.W.; Kim, D.K. Phytochemical constituents from *Diodia teres*. *Arch. Pharm. Res.*, **2004**, *27*, 40-3.
- [482] Gao, H.; Kuroyanagi, M.; Wu, L.; Kawahara, N.; Yasuno, T.; Nakamura, Y. Antitumor-promoting constituents from *Dioscorea bulbifera* L. in JB6 mouse epidermal cells. *Biol. Pharm. Bull.*, **2002**, *25*, 1241-3.
- [483] Mvot, A.C.; Mbazoa, D.C.; Ephrem, N.A.; Tu, P.F.; Lei, L.D. New Coumarin Glycosides from the Leaves of *Diospyros crassiflora* (Hiern). *Fitoterapia*, **2010**, *81*, 873-7.
- [484] Chen, G.; Lu, H.; Wang, C.; Yamashita, K.; Manabe, M.; Meng, Z.; Xu, S.; Kodama, H. Effect of five flavonoid compounds isolated from leaves of *Diospyros kaki* on stimulus-induced superoxide generation and tyrosyl phosphorylation of proteins in human neutrophils. *Clin. Chim. Acta*, **2002**, *326*, 169-75.
- [485] Loizzo, M.R.; Said, A.; Tundis, R.; Hawas, U.W.; Rashed, K.; Menichini, F.; Frega, N.G.; Menichini, F. Antioxidant and antiproliferative activity of *Diospyros lotus* L. extract and isolated compounds. *Plant Foods Hum. Nutr.*, **2009**, *64*, 264-70.
- [486] de Carvalho, M.G.; Cranchi, D.C.; Kingston, D.G.; Werle, A.A. Proposed active constituents of *Dipladenia martiana*. *Phytother. Res.*, **2001**, *15*, 715-7.
- [487] Moreno, A.; Martín-Cordero, C.; Iglesias-Guerra, F.; Toro, M.V. Flavonoids from *Dorycnium rectum*. *Biochem. Syst. Ecol.*, **2002**, *30*, 73-4.
- [488] Moon, S.S.; Rahman, A.A.; Manir, M.; Jamal, A.; V Kaempferol glycosides and cardenolide glycosides, cytotoxic constituents from the seeds of *Draba nemorosa* (Brassicaceae). *Arch. Pharm. Res.*, **2010**, *33*, 1169-73.
- [489] Dai, L.M.; Zhao, C.C.; Jin, H.Z.; Tang, J.; Shen, Y.H.; Li, H.L.; Peng, C.Y.; Zhang, W.D. A new ferulic acid ester and other constituents from *Dracocephalum pterigrinum*. *Arch. Pharm. Res.*, **2008**, *31*, 1325-9.
- [490] Wang, X.L.; Wang, N.L.; Zhang, Y.; Gao, H.; Pang, W.Y.; Wong, M.S.; Zhang, G.; Qin, L.; Yao, X.S. Effects of eleven flavonoids from the osteoprotective fraction of *Drynaria fortunei* (KUNZE) J. SM. on osteoblastic proliferation using an osteoblast-like cell line. *Chem. Pharm. Bull. (Tokyo)*, **2008**, *56*, 46-51.
- [491] Jiang, R.W.; Zhou, J.R.; Hon, P.M.; Li, S.L.; Zhou, Y.; Li, L.L.; Ye, W.C.; Xu, H.X.; Shaw, P.C.; But, P.P. Lignans from *Dyosma versipellis* with inhibitory effects on prostate cancer cell lines. *J. Nat. Prod.*, **2007**, *70*, 283-6.

- [492] Singh, S.; Upadhyay, R.K.; Pandey, M.B.; Singh, J.P.; Pandey, V.B. Flavonoids from *Echinops echinatus*. *J. Asian Nat. Prod. Res.*, **2006**, *8*, 197-200.
- [493] Chien, M.M.; Svoboda, G.H.; Schiff, P.L., Jr.; Slatkin, D.J.; Knapp, J.E. Chemical constituents of *Echites hirsuta* (Apocynaceae). *J. Pharm. Sci.*, **1979**, *68*, 247-9.
- [494] Pattamadilok, D. and Suttisri, R. Seco-terpenoids and other constituents from *Elateriospermum tapos*. *J. Nat. Prod.*, **2008**, *71*, 292-4.
- [495] Wirasathien, L.; Pengsuparp, T.; Moriyasu, M.; Kawanishi, K.; Suttisri, R. Cytotoxic C-benzylated chalcone and other constituents of *Ellipeiopsis cherevensis*. *Arch. Pharm. Res.*, **2006**, *29*, 497-502.
- [496] Chen, C.C.; Huang, Y.L.; Sun, C.M.; Shen, C.C. New prenylflavones from the leaves of *Epimedium sagittatum*. *J. Nat. Prod.*, **1996**, *59*, 412-4.
- [497] Wang, G.J.; Tsai, T.H.; Lin, L.C. Prenylflavonol, acylated flavonol glycosides and related compounds from *Epimedium sagittatum*. *Phytochemistry*, **2007**, *68*, 2455-64.
- [498] Oh, H.; Kim, D.H.; Cho, J.H.; Kim, Y.C. Hepatoprotective and free radical scavenging activities of phenolic petrosins and flavonoids isolated from *Equisetum arvense*. *J. Ethnopharmacol.*, **2004**, *95*, 421-4.
- [499] Kanchanapoom, T.; Otsuka, H.; Ruchirawat, S. Megastigmane glucosides from *Equisetum debile* and *E. diffusum*. *Chem. Pharm. Bull. (Tokyo)*, **2007**, *55*, 1277-80.
- [500] Gurbuz, I.; Yesilada, E.; Ito, S. An anti-ulcerogenic flavonol diglucoside from *Equisetum palustre* L. *J. Ethnopharmacol.*, **2009**, *121*, 360-5.
- [501] Jung, H.A.; Park, J.C.; Chung, H.Y.; Kim, J.; Choi, J.S. Antioxidant flavonoids and chlorogenic acid from the leaves of *Eriobotrya japonica*. *Arch. Pharm. Res.*, **1999**, *22*, 213-8.
- [502] Amakura, Y.; Yoshimura, M.; Sugimoto, N.; Yamazaki, T.; Yoshida, T. Marker Constituents of the Natural Antioxidant Eucalyptus Leaf Extract for the Evaluation of Food Additives. *Biosci. Biotechnol. Biochem.*, **2009**, *73*, 1060-5.
- [503] Oksuz, S.; Gurek, F.; Lin, L.Z.; Gil, R.R.; Pezzuto, J.M.; Cordell, G.A. Aleppicatinines A and B from *Euphorbia aleppica*. *Phytochemistry*, **1996**, *42*, 473-8.
- [504] Nazemiyeh, H.; Kazemi, E.M.; Zare, K.; Jodari, M.; Nahar, L.; Sarker, S.D. Free radical scavengers from the aerial parts of *Euphorbia petiolata*. *J. Nat. Med.*, **2009**.
- [505] Lee, M.K.; Jeon, H.Y.; Lee, K.Y.; Kim, S.H.; Ma, C.J.; Sung, S.H.; Lee, H.S.; Park, M.J.; Kim, Y.C. Inhibitory constituents of *Euscaphis japonica* on lipopolysaccharide-induced nitric oxide production in BV2 microglia. *Planta Med.*, **2007**, *73*, 782-6.
- [506] Gupta, P.; Akanksha, S.; Siripurapu, K.B.; Ahmad, A.; Palit, G.; Arora, A.; Maurya, R. Anti-stress constituents of *Evolvulus alsinoides*: an ayurvedic crude drug. *Chem. Pharm. Bull. (Tokyo)*, **2007**, *55*, 771-5.
- [507] Kumar, M.; Ahmad, A.; Rawat, P.; Khan, M.F.; Rasheed, N.; Gupta, P.; Sathiamoorthy, B.; Bhatia, G.; Palit, G.; Maurya, R. Antioxidant flavonoid glycosides from *Evolvulus alsinoides*. *Fitoterapia*, **2010**, *81*, 234-42.
- [508] El Wakil, E.A. Phytochemical and molluscicidal investigations of *Fagonia arabica*. *Z. Naturforsch. [C.]*, **2007**, *62*, 661-7.
- [509] Ibrahim, L.F.; Kawashty, S.A.; El Hagrassy, A.M.; Nassar, M.I.; Mabry, T.J. A new kaempferol triglycoside from *Fagonia taekholmiana*: cytotoxic activity of its extracts. *Carbohydr. Res.*, **2008**, *343*, 155-8.
- [510] Tomczyk, M.; Gudej, J.; Sochacki, M. Flavonoids from *Ficaria verna* Huds. *Z. Naturforsch. [C.]*, **2002**, *57*, 440-4.
- [511] Ramadan, M.A.; Ahmad, A.S.; Nafady, A.M.; Mansour, A.I. Chemical composition of the stem bark and leaves of *Ficus pandurata* Hance. *Nat. Prod. Res.*, **2009**, *23*, 1218-30.
- [512] Parejo, I.; Viladomat, F.; Bastida, J.; Schmeda-Hirschmann, G.; Burillo, J.; Codina, C. Bioguided isolation and identification of the nonvolatile antioxidant compounds from fennel (*Foeniculum vulgare* Mill.) waste. *J. Agric. Food Chem.*, **2004**, *52*, 1890-7.
- [513] Xu, Y.M.; Smith, J.A.; Lannigan, D.A.; Hecht, S.M. Three acetylated flavonol glycosides from *Forsteronia refracta* that specifically inhibit p90 RSK. *Bioorg. Med. Chem.*, **2006**, *14*, 3974-7.
- [514] Tsukamoto, S.; Tomise, K.; Aburatani, M.; Onuki, H.; Hirorta, H.; Ishiharajima, E.; Ohta, T. Isolation of cytochrome P450 inhibitors from strawberry fruit, *Fragaria ananassa*. *J. Nat. Prod.*, **2004**, *67*, 1839-41.
- [515] Aaby, K.; Ekeberg, D.; Skrede, G. Characterization of phenolic compounds in strawberry (*Fragaria x ananassa*) fruits by different HPLC detectors and contribution of individual compounds to total antioxidant capacity. *J. Agric. Food Chem.*, **2007**, *55*, 4395-406.
- [516] Hussein, S.A. Flavonoid and methoxycinnamic acid sodium sulphates from *Frankenia laevis* L. *Pharmazie*, **2004**, *59*, 484-7.
- [517] Champavier, Y.; Allais, D.P.; Chulia, A.J.; Kaouadji, M. Acetylated and non-acetylated flavonol triglycosides from *Galega officinalis*. *Chem. Pharm. Bull. (Tokyo)*, **2000**, *48*, 281-2.
- [518] Gayosso-De-Lucio, J.A.; Torres-Valencia, J.M.; Cerda-Garcia-Rojas, C.M.; Joseph-Nathan, P. Ellagitannins from *Geranium potentillaefolium* and *G. bellum*. *Nat. Prod. Commun.*, **2010**, *5*, 531-4.
- [519] Li, J.; Huang, H.; Zhou, W.; Feng, M.; Zhou, P. Anti-hepatitis B virus activities of *Geranium carolinianum* L. extracts and identification of the active components. *Biol. Pharm. Bull.*, **2008**, *31*, 743-7.
- [520] Ho, H.M.; Chen, R.; Huang, Y.; Chen, Z.Y. Vascular effects of a soy leaves (*Glycine max*) extract and kaempferol glycosides in isolated rat carotid arteries. *Planta Med.*, **2002**, *68*, 487-91.
- [521] Hatano, T.; Yasuhara, T.; Fukuda, T.; Noro, T.; Okuda, T. Phenolic constituents of licorice. II. Structures of licopyranocoumarin, licoaryl coumarin and glisoflavone, and inhibitory effects of licorice phenolics on xanthine oxidase. *Chem. Pharm. Bull. (Tokyo)*, **1989**, *37*, 3005-9.
- [522] Ferrari, J.; Terreaux, C.; Sahpaz, S.; Msonthi, J.D.; Wolfender, J.L.; Hostettmann, K. Benzophenone glycosides from *Gnidia involucreta*. *Phytochemistry*, **2000**, *54*, 883-9.
- [523] Du, X.M.; Sun, N.Y.; Shoyama, Y. Flavonoids from *Goodyera schlechtendaliana*. *Phytochemistry*, **2000**, *53*, 997-1000.
- [524] Krenn, L.; Wollenweber, E.; Steyrlreuthner, K.; Gorick, C.; Melzig, M.F. Contribution of methylated exudate flavonoids to the anti-inflammatory activity of *Grindelia robusta*. *Fitoterapia*, **2009**, *80*, 267-9.
- [525] Liu, X.; Ye, W.; Yu, B.; Zhao, S.; Wu, H.; Che, C. Two new flavonol glycosides from *Gymnema sylvestris* and *Euphorbia ebracteolata*. *Carbohydr. Res.*, **2004**, *339*, 891-5.
- [526] Yin, F.; Zhang, Y.; Yang, Z.; Cheng, Q.; Hu, L. Triterpene saponins from *Gynostemma cardiospermum*. *J. Nat. Prod.*, **2006**, *69*, 1394-8.
- [527] Sala, A.; Recio, M.C.; Schinella, G.R.; Manez, S.; Giner, R.M.; Cerda-Nicolas, M.; Rosi, J.L. Assessment of the anti-inflammatory activity and free radical scavenger activity of tiliroside. *Eur. J. Pharmacol.*, **2003**, *461*, 53-61.
- [528] Vitalini, S.; Braca, A.; Fico, G. Study on secondary metabolite content of *Helleborus niger* L. leaves. *Fitoterapia*, **2011**, *82*, 152-4.
- [529] Cichewicz, R.H. and Nair, M.G. Isolation and characterization of stelladerol, a new antioxidant naphthalene glycoside, and other antioxidant glycosides from edible daylily (*hemerocallis*) flowers. *J. Agric. Food Chem.*, **2002**, *50*, 87-91.
- [530] Haraguchi, H.; Ishikawa, H.; Sanchez, Y.; Ogura, T.; Kubo, Y.; Kubo, I. Antioxidative constituents in *Heterotheca inuloides*. *Bioorg. Med. Chem.*, **1997**, *5*, 865-71.
- [531] Sharma, U.K.; Sharma, K.; Sharma, N.; Sharma, A.; Singh, H.P.; Sinha, A.K. Microwave-assisted efficient extraction of different parts of *Hippophae rhamnoides* for the comparative evaluation of antioxidant activity and quantification of its phenolic constituents by reverse-phase high-performance liquid chromatography (RP-HPLC). *J. Agric. Food Chem.*, **2008**, *56*, 374-9.
- [532] Budzianowski, J. Kaempferol glycosides from *Hosta ventricosa*. *Phytochemistry*, **1990**, *29*, 3643-7.
- [533] Yoshikawa, M.; Matsuda, H.; Shimoda, H.; Shimada, H.; Harada, E.; Naitoh, Y.; Miki, A.; Yamahara, J.; Murakami, N. Development of bioactive functions in *hydrangeae dulcis* folium. V. On the antiallergic and antimicrobial principles of *hydrangeae dulcis* folium. (2). Thunberginol C, D, and E, thunberginol G 3'-O-glucoside, (-)-hydrangenol 4'-o-glucoside, and (+)-hydrangenol 4'-O-glucoside. *Chem. Pharm. Bull. (Tokyo)*, **1996**, *44*, 1440-7.
- [534] Rocha, L.; Marston, A.; Potterat, O.; Kaplan, M.A.; Stoeckli-Evans, H.; Hostettmann, K. Antibacterial phloroglucinols and flavonoids from *Hypericum brasiliense*. *Phytochemistry*, **1995**, *40*, 1447-52.

- [535] Odabas, M.S.; Camas, N.; Cirak, C.; Radusiene, J.; Janulis, V.; Ivanauskas, L. The quantitative effects of temperature and light intensity on phenolics accumulation in St. John's wort (*Hypericum perforatum*). *Nat. Prod. Commun.*, **2010**, *5*, 535-40.
- [536] Xie, G.B.; Niu, F.; Wang, X.J.; Lei, L.D.; Tu, P.F. Chemical constituents from the leaves of *Ilex pernyi*. *Yao Xue. Xue. Bao.*, **2008**, *43*, 60-2.
- [537] Hua, L.; Peng, Z.; Chia, L.S.; Goh, N.K.; Tan, S.N. Separation of kaempferols in *Impatiens balsamina* flowers by capillary electrophoresis with electrochemical detection. *J. Chromatogr. A.*, **2001**, *909*, 297-303.
- [538] Ishiguro, K.; Ohira, Y.; Oku, H. Preventive effects of *Impatiens balsamina* on the hen egg-white lysozyme (HEL)-induced decrease in blood flow. *Biol. Pharm. Bull.*, **2002**, *25*, 505-8.
- [539] Ueda, Y.; Oku, H.; Iinuma, M.; Ishiguro, K. Effects on blood pressure decrease in response to PAF of *Impatiens textori* MIQ. *Biol. Pharm. Bull.*, **2003**, *26*, 1505-7.
- [540] Hasan, A.; Ahmad, I.; Khan, M.A.; Chudhary, M.I. Two flavonol triglycosides from flowers of *Indigofera hebeptala*. *Phytochemistry*, **1996**, *43*, 1115-8.
- [541] Calvo, T.R.; Cardoso, C.R.; Silva Moura, A.C.; dos Santos, L.C.; Colus, I.M.; Vilegas, W.; Varanda, E.A. Mutagenic Activity of *Indigofera truxillensis* and *I. suffruticosa* Aerial Parts. *Evid. Based. Complement Alternat. Med.*, **2009**, *20*.
- [542] Zhang, Y.; Zhao, L.; Shi, Y.P. Separation and determination of flavonoids in *Ixeridium gracile* by capillary electrophoresis. *J. Chromatogr. Sci.*, **2007**, *45*, 600-4.
- [543] Clarkson, C.; Staerk, D.; Hansen, S.H.; Jaroszewski, J.W. Hypheation of solid-phase extraction with liquid chromatography and nuclear magnetic resonance: application of HPLC-DAD-SPE-NMR to identification of constituents of *Kanahia laniflora*. *Anal. Chem.*, **2005**, *77*, 3547-53.
- [544] Matlawska, I. Flavonoid compounds in the flowers of *Kitaibelia vitifolia* Willd. (Malvaceae). *Acta Pol. Pharm.*, **2001**, *58*, 127-31.
- [545] Abou-Shoer, M.; Ma, G.E.; Li, X.H.; Koonchanok, N.M.; Geahlen, R.L.; Chang, C.J. Flavonoids from *Koeleruteria henryi* and other sources as protein-tyrosine kinase inhibitors. *J. Nat. Prod.*, **1993**, *56*, 967-9.
- [546] Lee, T.H.; Chiang, Y.H.; Chen, C.H.; Chen, P.Y.; Lee, C.K. A new flavonol galloylrhamnoside and a new lignan glucoside from the leaves of *Koeleruteria henryi* Dummer. *Nat. Med. (Tokyo)*, **2009**, *63*, 209-14.
- [547] Lin, W.H.; Deng, Z.W.; Lei, H.M.; Fu, H.Z.; Li, J. Polyphenolic compounds from the leaves of *Koeleruteria paniculata* Laxm. *J. Asian Nat. Prod. Res.*, **2002**, *4*, 287-95.
- [548] Kim, D.K. Antioxidative components from the aerial parts of *Lactuca scariola* L. *Arch. Pharm. Res.*, **2001**, *24*, 427-30.
- [549] Budzianowski, J. and Skrzypczak, L. Phenylpropanoid esters from *Lamium album* flowers. *Phytochemistry*, **1995**, *38*, 997-1001.
- [550] Nugroho, A.; Choi, J.K.; Park, J.H.; Lee, K.T.; Cha, B.C.; Park, H.J. Two New Flavonol Glycosides from *Lamium amplexicaule* L. and Their *in vitro* Free Radical Scavenging and Tyrosinase Inhibitory Activities. *Planta Med.*, **2009**, *75*, 358-9.
- [551] Skalicka-Wozniak, K.; Melliou, E.; Gortzi, O.; Glowniak, K.; Chinou, I.B. Chemical constituents of *Lavatera trimestris* L.--antioxidant and antimicrobial activities. *Z. Naturforsch. [C.]*, **2007**, *62*, 797-800.
- [552] Tasdemir, D.; Scapozza, L.; Zerbe, O.; Linden, A.; Calis, I.; Sticher, O. Iridoid glycosides of *Leonurus persicus*. *J. Nat. Prod.*, **1999**, *62*, 811-6.
- [553] Li, X.C.; Dunbar, D.C.; ElSohly, H.N.; Walker, L.A.; Clark, A.M. Indolopyridoquinazoline alkaloid from *Leptothyrsa sprucei*. *Phytochemistry*, **2001**, *58*, 627-9.
- [554] Braca, A.; Sortino, C.; Politi, M.; Morelli, I.; Mendez, J. Antioxidant activity of flavonoids from *Licania licaniaeflora*. *J. Ethnopharmacol.*, **2002**, *79*, 379-81.
- [555] Ouyang, M.A.; He, Z.D.; Wu, C.L. Anti-oxidative activity of glycosides from *Ligustrum sinense*. *Nat. Prod. Res.*, **2003**, *17*, 381-7.
- [556] Vachalkova, A.; Eisenreichova, E.; Haladova, M.; Mucaji, P.; Jozova, B.; Novotny, L. Potential carcinogenic and inhibitory activity of compounds isolated from *Lilium candidum* L. *Neoplasma.*, **2000**, *47*, 313-8.
- [557] Francis, J.A.; Rumbelha, W.; Nair, M.G. Constituents in Easter lily flowers with medicinal activity. *Life Sci.*, **2004**, *76*, 671-83.
- [558] Obmann, A.; Tsendayush, D.; Thalhammer, T.; Zehl, M.; Vo, T.P.; Purevsuren, S.; Natsagdorj, D.; Narantuya, S.; Kletter, C.; Glasl, S. Extracts from the Mongolian traditional medicinal plants *Dianthus versicolor* Fisch. and *Lilium pumilum* Delile stimulate bile flow in an isolated perfused rat liver model. *J. Ethnopharmacol.*, **2010**, *131*, 555-61.
- [559] Choi, C.W.; Jung, H.A.; Kang, S.S.; Choi, J.S. Antioxidant constituents and a new triterpenoid glycoside from *Flos Lonicerae*. *Arch. Pharm. Res.*, **2007**, *30*, 1-7.
- [560] el Mousallami, A.M.; Afifi, M.S.; Hussein, S.A. Acylated flavonol diglycosides from *Lotus polyphyllus*. *Phytochemistry*, **2002**, *60*, 807-11.
- [561] Christen, P. and Kapetanidis, I. Flavonoids from *Lycium halimifolium* L. *Planta Med.*, **1987**, *53*, 571-2.
- [562] Inbaraj, B.S.; Lu, H.; Kao, T.H.; Chen, B.H. Simultaneous determination of phenolic acids and flavonoids in *Lycium barbarum* Linnaeus by HPLC-DAD-ESI-MS. *J. Pharm. Biomed. Anal.*, **2010**, *51*, 549-56.
- [563] Achari, B.; Basu, K.; Saha, C.R.; Pakrashi, S.C. A New Triterpene Ester, an Anthraquinone and Other Constituents of the Fern *Lygodium flexuosum*. *Planta Med.*, **1986**, *52*, 329-30.
- [564] Rzadkowska-Bodalska, H. and Olechnowicz-Stepien, W. Flavonoids in the herb of yellow loosestrife (*Lysimachia vulgaris* L.). *Pol. J. Pharmacol. Pharm.*, **1975**, *27*, 345-8.
- [565] Lee, S.S.; Lin, H.C.; Chen, C.K. Acylated flavonol monorhamnosides, alpha-glucosidase inhibitors, from *Machilus philippinensis*. *Phytochemistry*, **2008**, *69*, 2347-53.
- [566] Jung, K.Y.; Oh, S.R.; Park, S.H.; Lee, I.S.; Ahn, K.S.; Lee, J.J.; Lee, H.K. Anti-complement activity of tiliroside from the flower buds of *Magnolia fargesii*. *Biol. Pharm. Bull.*, **1998**, *21*, 1077-8.
- [567] Matlawska, I. and Sikorska, M. Flavonoids from flowers of *Malva crispa* L. (Malvaceae). *Acta Pol. Pharm.*, **2004**, *61*, 65-8.
- [568] Stingl, C.; Knapp, H.; Winterhalter, P. 3,4-Dihydroxy-7,8-dihydro-beta-ionone 3-O-beta-D-glucopyranoside and other glycosidic constituents from apple leaves. *Nat. Prod. Lett.*, **2002**, *16*, 87-93.
- [569] Prawat, H.; Mahidol, C.; Ruchirawat, S.; Prawat, U.; Tuntiwachwut-tikul, P.; Tooptakong, U.; Taylor, W.C.; Pakawatchai, C.; Skeleton, B.W.; White, A.H. Cyanogenic and non-cyanogenic glycosides from *Manihot esculenta*. *Phytochemistry*, **1995**, *40*, 1167-73.
- [570] Vilegas, W.; Sanonmiya, M.; Rastrelli, L.; Pizza, C. Isolation and structure elucidation of two new flavonoid glycosides from the infusion of *Maytenus aquifolium* leaves. Evaluation of the antiulcer activity of the infusion. *J. Agric. Food Chem.*, **1999**, *47*, 403-6.
- [571] Leite, J.P.; Rastrelli, L.; Romussi, G.; Oliveira, A.B.; Vilegas, J.H.; Vilegas, W.; Pizza, C. Isolation and HPLC quantitative analysis of flavonoid glycosides from Brazilian beverages (*Maytenus ilicifolia* and *M. aquifolium*). *J. Agric. Food Chem.*, **2001**, *49*, 3796-801.
- [572] Tanaka, M.; Fujimori, T.; Uchida, I.; Yamaguchi, S.; Takeda, K. A malonylated anthocyanin and flavonols in blue *Meconopsis* flowers. *Phytochemistry*, **2001**, *56*, 373-6.
- [573] Shang, X.Y.; Wang, Y.H.; Li, C.; Zhang, C.Z.; Yang, Y.C.; Shi, J.G. Acetylated flavonol diglycosides from *Meconopsis quintuplinervia*. *Phytochemistry*, **2006**, *67*, 511-5.
- [574] Sirat, H.M.; Susanti, D.; Ahmad, F.; Takayama, H.; Kitajima, M. Amides, triterpene and flavonoids from the leaves of *Melastoma malabathricum* L. *J. Nat. Med.*, **2010**, *64*, 492-5.
- [575] Fiorentino, A.; D'Abrosca, B.; Pacifico, S.; Golino, A.; Mastellone, C.; Oriano, P.; Monaco, P. Reactive oxygen species scavenging activity of flavone glycosides from *Melilotus neapolitana*. *Molecules*, **2007**, *12*, 263-70.
- [576] Rodrigues, J.; Rinaldo, D.; dos Santos, L.C.; Vilegas, W. An unusual C6-C6' linked flavonoid from *Miconia cabucu* (Melastomataceae). *Phytochemistry*, **2007**, *68*, 1781-4.
- [577] Bisignano, G.; Sanogo, R.; Marino, A.; Aquino, R.; D'Angelo, V.; Germano, M.P.; De Pasquale, R.; Pizza, C. Antimicrobial activity of *Mitracarpus scaber* extract and isolated constituents. *Let. Appl. Microbiol.*, **2000**, *30*, 105-8.
- [578] Bashir, A.; Hamburger, M.; Gupta, M.P.; Solis, P.N.; Hostettmann, K. Flavonol glycosides from *Monnina sylvatica*. *Phytochemistry*, **1991**, *30*, 3781-4.
- [579] Cimanga, K.; De Bruyne, T.; Lasure, A.; Van Poel, B.; Pieters, L.; Vanden Berghe, D.; Vlietinck, A.; Kambu, K.; Tona, L. *In vitro* anticomplementary activity of constituents from *Morinda morindoides*. *J. Nat. Prod.*, **1995**, *58*, 372-8.

- [580] Cimanga, R.K.; Kambu, K.; Tona, L.; Hermans, N.; Apers, S.; Totte, J.; Pieters, L.; Vlietinck, A.J. Cytotoxicity and *in vitro* susceptibility of *Entamoeba histolytica* to *Morinda morindoides* leaf extracts and its isolated constituents. *J. Ethnopharmacol.*, **2006**, *107*, 83-90.
- [581] Cimanga, K.; De Bruyne, T.; Van Poel, B.; Ma, Y.; Claeys, M.; Pieters, L.; Kambu, K.; Tona, L.; Bakana, P.; Vanden Berghe, D.; Vlietinck, A.J. Complement-modulating properties of a kaempferol 7-O-rhamnosylsophoroside from the leaves of *Morinda morindoides*. *Planta Med.*, **1997**, *63*, 220-3.
- [582] Siddhuraju, P. and Becker, K. Antioxidant properties of various solvent extracts of total phenolic constituents from three different agroclimatic origins of drumstick tree (*Moringa oleifera* Lam.) leaves. *J. Agric. Food Chem.*, **2003**, *51*, 2144-55.
- [583] Manguro, L.O. and Lemmen, P. Phenolics of *Moringa oleifera* leaves. *Nat. Prod. Res.*, **2007**, *21*, 56-68.
- [584] Singh, B.N.; Singh, B.R.; Singh, R.L.; Prakash, D.; Dhakarey, R.; Upadhyay, G.; Singh, H.B. Oxidative DNA damage protective activity, antioxidant and anti-quorum sensing potentials of *Moringa oleifera*. *Food Chem. Toxicol.*, **2009**, *47*, 1109-16.
- [585] Kim, S.Y.; Gao, J.J.; Lee, W.C.; Ryu, K.S.; Lee, K.R.; Kim, Y.C. Antioxidative flavonoids from the leaves of *Morus alba*. *Arch. Pharm. Res.*, **1999**, *22*, 81-5.
- [586] Pothavorn, P.; Kitdamrongsont, K.; Swangpol, S.; Wongniam, S.; Atawongsa, K.; Savasti, J.; Somana, J. Sap phytochemical compositions of some bananas in Thailand. *J. Agric. Food Chem.*, **2010**, *58*, 8782-7.
- [587] Lim, S.S.; Jung, Y.J.; Hyun, S.K.; Lee, Y.S.; Choi, J.S. Rat lens aldose reductase inhibitory constituents of *Nelumbo nucifera* stem. *Phytother. Res.*, **2006**, *20*, 825-30.
- [588] Yang, J.H.; Kondratyuk, T.P.; Marler, L.E.; Qiu, X.; Choi, Y.; Cao, H.; Yu, R.; Sturdy, M.; Pegan, S.; Liu, Y.; Wang, L.Q.; Mesecar, A.D.; Breemen, R.B.; Pezzuto, J.M.; Fong, H.H.; Chen, Y.G.; Zhang, H.J. Isolation and evaluation of kaempferol glycosides from the fern *Neocheiropteris palmatopedata*. *Phytochemistry*, **2010**, *71*, 641-7.
- [589] Aung, H.H.; Chia, L.S.; Goh, N.K.; Chia, T.F.; Ahmed, A.A.; Pare, P.W.; Mabry, T.J. Phenolic constituents from the leaves of the carnivorous plant *Nepenthes gracilis*. *Fitoterapia*, **2002**, *73*, 445-7.
- [590] Ragasa, C.Y.; de Luna, R.D.; Cruz, W.C., Jr.; Rideout, J.A. Monoterpene lactones from the seeds of *Nephelium lappaceum*. *J. Nat. Prod.*, **2005**, *68*, 1394-6.
- [591] Pang, T.; Yuan, Z.; Dai, Y.; Wang, C.; Yang, J.; Peng, L.; Xu, G. Identification and determination of glycosides in tobacco leaves by liquid chromatography with atmospheric pressure chemical ionization tandem mass spectrometry. *J. Sep. Sci.*, **2007**, *30*, 289-96.
- [592] Zhang, Z.; ElSohly, H.N.; Li, X.C.; Khan, S.I.; Broedel, S.E., Jr.; Raulli, R.E.; Cihlar, R.L.; Burandt, C.; Walker, L.A. Phenolic compounds from *Nymphaea odorata*. *J. Nat. Prod.*, **2003**, *66*, 548-50.
- [593] Jayaprakasam, B.; Damu, A.G.; Rao, K.V.; Gunasekar, D.; Blond, A.; Bodo, B. 7-O-Methyltetrahydrochonaflavone, a new biflavanone from *Ochna beddomei*. *J. Nat. Prod.*, **2000**, *63*, 507-8.
- [594] Barakat, H.H.; el Mousallamy, A.M.; Souleman, A.M.; Awadalla, S. Flavonoids of *Ochradenus baccatus*. *Phytochemistry*, **1991**, *30*, 3777-9.
- [595] de Laurentis, N.; Stefanizzi, L.; Milillo, M.A.; Tantillo, G. Flavonoids from leaves of *Olea europaea* L. cultivars. *Ann. Pharm. Fr.*, **1998**, *56*, 268-73.
- [596] Lin, Y.L.; Shen, C.C.; Huang, Y.J.; Chang, Y.Y. Homoflavonoids from *Ophioglossum petiolatum*. *J. Nat. Prod.*, **2005**, *68*, 381-4.
- [597] Qiu, Y.; Chen, Y.; Pei, Y.; Matsuda, H.; Yoshikawa, M. Constituents with radical scavenging effect from *Opuntia dillenii*: structures of new alpha-pyrones and flavonol glycoside. *Chem. Pharm. Bull. (Tokyo)*, **2002**, *50*, 1507-10.
- [598] Lee, E.H.; Kim, H.J.; Song, Y.S.; Jin, C.; Lee, K.T.; Cho, J.; Lee, Y.S. Constituents of the stems and fruits of *Opuntia ficus-indica* var. *saboten*. *Arch. Pharm. Res.*, **2003**, *26*, 1018-23.
- [599] Chatzopoulou, A.; Karioti, A.; Gousiadou, C.; Lax, V., V; Kyriazopoulos, P.; Golegou, S.; Skaltsa, H. Depsides and Other Polar Constituents from *Origanum dictamnus* L. and Their *in vitro* Antimicrobial Activity in Clinical Strains. *J. Agric. Food Chem.*, **2010**, *26*, 6064-8.
- [600] Je, M.C.; Jung, W.J.; Lee, K.Y.; Kim, Y.C.; Sung, S.H. Calpain inhibitory flavonoids isolated from *Orostachys japonicus*. *J. Enzyme Inhib. Med. Chem.*, **2008**, *24*, 676-9.
- [601] Jiang, H.; Zhan, W.Q.; Liu, X.; Jiang, S.X. Antioxidant activities of extracts and flavonoid compounds from *Oxytropis falcate* Bunge. *Nat. Prod. Res.*, **2008**, *22*, 1650-6.
- [602] Wang, X.; Cheng, C.; Sun, Q.; Li, F.; Liu, J.; Zheng, C. Isolation and purification of four flavonoid constituents from the flowers of *Paeonia suffruticosa* by high-speed counter-current chromatography. *J. Chromatogr. A.*, **2005**, *20*, 127-31.
- [603] Schliemann, W.; Schneider, B.; Wray, V.; Schmidt, J.; Nimtz, M.; Porzel, A.; Bohm, H. Flavonols and an indole alkaloid skeleton bearing identical acylated glycosidic groups from yellow petals of *Papaver nudicaule*. *Phytochemistry*, **2006**, *67*, 191-201.
- [604] Saleem, M.; Kim, H.J.; Jin, C.; Lee, Y.S. Antioxidant caffeic acid derivatives from leaves of *Parthenocissus tricuspidata*. *Arch. Pharm. Res.*, **2004**, *27*, 300-4.
- [605] Abreu, P.M.; Matthew, S.; Gonzalez, T.; Vanickova, L.; Costa, D.; Gomes, A.; Segundo, M.A.; Fernandes, E. Isolation and identification of antioxidants from *Pedilanthus tithymaloides*. *Nat. Med. (Tokyo)*, **2008**, *62*, 67-70.
- [606] Williams, C.A.; Harborne, J.B.; Newman, M.; Greenham, J.; Eagles, J. Chrysin and other leaf exudate flavonoids in the genus *Pelargonium*. *Phytochemistry*, **1997**, *46*, 1349-53.
- [607] Masuda, T.; Iritani, K.; Yonemori, S.; Oyama, Y.; Takeda, Y. Isolation and antioxidant activity of galloyl flavonol glycosides from the seashore plant, *Pemphis acida*. *Biosci. Biotechnol. Biochem.*, **2001**, *65*, 1302-9.
- [608] Bloor, S.J. An antimicrobial kaempferol-diacetyl-rhamnoside from *Pentachondra pumila*. *Phytochemistry*, **1995**, *38*, 1033-5.
- [609] Park, S.H.; Oh, S.R.; Jung, K.Y.; Lee, I.S.; Ahn, K.S.; Kim, J.H.; Kim, Y.S.; Lee, J.J.; Lee, H.K. Acylated flavonol glycosides with anti-complement activity from *Persicaria lapathifolia*. *Chem. Pharm. Bull. (Tokyo)*, **1999**, *47*, 1484-6.
- [610] Simirgiotis, M.J. and Schmeda-Hirschmann, G. Direct identification of phenolic constituents in *Boldo Folium* (*Peumus boldus* Mol.) infusions by high-performance liquid chromatography with diode array detection and electrospray ionization tandem mass spectrometry. *J. Chromatogr. A.*, **2010**, *1217*, 443-9.
- [611] Zhang, Y.B.; Xu, X.J.; Liu, H.M. Chemical constituents from *Mahkota dewa*. *J. Asian Nat. Prod. Res.*, **2006**, *8*, 119-23.
- [612] Beninger, C.W. and Hosfield, G.L. Flavonol glycosides from *Montcalm* dark red kidney bean: implications for the genetics of seed coat color in *Phaseolus vulgaris* L. *J. Agric. Food Chem.*, **1999**, *47*, 4079-82.
- [613] Plumb, G.W.; Price, K.R.; Williamson, G. Antioxidant properties of flavonol glycosides from green beans. *Redox. Rep.*, **1999**, *4*, 123-7.
- [614] Leu, C.H.; Li, C.Y.; Yao, X.; Wu, T.S. Constituents from the leaves of *Phellodendron amurense* and their antioxidant activity. *Chem. Pharm. Bull. (Tokyo)*, **2006**, *54*, 1308-11.
- [615] Aboutabl, E.A.; Meselhy, M.R.; Afifi, M.S. Iridoids from *Phlomis aurea* Decne growing in Egypt. *Pharmazie*, **2002**, *57*, 646-7.
- [616] Delazar, A.; Sabzevari, A.; Mojarrah, M.; Nazemiyeh, H.; Esnaashari, S.; Nahar, L.; Razavi, S.M.; Sarker, S.D. Free-radical-scavenging principles from *Phlomis caucasica*. *Nat. Med. (Tokyo)*, **2008**, *62*, 464-6.
- [617] Sousa, M.; Ousingsawat, J.; Seitz, R.; Puntheeranurak, S.; Regalado, A.; Schmidt, A.; Grego, T.; Jansakul, C.; Amaral, M.D.; Schreiber, R.; Kunzelmann, K. An extract from the medicinal plant *Phyllanthus acidus* and its isolated compounds induce airway chloride secretion: A potential treatment for cystic fibrosis. *Mol. Pharmacol.*, **2007**, *71*, 366-76.
- [618] Habib, u.R.; Yasin, K.A.; Choudhary, M.A.; Khaliq, N.; Atta, u.R.; Choudhary, M.I.; Malik, S. Studies on the chemical constituents of *Phyllanthus emblica*. *Nat. Prod. Res.*, **2007**, *20*, 775-81.
- [619] Bylka, W. and Matlowska, I. Flavonoids and free phenolic acids from *Phytolacca americana* L. leaves. *Acta Pol. Pharm.*, **2001**, *58*, 69-72.
- [620] Jung, M.J.; Jung, H.A.; Kang, S.S.; Hwang, G.S.; Choi, J.S. A new abietic acid-type diterpene glucoside from the needles of *Pinus densiflora*. *Arch. Pharm. Res.*, **2009**, *32*, 1699-704.
- [621] Tomaino, A.; Martorana, M.; Arcoraci, T.; Monteleone, D.; Giovino, C.; Saija, A. Antioxidant activity and phenolic profile

- of pistachio (*Pistacia vera* L., variety Bronte) seeds and skins. *Biochimie*, **2010**, *92*, 1115-22.
- [622] Ferreres, F.; Esteban, E.; Carpena-Ruiz, R.; Jimenez, M.A.; Tomas-Barberan, F.A. Acylated flavonol sophorotriosides from pea shoots. *Phytochemistry*, **1995**, *39*, 1443-6.
- [623] Crublet, M.L.; Long, C.; Sevenet, T.; Hadi, H.A.; Lavaud, C. Acylated flavonol glycosides from leaves of *Planchonia grandis*. *Phytochemistry*, **2003**, *64*, 589-94.
- [624] Ibrahim, M.A.; Mansoor, A.A.; Gross, A.; Ashfaq, M.K.; Jacob, M.; Khan, S.I.; Hamann, M.T. Methicillin-resistant *Staphylococcus aureus* (MRSA)-active metabolites from *Platanus occidentalis* (American Sycamore). *J Nat. Prod.*, **2009**, *72*, 2141-4.
- [625] Mitrokovska, D.; Mitaku, S.; Demetzos, C.; Harvala, C.; Mentis, A.; Perez, S.; Kokkinopoulos, D. Bioactive compounds from the buds of *Platanus orientalis* and isolation of a new kaempferol glycoside. *Planta Med.*, **1993**, *59*, 517-20.
- [626] Luo, Y.G.; Li, B.G.; Zhang, G.L. Four new glycosides from *Pleurospermum franchetianum*. *J. Asian Nat. Prod. Res.*, **2002**, *4*, 155-63.
- [627] Wang, G.J.; Chen, Y.M.; Wang, T.M.; Lee, C.K.; Chen, K.J.; Lee, T.H. Flavonoids with iNOS inhibitory activity from *Pogonatherum crinitum*. *J. Ethnopharmacol.*, **2008**, *19*:118, 71-8.
- [628] Kim, S.H.; Jang, S.D.; Lee, K.Y.; Sung, S.H.; Kim, Y.C. Chemical constituents isolated from *Polygala japonica* leaves and their inhibitory effect on nitric oxide production *in vitro*. *J. Enzyme Inhib. Med. Chem.*, **2009**, *24*, 230-3.
- [629] Liu, G.; Wang, W.; Masuoka, N.; Isobe, T.; Yamashita, K.; Manabe, M.; Kodama, H. Effect of three flavonoids isolated from Japanese *Polygonum* species on superoxide generation in human neutrophils. *Planta Med.*, **2005**, *71*, 933-7.
- [630] Smolarz, H.D.; Budzianowski, J.; Bogucka-Kocka, A.; Kocki, J.; Mendyk, E. Flavonoid glucuronides with anti-leukaemic activity from *Polygonum amphibium* L. *Phytochem. Anal.*, **2008**, *19*, 506-13.
- [631] Calis, I.; Kuruuzum, A.; Demirezer, L.O.; Sticher, O.; Ganci, W.; Ruedi, P. Phenylvaleric acid and flavonoid glycosides from *Polygonum salicifolium*. *J. Nat. Prod.*, **1999**, *62*, 1101-5.
- [632] Vasange, M.; Liu, B.; Welch, C.J.; Rolfsen, W.; Bohlin, L. The flavonoid constituents of two *Polypodium* species (*Calaguala*) and their effect on the elastase release in human neutrophils. *Planta Med.*, **1997**, *63*, 511-7.
- [633] Marzouk, M.S.; Ibrahim, M.T.; El Gindi, O.R.; Abou Bakr, M.S. Isoflavonoid glycosides and rotenoids from *Pongamia pinnata* leaves. *Z. Naturforsch. [C. J.]*, **2008**, *63*, 1-7.
- [634] Zhang, X.; Hung, T.M.; Phuong, P.T.; Ngoc, T.M.; Min, B.S.; Song, K.S.; Seong, Y.H.; Bae, K. Anti-inflammatory activity of flavonoids from *Populus davidiana*. *Arch. Pharm. Res.*, **2006**, *29*, 1102-8.
- [635] Fico, G.; Rodondi, G.; Flamini, G.; Passarella, D.; Tome, F. Comparative phytochemical and morphological analyses of three Italian *Primula* species. *Phytochemistry*, **2007**, *68*, 1683-91.
- [636] Sang, S.; Lapsley, K.; Jeong, W.S.; Lachance, P.A.; Ho, C.T.; Rosen, R.T. Antioxidative phenolic compounds isolated from almond skins (*Prunus amygdalus* Batsch). *J. Agric. Food Chem.*, **2002**, *50*, 2459-63.
- [637] Jung, H.A.; Jung, M.J.; Kim, J.Y.; Chung, H.Y.; Choi, J.S. Inhibitory activity of flavonoids from *Prunus davidiana* and other flavonoids on total ROS and hydroxyl radical generation. *Arch. Pharm. Res.*, **2003**, *26*, 809-15.
- [638] Olszewska, M. High-performance liquid chromatographic identification of flavonoid monoglycosides from *Prunus serotina* Ehrh. *Acta Pol. Pharm.*, **2005**, *62*, 435-41.
- [639] Jung, H.A.; Kim, A.R.; Chung, H.Y.; Choi, J.S. *In vitro* antioxidant activity of some selected *Prunus* species in Korea. *Arch. Pharm. Res.*, **2002**, *25*, 865-72.
- [640] Olszewska, M. and Wolbis, M. Flavonoids from the flowers of *Prunus spinosa* L. *Acta Pol. Pharm.*, **2001**, *58*, 367-72.
- [641] Olszewska, M. and Wolbis, M. Further flavonoids from the flowers of *Prunus spinosa* L. *Acta Pol. Pharm.*, **2002**, *59*, 133-7.
- [642] Chen, K.C.; Chuang, C.M.; Lin, L.Y.; Chiu, W.T.; Wang, H.E.; Hsieh, C.L.; Tsai, T.; Peng, R.Y. The polyphenolics in the aqueous extract of *Psidium guajava* kinetically reveal an inhibition model on LDL glycation. *Pharm. Biol.*, **2010**, *48*, 23-31.
- [643] Chen, Y.; Zhao, Y.; Hu, Y.; Wang, L.; Ding, Z.; Liu, Y.; Wang, J. Isolation of 5-hydroxyppyrolidin-2-one and other constituents from the young fronds of *Pteridium aquilinum*. *Nat. Med. (Tokyo)*, **2008**, *62*, 358-9.
- [644] Regasini, L.O.; Velloso, J.C.; Silva, D.H.; Furlan, M.; de Oliveira, O.M.; Khalil, N.M.; Brunetti, I.L.; Young, M.C.; Barreiro, E.J.; Bolzani, V.S. Flavonols from *Pterogyne nitens* and their evaluation as myeloperoxidase inhibitors. *Phytochemistry*, **2008**, *69*, 1739-44.
- [645] van Elswijk, D.A.; Schobel, U.P.; Lansky, E.P.; Irth, H.; van der, G.J. Rapid dereplication of estrogenic compounds in pomegranate (*Punica granatum*) using on-line biochemical detection coupled to mass spectrometry. *Phytochemistry*, **2004**, *65*, 233-41.
- [646] Falodun, A.; Qadir, M.I.; Chouldary, M.I. Isolation and characterization of xanthine oxidase inhibitory constituents of *Pyrenacantha staudtii*. *Yao Xue. Xue. Bao.*, **2009**, *44*, 390-4.
- [647] Wang, N.; Wang, J.H.; Li, X.; Ling, J.H.; Li, N. Flavonoids from *Pyrrosia petiolosa* (Christ) Ching. *J. Asian Nat. Prod. Res.*, **2006**, *8*, 753-6.
- [648] Rychlinska, I. and Gudej, J. Flavonoid compounds from *Pyrus communis* L. flowers. *Acta Pol. Pharm.*, **2002**, *59*, 53-6.
- [649] Meng, Z.; Zhou, Y.; Lu, J.; Sugahara, K.; Xu, S.; Kodama, H. Effect of five flavonoid compounds isolated from *Quercus dentata* Thunb on superoxide generation in human neutrophils and phosphorylation of neutrophil proteins. *Clin. Chim. Acta*, **2001**, *306*, 97-102.
- [650] Sahpaz, S.; Gupta, M.P.; Hostettmann, K. Triterpene saponins from *Randia formosa*. *Phytochemistry*, **2000**, *54*, 77-84.
- [651] el Sayed, N.H.; Omara, N.M.; Yousef, A.K.; Farag, A.M.; Mabry, T.J. Kaempferol triosides from *Reseda muricata*. *Phytochemistry*, **2001**, *57*, 575-8.
- [652] Wei, B.L.; Lu, C.M.; Tsao, L.T.; Wang, J.P.; Lin, C.N. *In vitro* anti-inflammatory effects of quercetin 3-O-methyl ether and other constituents from *Rhamnus* species. *Planta Med.*, **2001**, *67*, 745-7.
- [653] Pandey, M.B.; Singh, A.K.; Singh, U.; Singh, S.; Pandey, V.B. A new chalcone glycoside from *Rhamnus nipalensis*. *Nat. Prod. Res.*, **2008**, *22*, 1657-9.
- [654] Ozipek, M.; Calis, I.; Ertan, M.; Ruedi, P. Rhamnetin 3-p-coumaroylrhamnoside from *Rhamnus petiolaris*. *Phytochemistry*, **1994**, *37*, 249-53.
- [655] Goel, R.K.; Pandey, V.B.; Dwivedi, S.P.; Rao, Y.V. Antiinflammatory and antiulcer effects of kaempferol, a flavone, isolated from *Rhamnus procumbens*. *Indian J. Exp. Biol.*, **1988**, *26*, 121-4.
- [656] Satake, T.; Hori, K.; Kamiya, K.; Saiki, Y.; Fujimoto, Y.; Kimura, Y.; Maksud, C.; Mekin, T. Studies on the constituents of Turkish plants. I. Flavonol triglycosides from the fruit of *Rhamnus thymifolius*. *Chem. Pharm. Bull. (Tokyo)*, **1993**, *41*, 1743-5.
- [657] Iwashina, T.; Omori, Y.; Kitajima, J.; Akiyama, S.; Suzuki, T.; Ohba, H. Flavonoids in translucent bracts of the Himalayan *Rheum nobile* (Polygonaceae) as ultraviolet shields. *J. Plant Res.*, **2004**, *117*, 101-7.
- [658] Song, E.K.; Kim, J.H.; Kim, J.S.; Cho, H.; Nan, J.X.; Sohn, D.H.; Ko, G.I.; Oh, H.; Kim, Y.C. Hepatoprotective phenolic constituents of *Rhodiola sachalinensis* on tacrine-induced cytotoxicity in Hep G2 cells. *Phytother. Res.*, **2003**, *17*, 563-5.
- [659] Lee, M.W.; Lee, Y.A.; Park, H.M.; Toh, S.H.; Lee, E.J.; Jang, H.D.; Kim, Y.H. Antioxidative phenolic compounds from the roots of *Rhodiola sachalinensis* A. Bor. *Arch. Pharm. Res.*, **2000**, *23*, 455-8.
- [660] Fan, W.; Tezuka, Y.; Ni, K.M.; Kadota, S. Prolyl endopeptidase inhibitors from the underground part of *Rhodiola sachalinensis*. *Chem. Pharm. Bull. (Tokyo)*, **2001**, *49*, 396-401.
- [661] Sandell, M.; Laaksonen, O.; Jarvinen, R.; Rostiala, N.; Pohjanheimo, T.; Tiitinen, K.; Kallio, H. Orosensory profiles and chemical composition of black currant (*Ribes nigrum*) juice and fractions of press residue. *J. Agric. Food Chem.*, **2009**, *57*, 3718-28.
- [662] Chin, Y.W. and Kim, J. Three new flavonol glycosides from the aerial parts of *Rodgersia podophylla*. *Chem. Pharm. Bull. (Tokyo)*, **2006**, *54*, 234-6.
- [663] Nowak, R. and Gawlik-Dziki, U. Polyphenols of *Rosa* L. leaves extracts and their radical scavenging activity. *Z. Naturforsch. [C. J.]*, **2007**, *62*, 32-8.
- [664] Ninomiya, K.; Matsuda, H.; Kubo, M.; Morikawa, T.; Nishida, N.; Yoshikawa, M. Potent anti-obese principle from *Rosa canina*: structural requirements and mode of action of trans-tiliroside. *Bioorg. Med. Chem. Lett.*, **2007**, *17*, 3059-64.

- [665] Schiber, A.; Mihalev, K.; Berardini, N.; Mollov, P.; Carle, R. Flavonol glycosides from distilled petals of *Rosa damascena* Mill. *Z. Naturforsch. [C.]*, **2005**, *60*, 379-84.
- [666] Bai, N.; He, K.; Roller, M.; Lai, C.S.; Shao, X.; Pan, M.H.; Ho, C.T. Flavonoids and phenolic compounds from *Rosmarinus officinalis*. *J. Agric. Food Chem.*, **2010**, *58*, 5363-7.
- [667] Zafrilla, P.; Ferreres, F.; Tomas-Barberan, F.A. Effect of processing and storage on the antioxidant ellagic acid derivatives and flavonoids of red raspberry (*Rubus idaeus*) jams. *J. Agric. Food Chem.*, **2001**, *49*, 3651-5.
- [668] Gudej, J. Kaempferol and quercetin glycosides from *Rubus idaeus* L. leaves. *Acta Pol. Pharm.*, **2003**, *60*, 313-5.
- [669] Badr, A.M.; El Demerdash, E.; Khalifa, A.E.; Ghoneim, A.I.; Ayoub, N.A.; Abdel-Naim, A.B. *Rubus sanctus* protects against carbon tetrachloride-induced toxicity in rat isolated hepatocytes: isolation and characterization of its galloylated flavonoids. *J. Pharm. Pharmacol.*, **2009**, *61*, 1511-20.
- [670] Panizzi, L.; Caponi, C.; Catalano, S.; Cioni, P.L.; Morelli, I. *In vitro* antimicrobial activity of extracts and isolated constituents of *Rubus ulmifolius*. *J. Ethnopharmacol.*, **2002**, *79*, 165-8.
- [671] Martini, S.; D'Addario, C.; Colacevich, A.; Focardi, S.; Borghini, F.; Santucci, A.; Figura, N.; Rossi, C. Antimicrobial activity against *Helicobacter pylori* strains and antioxidant properties of blackberry leaves (*Rubus ulmifolius*) and isolated compounds. *Int. J. Antimicrob. Agents*, **2009**, *34*, 50-9.
- [672] Hasan, A.; Ahmed, I.; Jay, M.; Voirin, B. Flavonoid glycosides and an anthraquinone from *Rumex chalepensis*. *Phytochemistry*, **1995**, *39*, 1211-3.
- [673] Jang, D.S.; Kim, J.M.; Kim, J.; Yoo, J.L.; Kim, Y.S.; Kim, J.S. Effects of compounds isolated from the fruits of *Rumex japonicus* on the protein glycation. *Chem. Biodivers.*, **2008**, *5*, 2718-23.
- [674] Chung, S.K.; Kim, Y.C.; Takaya, Y.; Terashima, K.; Niwa, M. Novel flavonol glycoside, 7-O-methyl mearnsitrin, from *Sageretia theezans* and its antioxidant effect. *J. Agric. Food Chem.*, **2004**, *52*, 4664-8.
- [675] Xu, C.L.; Zheng, Y.N.; Yang, X.W.; Li, X.G.; Li, X.; Chen, Q.C. Raddeanalin, a new flavonoid glycoside from the leaves of *Salix raddeana* Laksh. *J. Asian Nat. Prod. Res.*, **2007**, *9*, 415-9.
- [676] Schmitzer, V.; Veberic, R.; Slatnar, A.; Stampar, F. Elderberry (*Sambucus nigra* L.) Wine: A Product Rich in Health Promoting Compounds. *J. Agric. Food Chem.*, **2010**, *58*, 10143-6.
- [677] el Mousallamy, A.M. Polyphenols of Egyptian Rosaceae plants--two new flavonoid glycosides from *Sanguisorba minor* Scop. *Pharmazie*, **2002**, *57*, 702-4.
- [678] Christopoulou, C.; Graikou, K.; Chinou, I. Chemosystematic value of chemical constituents from *Scabiosa hymettia* (Dipsacaceae). *Chem. Biodivers.*, **2008**, *5*, 318-23.
- [679] Nowak, S. and Wolbis, M. Flavonoids from some species of genus *Scopolia* Jacq. *Acta Pol. Pharm.*, **2002**, *59*, 275-80.
- [680] Wolbis, M.; Nowak, S.; Kicel, A. Polyphenolic compounds in *Scopolia caucasica* Kolesn. ex Kreyer (Solanaceae). *Acta Pol. Pharm.*, **2007**, *64*, 241-6.
- [681] Calis, I.; Zor, M.; Basaran, A.A.; Wright, A.D.; Sticher, O. Karsoside and scropolioside D, two new iridoid glycosides from *Scrophularia ilwensis*. *J. Nat. Prod.*, **1993**, *56*, 606-9.
- [682] Penna, C.; Marino, S.; Vivot, E.; Cruanes, M.C.; de, D.M.; Cruanes, J.; Ferraro, G.; Gutkind, G.; Martino, V. Antimicrobial activity of Argentine plants used in the treatment of infectious diseases. Isolation of active compounds from *Sebastiania brasiliensis*. *J. Ethnopharmacol.*, **2001**, *77*, 37-40.
- [683] Ali, A.A.; Mohamed, M.H.; Kamel, M.S.; Fouad, M.A.; Spring, O. Studies on *Securigera securidacea* (L.) Deg. et Dorfl. (Fabaceae) seeds, an antidiabetic Egyptian folk medicine. *Pharmazie*, **1998**, *53*, 710-5.
- [684] Sanogo, R.; Vassallo, A.; Malafrente, N.; Imperato, S.; Russo, A.; Dal Piaz, F. New phenolic glycosides from *Securinega virosa* and their antioxidant activity. *Nat. Prod. Commun.*, **2009**, *4*, 1645-50.
- [685] Wang, W.S.; Lu, P.; Duan, C.H.; Feng, J.C. A new jacaranone derivative from *Senecio scandens* var. *incisus*. *Nat. Prod. Res.*, **2010**, *24*, 370-4.
- [686] el Sayed, N.H.; Wojcinska, M.; Drost-Karbowska, K.; Matlawska, I.; Williams, J.; Mabry, T.J. Kaempferol triosides from *Silphium perfoliatum*. *Phytochemistry*, **2002**, *60*, 835-8.
- [687] Arriaga, A.C.; de Mesquita, A.C.; Pouliquen, Y.B.; de Lima, R.A.; Cavalcante, S.H.; de Carvalho, M.G.; de Siqueira, J.A.; Alegrio, L.V.; Braz-Filho, R. Chemical constituents of *Simarouba versicolor*. *An. Acad. Bras. Cienc.*, **2002**, *74*, 415-24.
- [688] Leitao, G.G.; Simas, N.K.; Soares, S.S.; de Brito, A.P.; Claros, B.M.; Brito, T.B.; Delle, M.F. Chemistry and pharmacology of Monimiaceae: a special focus on *Siparuna* and *Mollinedia*. *J. Ethnopharmacol.*, **1999**, *65*, 87-102.
- [689] Leitao, G.G.; Soares, S.S.; Brito, T.D.; Delle, M.F. Kaempferol glycosides from *Siparuna apiosyce*. *Phytochemistry*, **2000**, *55*, 679-82.
- [690] Li, D.; Ikeda, T.; Matsuoka, N.; Nohara, T.; Zhang, H.; Sakamoto, T.; Nonaka, G. Cucurbitane glycosides from unripe fruits of *Lo Han Kuo* (*Siraitia grosvenori*). *Chem. Pharm. Bull. (Tokyo)*, **2006**, *54*, 1425-8.
- [691] Xu, J.; Li, X.; Zhang, P.; Li, Z.L.; Wang, Y. Antiinflammatory constituents from the roots of *Smilax bockii* warb. *Arch. Pharm. Res.*, **2005**, *28*, 395-9.
- [692] Slimestad, R. and Verheul, M.J. Seasonal variations in the level of plant constituents in greenhouse production of cherry tomatoes. *J. Agric. Food Chem.*, **2005**, *20*, 3114-9.
- [693] Chassy, A.W.; Bui, L.; Renaud, E.N.; Van Horn, M.; Mitchell, A.E. Three-year comparison of the content of antioxidant microconstituents and several quality characteristics in organic and conventionally managed tomatoes and bell peppers. *J. Agric. Food Chem.*, **2006**, *54*, 8244-52.
- [694] Ferreres, F.; Taveira, M.; Pereira, D.M.; Valentao, P.; Andrade, B. Tomato (*Lycopersicon esculentum*) seeds: new flavonols and cytotoxic effect. *J. Agric. Food Chem.*, **2010**, *58*, 2854-61.
- [695] Huang, H.C.; Syu, K.Y.; Lin, J.K. Chemical composition of *Solanum nigrum* linn extract and induction of autophagy by leaf water extract and its major flavonoids in AU565 breast cancer cells. *J. Agric. Food Chem.*, **2010**, *58*, 8699-708.
- [696] Kamel, M.S.; Ohtani, K.; Hasanain, H.A.; Mohamed, M.H.; Kasai, R.; Yamasaki, K. Monoterpene and pregnane glucosides from *Solenostemma argel*. *Phytochemistry*, **2000**, *53*, 937-40.
- [697] Wu, B.; Takahashi, T.; Kashiwagi, T.; Tebayashi, S.; Kim, C.S. New flavonoid glycosides from the leaves of *Solidago altissima*. *Chem. Pharm. Bull. (Tokyo)*, **2007**, *55*, 815-6.
- [698] Choi, S.Z.; Choi, S.U.; Lee, K.R. Phytochemical constituents of the aerial parts from *Solidago virga-aurea* var. *gigantea*. *Arch. Pharm. Res.*, **2004**, *27*, 164-8.
- [699] Tang, Y.; Lou, F.; Wang, J.; Zhuang, S. Four new isoflavone triglycosides from *Sophora japonica*. *J. Nat. Prod.*, **2001**, *64*, 1107-10.
- [700] Tang, Y.P.; Li, Y.F.; Hu, J.; Lou, F.C. Isolation and identification of antioxidants from *Sophora japonica*. *J. Asian Nat. Prod. Res.*, **2002**, *4*, 123-8.
- [701] Tang, Y.P.; Hu, J.; Wang, J.H.; Lou, F.C. A new coumaronochromone from *Sophora japonica*. *J. Asian Nat. Prod. Res.*, **2002**, *4*, 1-5.
- [702] Wang, J.H.; Lou, F.C.; Wang, Y.L.; Tang, Y.P. A flavonol tetraglycoside from *Sophora japonica* seeds. *Phytochemistry*, **2003**, *63*, 463-5.
- [703] Sun, A.; Sun, Q.; Liu, R. Preparative isolation and purification of flavone compounds from *sophora japonica* L. by high-speed counter-current chromatography combined with macroporous resin column separation. *J. Sep. Sci.*, **2007**, *30*, 1013-8.
- [704] Choudhary, M.I.; Naheed, N.; Abbaskhan, A.; Ali, S.; Atta, u.R. Hemiterpene glucosides and other constituents from *Spiraea canescens*. *Phytochemistry*, **2009**, *70*, 1467-73.
- [705] Dong, M.L.; Chen, G.; Zhou, Z.M. Flavonoid constituents from *Spiranthes australis* LINDL. *Chem. Pharm. Bull. (Tokyo)*, **2008**, *56*, 1600-3.
- [706] Liu, H.; Orjala, J.; Sticher, O.; Rali, T. Acylated flavonol glycosides from leaves of *Stenochlaena palustris*. *J. Nat. Prod.*, **1999**, *62*, 70-5.
- [707] Ahmad, V.U.; Bader, S.; Arshad, S.; Iqbal, S.; Ahmed, A.; Mohammad, F.V.; Kann, A.; Tareen, R.B. A new acylated flavone glycoside from the fruits of *Stocksia brauhica*. *J. Asian Nat. Prod. Res.*, **2007**, *9*, 299-305.
- [708] Ahmad, V.U.; Bader, S.; Arshad, S.; Ahmed, A.; Khan, A.; Iqbal, S.; Rasheed, M.; Tareen, R.B. Brauheneferoside E and F: acylated flavonol glycosides from *Stocksia brauhica* Linn. *Magn Reson. Chem.*, **2010**, *48*, 304-8.
- [709] Fu, X.; Li, X.C.; Wang, Y.H.; Avula, B.; Smillie, T.J.; Mabusela, W.; Syce, J.; Johnson, Q.; Folk, W.; Khan, I.A. Flavonol glycosides

- from the south African medicinal plant *Sutherlandia frutescens*. *Planta Med.*, **2010**, *76*, 178-81.
- [710] Whang, W.K. and Lee, M.T. New flavonol glycosides from leaves of *Symplocarpus renifolius*. *Arch. Pharm. Res.*, **1999**, *22*, 423-9.
- [711] Cai, L. and Wu, C.D. Compounds from *Syzygium aromaticum* possessing growth inhibitory activity against oral pathogens. *J. Nat. Prod.*, **1996**, *59*, 987-90.
- [712] Xiang, W.; Li, R.T.; Mao, Y.L.; Zhang, H.J.; Li, S.H.; Song, Q.S.; Sun, H.D. Four new prenylated isoflavonoids in *Tadehagi triquetrum*. *J. Agric. Food Chem.*, **2005**, *53*, 267-71.
- [713] Abouzid, S.F.; Ali, S.A.; Choudhary, M.I. A new ferulic acid ester and other constituents from *Tamarix nilotica* leaves. *Chem. Pharm. Bull. (Tokyo)*, **2009**, *57*, 740-2.
- [714] Hari, K.P.; Vijaya Bhaskar, R.M.; Gunasekar, D.; Marthanda, M.M.; Caux, C.; Bodo, B. A new coumestan from *Tephrosia calophylla*. *Chem. Pharm. Bull. (Tokyo)*, **2003**, *51*, 194-6.
- [715] Marzouk, M.S.; El Toumy, S.A.; Moharram, F.A.; Shalaby, N.M.; Ahmed, A.A. Pharmacologically active ellagitannins from *Terminalia myriocarpa*. *Planta Med.*, **2002**, *68*, 523-7.
- [716] Jo, Y.; Kim, M.; Shin, M.H.; Chung, H.Y.; Jung, J.H.; Im, K.S. Antioxidative phenolics from the fresh leaves of *Ternstroemia japonica*. *J. Nat. Prod.*, **2006**, *69*, 1399-403.
- [717] Ho, J.C.; Chen, C.M.; Row, L.C. Flavonoids and benzene derivatives from the flowers and fruit of *Tetrapanax papyriferus*. *J. Nat. Prod.*, **2005**, *68*, 1773-5.
- [718] Yang, H.; Protiva, P.; Cui, B.; Ma, C.; Baggett, S.; Hequet, V.; Mori, S.; Weinstein, I.B.; Kennelly, E.J. New bioactive polyphenols from *Theobroma grandiflorum* ("cupuacu"). *J. Nat. Prod.*, **2003**, *66*, 1501-4.
- [719] Parveen, Z.; Deng, Y.; Saeed, M.K.; Dai, R.; Ahamad, W.; Yu, Y.H. Antiinflammatory and analgesic activities of *Thesium chinense* Turcz extracts and its major flavonoids, kaempferol and kaempferol-3-O-glucoside. *Yakugaku Zasshi.*, **2007**, *127*, 1275-9.
- [720] Abe, F.; Iwase, Y.; Yamauchi, T.; Yahara, S.; Nohara, T. Flavonol sinapoyl glycosides from leaves of *Thevetia peruviana*. *Phytochemistry*, **1995**, *40*, 577-81.
- [721] Luo, Y.; Deng, Y.; Chen, B.; Ding, L.; Wu, F.E. A new amide from *Thyrocarpus glochidiatus*. *Nat. Prod. Res.*, **2006**, *20*, 1063-6.
- [722] Matsuda, H.; Ninomiya, K.; Shimoda, H.; Yoshikawa, M. Hepatoprotective principles from the flowers of *Tilia argentea* (linden): structure requirements of tiliroside and mechanisms of action. *Bioorg. Med. Chem.*, **2002**, *10*, 707-12.
- [723] Viola, H.; Wolfman, C.; Levi, d.S.; Wasowski, C.; Pena, C.; Medina, J.H.; Paladini, A.C. Isolation of pharmacologically active benzodiazepine receptor ligands from *Tilia tomentosa* (Tiliaceae). *J. Ethnopharmacol.*, **1994**, *44*, 47-53.
- [724] Su, L.; Feng, S.G.; Qiao, L.; Zhou, Y.Z.; Yang, R.P.; Pei, Y.H. Two new steroidal saponins from *Tribulus terrestris*. *J. Asian Nat. Prod. Res.*, **2009**, *11*, 38-43.
- [725] Sharaf, M. Chemical constituents from the seeds of *Trifolium alexandrinum*. *Nat. Prod. Res.*, **2008**, *22*, 1620-3.
- [726] Janda, B.; Stochmal, A.; Montoro, P.; Piacente, S.; Oleszek, W. Phenolics in aerial parts of Persian clover *Trifolium resupinatum*. *Nat. Prod. Commun.*, **2009**, *4*, 1661-4.
- [727] Han, Y.; Nishibe, S.; Noguchi, Y.; Jin, Z. Flavonol glycosides from the stems of *Trigonella foenum-graecum*. *Phytochemistry*, **2001**, *58*, 577-80.
- [728] Budzianowski, J.; Korzeniowska, K.; Chmara, E.; Mrozikiewicz, A. Microvascular protective activity of flavonoid glucuronides fraction from *Tulipa gesneriana*. *Phytother. Res.*, **1999**, *13*, 166-8.
- [729] Dini, A.; Rastrelli, L.; Saturnino, P.; Schettino, O. Minor components in food plants--Note I. Flavonol glycosides from *Ullucus tuberosus*. *Boll. Soc. Ital. Biol. Sper.*, **1991**, *67*, 1053-8.
- [730] Liu, A.; Zou, Z.M.; Xu, L.Z.; Yang, S.L. A new cerebroside from *Uvaria tonkinensis* var. *subglabra*. *J. Asian Nat. Prod. Res.*, **2005**, *7*, 861-5.
- [731] Ek, S.; Kartimo, H.; Mattila, S.; Tolonen, A. Characterization of phenolic compounds from lingonberry (*Vaccinium vitis-idaea*). *J. Agric. Food Chem.*, **2006**, *54*, 9834-42.
- [732] Majinda, R.R.; Motswaledi, M.; Waigh, R.D.; Waterman, P.G. Phenolic and antibacterial constituents of *Vahlia capensis*. *Planta Med.*, **1997**, *63*, 268-70.
- [733] Malafronte, N.; Pesca, M.S.; Bisio, A.; Morales, E.L.; De Tommasi, N. New flavonoid glycosides from *Vernonia ferruginea*. *Nat. Prod. Commun.*, **2009**, *4*, 1639-42.
- [734] Seetharaman, T.R. and Petrus, A.J. Novel acylkaempferol glycoside from the endemic species, *Vernonia travancorica* Hook. f. *J. Asian Nat. Prod. Res.*, **2004**, *6*, 295-9.
- [735] Vierstra, R.D.; John, T.R.; Poff, K.L. Kaempferol 3-O-Galactoside, 7-O-Rhamnoside is the Major Green Fluorescing Compound in the Epidermis of *Vicia faba*. *Plant Physiol.*, **1982**, *69*, 522-5.
- [736] Szostak, H. and Kowalewski, Z. The flavonoids in the leaves of *Vinca minor* L. (Apocynaceae). *Pol. J. Pharmacol. Pharm.*, **1975**, *27*, 657-63.
- [737] Vukics, V.; Ringer, T.; Kery, A.; Bonn, G.K.; Guttman, A. Analysis of heartsease (*Viola tricolor* L.) flavonoid glycosides by micro-liquid chromatography coupled to multistage mass spectrometry. *J. Chromatogr. A.*, **2008**, *1206*, 11-20.
- [738] Nguemaving, J.R.; Azebaze, A.G.; Kuete, V.; Eric Carly, N.N.; Beng, V.P.; Meyer, M.; Blond, A.; Bodo, B.; Nkengfack, A.E. Laurentixanthones A and B, antimicrobial xanthones from *Vismia laurentii*. *Phytochemistry*, **2006**, *67*, 1341-6.
- [739] Sandhu, A.K. and Gu, L. Antioxidant capacity, phenolic content, and profiling of phenolic compounds in the seeds, skin, and pulp of *Vitis rotundifolia* (Muscadine Grapes) As determined by HPLC-DAD-ESI-MS(n). *J. Agric. Food Chem.*, **2010**, *58*, 4681-92.
- [740] Castillo-Munoz, N.; Gomez-Alonso, S.; Garcia-Romero, E.; Hermosin-Gutierrez, I. Flavonol profiles of *Vitis vinifera* red grapes and their single-cultivar wines. *J. Agric. Food Chem.*, **2007**, *55*, 992-1002.
- [741] Rao, Y.K.; Fang, S.H.; Tzeng, Y.M. Inhibitory effects of the flavonoids isolated from *Waltheria indica* on the production of NO, TNF-alpha and IL-12 in activated macrophages. *Biol. Pharm. Bull.*, **2005**, *28*, 912-5.
- [742] Manguro, L.O.; Ugi, I.; Hermann, R.; Lemmen, P. Flavonol and drimane-type sesquiterpene glycosides of *Warburgia stuhlmannii* leaves. *Phytochemistry*, **2003**, *63*, 497-502.
- [743] Manguro, L.O.; Ugi, I.; Lemmen, P.; Hermann, R. Flavonol glycosides of *Warburgia ugandensis* leaves. *Phytochemistry*, **2003**, *64*, 891-6.
- [744] Niklas, K.J. and Giannasi, D.E. Flavonoids and Other Chemical Constituents of Fossil Miocene *Zelkova* (Ulmaceae). *Science*, **1977**, *196*, 877-8.
- [745] Usia, T.; Watabe, T.; Kadota, S.; Tezuka, Y. Mechanism-based inhibition of CYP3A4 by constituents of *Zingiber aromaticum*. *Biol. Pharm. Bull.*, **2005**, *28*, 495-9.
- [746] Usia, T.; Iwata, H.; Hiratsuka, A.; Watabe, T.; Kadota, S.; Tezuka, Y. Sesquiterpenes and flavonol glycosides from *Zingiber aromaticum* and their CYP3A4 and CYP2D6 inhibitory activities. *J. Nat. Prod.*, **2004**, *67*, 1079-83.
- [747] Jang, D.S.; Han, A.R.; Park, G.; Jhon, G.J.; Seo, E.K. Flavonoids and aromatic compounds from the rhizomes of *Zingiber zerumbet*. *Arch. Pharm. Res.*, **2004**, *27*, 386-9.
- [748] Chung, S.Y.; Jang, D.S.; Han, A.R.; Jang, J.O.; Kwon, Y.; Seo, E.K.; Lee, H.J. Modulation of P-glycoprotein-mediated resistance by kaempferol derivatives isolated from *Zingiber zerumbet*. *Phytother. Res.*, **2007**, *21*, 565-9.
- [749] Coelho, R.G.; Di Stasi, L.C.; Vilegas, W. Chemical constituents from the infusion of *Zollernia ilicifolia* Vog. and comparison with *Maytenus* species. *Z. Naturforsch. [C.]*, **2003**, *58*, 47-52.
- [750] Lin, R.D.; Chin, Y.P.; Hou, W.C.; Lee, M.H. The effects of antibiotics combined with natural polyphenols against clinical methicillin-resistant *Staphylococcus aureus* (MRSA). *Planta Med.*, **2008**, *74*, 840-6.
- [751] Martini, N.D.; Katerere, D.R.; Eloff, J.N. Biological activity of five antibacterial flavonoids from *Combretum erythrophyllum* (Combretaceae). *J. Ethnopharmacol.*, **2004**, *93*, 207-12.